CHRISTMAS COLORS: COLOMORPH DISTRIBUTION OF 
SPIROBRANCHUS GIGANTEUS PALLAS 1766 ON MOOREA, 
FRENCH POLYNESIA

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Abstract. Spirobranchus giganteus Pallas 1766 is an obligate associate of coral. This study focused on the distribution of five branchial crown colormorphs (Blue, Brown, Marigold, Purple, and White) on eight coral species (Acropora I, Acropora II, Porites I, Porites II, Porites III, Porites IV, Porites V, Porites VI) by quadrat sampling method. White was the most abundant colormorph, representing 24.0% of the total. Blue was the least abundant colormorph at 9.5% of the total. There were no significant differences in Shannon-Weiner Diversity Index (H’) of colormorphs between coral species. Also, relative colormorph abundance did not differ significantly between coral species or between the Front and Back positions. Only Blue and Marigold differed significantly in relative abundance between Top, Midde, and Bottom positions. Findings support a colormorph distribution of colormorphs. There are two possible explanations: 1) mortality and selection effects on distribution and 2) phenotypic plasticity, a combination of genetic and environmental factors contributing to the occurrence of certain phenotypes.

Key words: density; coral species; suspension feeding; phenotype; branchial crown; Polychaeta

INTRODUCTION

Coral reefs are complex systems that provide a multitude of habitats for associates, which use the tissue and skeleton as substrate (Frank et al. 1995). Risk et al. (2001) define “associate” as sessile invertebrates that live in or on the coral. Spirobranchus giganteus Pallas 1766 is one of the most conspicuous and colorful of coral associates.

Spirobranchus giganteus is an obligate associate of coral and found in tropical and sub tropical waters (Marsden and Meeuwig 1990). Hove (1970) divided S. giganteus into two sub-species: Spirobranchus giganteus corniculatus Grube 1862 in the Pacific and Spirobranchus giganteus giganteus Pallas 1766 in the Atlantic and Caribbean.

Spirobranchus giganteus is sequentially hermaphroditic and is a broadcast spawner (Kurpiyanova et al. 2001). The pelagic larvae stage lasts between 9 and 12 days (Marsden 1987). The adult worm can live between 10 - 35 years (Smith 1985) with a few surviving over 40 years (Nishi and Nishihira 1996, Nishi and Kikuchi 1996). The species does not burrow, instead it builds a calcareous tube on the surface and the coral colony grows around it (Smith 1984).

As active suspension feeding adults, the paired branchial crowns protrude from the margins of the tube. Each crown is composed of successively smaller tentacle whorls layered with 4 - 6 spiraling about a central stalk (Fig. 1). Tentacles consist of radioles with cilia laden pinnules. The cilia serve to whip water in between the pinnules (Strathmann et al. 1984).

Branchial crowns appear in a wide variety of colormorphs (e.g. blue, brown, marigold,
purple, and white). The downward slope and colorful appearance may be the source of its common name, “Christmas Tree worm.”

The presence of branchial crowns and tubes facilitate the flow of water and excrements about the coral surface (Mokady et al. 1998). *Spirobranchus giganteus* also protects coral against predation by irritating the underside of starfish *Acanthaster planci* Linnaeus 1758, inducing it to move (Devantier et al. 1986).

Ben-Tzvi et al. (2006) found that 1) *S. giganteus* colonized area was kept alive and continued to grow while areas adjacent to colonization died and 2) that coral tissue surrounding *S. giganteus* showed no predatory damage and the colonies recovered.

Previous distribution studies of *S. giganteus* show that distribution among coral is non-random and worm densities are higher on certain coral species (Hunte et al. 1990a, Marsden 1990, Marsden and Meeuwig 1990).

Distribution patterns of sessile invertebrates on coral may be affected by mortality rates (Connell 1985). *Spirobranchus giganteus* tends to cluster with increasing number of worms per colony (Dai and Yang 1995).

Hunte et al. (1990b) found that the more heavily colonized the coral, the larger the worm; larger worms were found to possess increased gamete production. The density of *S. giganteus* were found to increase with depth (Floros et al. 2005).

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**FIG. 1.** Picture of a Blue *S. giganteus* colormorph. a) spiraling whorl, b) Individual tentacles with radioles and cilia, c) central stalk and d) operculum.
Colormorphs of *S. giganteus* have not been extensively studied as in the case of other organisms, such as snails. Snail shell colormorphs were found to vary with abiotic environmental factors such as nutrients (Slotow and Ward 1997) and stresses, such as high temperatures (Etter 1988).

One approach to clarifying the occurrence of different *Spirobranchus giganteus* phenotypes is to investigate the distribution of branchial crown colormorphs. The primary purpose of this study is to study the distribution of five branchial crown colormorphs. This paper focuses on the abundance and diversity of colormorphs at different coral positions and species.

**METHODS**

**Site description**

The distribution of branchial crown colormorphs was studied on Moorea, French Polynesia at two different sties: Temae public beach (Temae) and near the Sheraton Hotel (Sheraton; Fig. 2). The survey was conducted during October and November 2006.

Temae beach (-17°27’56”N, -151°30’34”W) is a government protected public beach, within the reef crest. The depth varied between 1 - 3 m.

The Sheraton (-17°28’49”N, -149°50’50”W) is within the Plan de Gestion de l’Espace Maritime (PGEM) Marine Protected Area (MPA). Sampling took place between 100 - 140 m from the Sheraton Hotel and depths varied between 2 - 4 m.

**Sampling**

Quadrats were 3 x 3 m in size and marked off with weighted flags. The quadrats were placed randomly at Temae Beach by kayak, inside the reef crest and at the Sheraton Hotel site by motorboat. To standardize quadrats, the bottom-right corner was set first and both the bottom and top edges were lined up as closely parallel to the reef crest as possible.

**Coral-host**

Eight different coral species were examined. Coral colonies were included if more than half the coral was present within the quadrat. They were only recorded if live worms were present. Individual coral colonies were determined as separate if they were not visibly attached at the base.

The water temperature at the base, the largest diameter, the tallest height, and GPS location (Garmin 76Cx) of each coral were recorded. Corals were identified to genus offsite with digital photographs and a field guide (Allen and Steen 1994, Vernon 2000). Photographs were taken with a Casio X-Slim and a Casio underwater housing unit.

**Colormorph count**

Worm presence was determined by visual evidence of living *S. giganteus* inside their tube. Branchial crown colormorphs were only recorded once, only when at least one crown was exposed.

When possible, I positioned myself downcurrent to reduce turbulence. Positions on corals were standardized by three equal horizontal and two equal vertical planes (Top, Middle, Bottom; Front, Back). The Front of the...
coral was determined as the plane facing the reef crest.

**Colormorph designation**

A preliminary survey of branchial crown colormorphs at Temae Beach determined the five colormorphs to study: Blue, Brown, Marigold, Purple, and White (Fig. 3). Colormorphs were designated visually, on-site, by the majority color. Each worm was only counted once and in only one colormorph category.

Variations in colormorph patterns were great. I observed instances where colormorphs were not homogenous. Crowns occurred in various shades and tints (i.e. light purple, dark yellow, tan) and patterns (i.e. banded, striped, dotted).

**Statistical analysis**

My question was aimed toward patterns on the island as a whole, thus I did not compare sites on the island.

Surface area of coral colonies was estimated to $= 2 \pi r^2$, under the assumption that the shape of coral colonies are approximately hemispherical (Dahl 1973). The radius was an average of the height and

![Colormorph categories](image)

Fig. 3. Colormorph categories: A) Marigold, B) White C) Brown D) Blue and E) Purple.
of half the diameter.

Shannon-Weiner (Shannon and Weiner 1953) Diversity Index, $H'$, was calculated for colormorphs at several levels: individual corals; the front and back; and the top, middle, and bottom. The Shapiro-Wilk W-statistic was used to test the worm and coral surface area data for normality.

Parametric analyses were not possible as the data were non-normally distributed and could not be normalized through transformations.

Kruskal-Wallis (K-W) test was used to calculate significant differences in mean values to determine: if $H'$ of individual corals differed between species; if the relative abundances differed between positions (Front & Back; Top, Middle, & Bottom); if the relative abundance of colormorphs and abundance of worms differed between positions (Front-Back; Top-Middle-Bottom); and if the relative colormorph abundance for each corals differed between coral species.

If significant differences were found, Tukey’s test was used to detect where the differences occurred.

Spearman’s rank correlation coefficient analyses were used to determine whether surface area correlated with relative abundance of colormorphs.

Kruskal-Wallis tests, Tukey’s tests, and Spearman’s correlation analyses were performed using JMP 5.1 (SAS Institute 2003).

**RESULTS**

**Distribution of S. giganteus**

A total of 1,314 *S. giganteus* were recorded on 8 species of coral (Table 1). The distribution of worms was not normal (S-W: $P < 0.001$). White (318 individuals) was most abundant and Blue (126) was least abundant. Porites I had the highest, 26.9 (worms / m$^2$) and Acropora I had the lowest overall density, 3.62 (worms / m$^2$).

**Coral species**

There were no significant differences in relative abundances of colormorphs between coral species (K-W: $P > 0.24$). There were no significant differences in $H'$ of colormorphs between coral species ($P = 0.09$; Appendix A).

Distribution of coral surface area was not normal (S-W: $P < 0.001$). Using Spearman’s rank correlation coefficient, Brown was the only colormorph not significantly correlated with surface area (Brown $r_s=0.13$, $P = 0.460$; Table 2).

### Table 1. Summary of worms and coral surveyed: coral species abundance (n), total surface area (SA, m$^2$), and *S. giganteus* abundance

<table>
<thead>
<tr>
<th>Species</th>
<th>n</th>
<th>SA</th>
<th>White</th>
<th>Brown</th>
<th>Purple</th>
<th>Marigold</th>
<th>Blue</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acropora I</td>
<td>1</td>
<td>9.12</td>
<td>7</td>
<td>4</td>
<td>1</td>
<td>18</td>
<td>3</td>
</tr>
<tr>
<td>Acropora II</td>
<td>2</td>
<td>2.47</td>
<td>9</td>
<td>13</td>
<td>6</td>
<td>8</td>
<td>3</td>
</tr>
<tr>
<td>Porites I</td>
<td>4</td>
<td>5.17</td>
<td>38</td>
<td>28</td>
<td>32</td>
<td>28</td>
<td>13</td>
</tr>
<tr>
<td>Porites II</td>
<td>8</td>
<td>16.67</td>
<td>46</td>
<td>42</td>
<td>36</td>
<td>39</td>
<td>14</td>
</tr>
<tr>
<td>Porites III</td>
<td>5</td>
<td>7.13</td>
<td>23</td>
<td>14</td>
<td>33</td>
<td>39</td>
<td>19</td>
</tr>
<tr>
<td>Porites IV</td>
<td>6</td>
<td>17.12</td>
<td>83</td>
<td>88</td>
<td>59</td>
<td>54</td>
<td>27</td>
</tr>
<tr>
<td>Porites V</td>
<td>5</td>
<td>10.79</td>
<td>53</td>
<td>49</td>
<td>47</td>
<td>75</td>
<td>24</td>
</tr>
<tr>
<td>Porites VI</td>
<td>4</td>
<td>9.14</td>
<td>59</td>
<td>53</td>
<td>51</td>
<td>53</td>
<td>23</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td>35</td>
<td>77.61</td>
<td>318</td>
<td>291</td>
<td>265</td>
<td>314</td>
<td>126</td>
</tr>
</tbody>
</table>

...
TABLE 2. Spearman’s rank correlation between colormorph abundance and coral surface area, Rho and probability values (P).

<table>
<thead>
<tr>
<th>Colormorph</th>
<th>Rho</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blue</td>
<td>0.3891</td>
<td>0.021</td>
</tr>
<tr>
<td>Brown</td>
<td>0.1292</td>
<td>0.460</td>
</tr>
<tr>
<td>Marigold</td>
<td>0.4257</td>
<td>0.012</td>
</tr>
<tr>
<td>Purple</td>
<td>0.4144</td>
<td>0.013</td>
</tr>
<tr>
<td>White</td>
<td>0.5485</td>
<td>0.001</td>
</tr>
</tbody>
</table>

Front-Back Position

The relative worm abundance differed significantly between the front and back positions (K-W: $P < 0.001$; Table 3). The front was colonized more ($\bar{x} = 0.59$) than the Back ($\bar{x} = 0.41$). The relative abundances of each colormorph did not differ significantly between the Front and Back (White, $P = 0.84$; Brown, $P = 0.56$; Marigold, $P = 0.84$; Purple, $P = 0.633$; Blue, $P = 0.96$).

Top-Middle-Bottom Position

Relative abundance of worms differed significantly Top to Bottom (K-W: $P < 0.001$). The Top position was colonized the most followed by the Middle and both were different from the Bottom position, the least colonized ($\bar{x} = 0.42$, $\bar{x} = 0.38$, & $\bar{x} = 0.20$, respectively).

The relative abundances of Blue and Marigold ($P = 0.002$ and $P = 0.008$ respectively) differed significantly from Top to Bottom, both colormorphs favoring the Top and Middle positions over the Bottom. The relative abundances of the other colormorphs were not significantly different at each position (Fig. 4).

DISCUSSION

In summary, this study found that there were no differences 1) in H’ between coral species and 2) in relative colormorph abundance.

The findings of this study suggest that there are no differences in colormorph diversity as well as abundance of colormorphs on coral.

Mortality and phenotypic plasticity may explain this study’s findings of colormorph distribution.

Death is a possible explanation for the apparent random distribution of colormorphs. Mortality by density-independent means, such as predation and habitat constraints, may affect distribution of marine sessile invertebrates (Connell 1985).

Like the polymorphic *Littorina sp.*, a Mangrove snail, which was found to change in colormorph frequencies over time (Hughes and Mather 1985, Johannesson and Ekendahl 2002) a similar change in distribution also studied in (*Argiope keyserlingi*), the St.

TABLE 3. Probability values ($P$) for the K-W test to determine significant differences in relative colormorph abundances between positions (mean ± SE).

<table>
<thead>
<tr>
<th>Color</th>
<th>$P$</th>
<th>Position</th>
<th>Front</th>
<th>Back</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blue</td>
<td>0.957</td>
<td>0.076 ± 0.063</td>
<td>0.112 ± 0.182</td>
<td></td>
</tr>
<tr>
<td>Brown</td>
<td>0.559</td>
<td>0.232 ± 0.162</td>
<td>0.212 ± 0.188</td>
<td></td>
</tr>
<tr>
<td>Marigold</td>
<td>0.841</td>
<td>0.196 ± 0.140</td>
<td>0.185 ± 0.136</td>
<td></td>
</tr>
<tr>
<td>Purple</td>
<td>0.633</td>
<td>0.230 ± 0.191</td>
<td>0.211 ± 0.198</td>
<td></td>
</tr>
<tr>
<td>White</td>
<td>0.837</td>
<td>0.266 ± 0.190</td>
<td>0.251 ± 0.166</td>
<td></td>
</tr>
</tbody>
</table>
Andrew’s Cross spider (Hoese et al. 2006). In both cases, distributions of colormorphs were attributed to predation.  

*S. giganteus* predation has not been well documented. On a few occasions, I did observe damaged branchial crowns as well as completely missing or partially broken opercula. This could indicate possible attempts at predation by.

The abiotic aspects of the environment may also have an effect on colormorph distribution. Sessile invertebrates were found to influence the flow of water around coral surface areas, thus affecting nutrition available to the worms (Ben-Tzvi et al. 2006).

This may lead to the accessibility of certain nutrients, such as in the case of a desert landsnail (*Trochoidea simulata*). Slotow and Ward (1997) found that CaCO$_3$ abundance was a predictor to the frequency of shell colormorphs in *T. simulata*.

Environmental stress is also a potential factor influencing polymorphism. In the case of *Nucella lapillus*, an intertidal snail, high energy wave action and temperature was related to increased polymorphism, where as sheltered snails exhibited a higher frequency of white shells (Etter 1988).

The depths of the coral colonies in both Temae and Sheraton were relatively low in comparison to some worms found in excess of 18 m (Floros et al. 2005). The close proximity to the surface of the water may influence the fauna composition as well as subject coral and worms to abiotic factors such as sun light, current speeds, and current direction.

The consideration of both environmental and genetic factors may also be a possible explanation for. Hadfield et al. (2006) found that phenotypes were poor proxies for insight in genetic patterns for color of *Parus caeruleus* (Tit bird) but found presence of common environmental factors to be important.

Phenotypic plasticity may help explain the dichotomy between the environment and genetics. Phenotypic plasticity is the consideration of both nature (genetics) and nurture (environment) to explain variations in phenotype (Pigliucci 2001).

In the example of *Daphnia magna*, depending on the abundance of food, *D. magna* produced varying clutch sizes, responding to environmental stresses. *D. magna* would produce larger clutch sizes when food was scarce and smaller clutch sizes for when food was abundant (Ebert et al. 1993).

As it relates to *S. giganteus*, there is the possibility that colormorphs may not be any one single factor influencing phenotype. It could be influenced by a polymorphic genotype, environmental factors, or even both.

The possibility exists that the cause of branchial crown colormorphs could be dictated by a combination of abiotic and genetic factors beyond the scope of this study.

The findings of this study calls prompts a meaningful question in regards to phenotype distributions: are colormorphs heritable?

A more proximal study could be conducted to determine spatial distribution

<table>
<thead>
<tr>
<th>Colormorph</th>
<th>$P$</th>
<th>Top</th>
<th>Middle</th>
<th>Bottom</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blue</td>
<td>0.002</td>
<td>0.086 ± 0.101</td>
<td>0.101 ± 0.091</td>
<td>0.037 ± 0.076</td>
</tr>
<tr>
<td>Brown</td>
<td>0.879</td>
<td>0.218 ± 0.171</td>
<td>0.200 ± 0.167</td>
<td>0.225 ± 0.260</td>
</tr>
<tr>
<td>Marigold</td>
<td>0.008</td>
<td>0.262 ± 0.219</td>
<td>0.199 ± 0.176</td>
<td>0.135 ± 0.176</td>
</tr>
<tr>
<td>Purple</td>
<td>0.900</td>
<td>0.212 ± 0.195</td>
<td>0.196 ± 0.154</td>
<td>0.240 ± 0.234</td>
</tr>
<tr>
<td>White</td>
<td>0.567</td>
<td>0.223 ± 0.169</td>
<td>0.276 ± 0.208</td>
<td>0.278 ± 0.262</td>
</tr>
</tbody>
</table>
patterns at the individual coral level (i.e. nearest neighbor method) done similarly with worm-density studies (Dai and Yang 1995).

The next step in uncovering the pattern of colormorph distribution would be to investigate the genetic factors. This would shed light as to the mechanisms that may be responsible for S. giganteus branchial crown colormorphs and for their distribution.

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LITERATURE CITED


**APPENDIX A**

Abundance of each coral species \((n)\), mean \(H'\) value for total coral species with standard deviation.

<table>
<thead>
<tr>
<th>Species</th>
<th>(n)</th>
<th>Mean</th>
<th>Standard deviation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acropora I</td>
<td>1</td>
<td>1.178</td>
<td>*</td>
</tr>
<tr>
<td>Acropora II</td>
<td>2</td>
<td>1.360</td>
<td>0.148</td>
</tr>
<tr>
<td>Porites I</td>
<td>4</td>
<td>1.200</td>
<td>0.407</td>
</tr>
<tr>
<td>Porites II</td>
<td>8</td>
<td>1.392</td>
<td>0.0959</td>
</tr>
<tr>
<td>Porites III</td>
<td>5</td>
<td>1.127</td>
<td>0.358</td>
</tr>
<tr>
<td>Porites IV</td>
<td>6</td>
<td>1.403</td>
<td>0.119</td>
</tr>
<tr>
<td>Porites V</td>
<td>5</td>
<td>1.497</td>
<td>0.032</td>
</tr>
<tr>
<td>Porites VI</td>
<td>4</td>
<td>1.540</td>
<td>0.019</td>
</tr>
</tbody>
</table>

*only 1 individual

**APPENDIX B**

Abundance of each coral species \((n)\), mean \(H'\) value (Mean) for total coral species with standard deviation.

<table>
<thead>
<tr>
<th>Colormorph</th>
<th>K-W</th>
<th>Tukey’s Test Results</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Blue</td>
<td>0.6657</td>
<td></td>
</tr>
<tr>
<td>Brown</td>
<td>0.2450</td>
<td></td>
</tr>
<tr>
<td>Marigold</td>
<td>0.2395</td>
<td>No Significant Differences Found</td>
</tr>
<tr>
<td>Purple</td>
<td>0.5402</td>
<td></td>
</tr>
<tr>
<td>White</td>
<td>0.7258</td>
<td></td>
</tr>
</tbody>
</table>

**APPENDIX C**

Abundance for each colormorph \((n)\) by position.

<table>
<thead>
<tr>
<th>Colormorph</th>
<th>Front</th>
<th>Back</th>
<th>Top</th>
<th>Middle</th>
<th>Bottom</th>
</tr>
</thead>
<tbody>
<tr>
<td>White</td>
<td>175</td>
<td>143</td>
<td>98</td>
<td>134</td>
<td>86</td>
</tr>
<tr>
<td>Brown</td>
<td>156</td>
<td>135</td>
<td>115</td>
<td>124</td>
<td>52</td>
</tr>
<tr>
<td>Marigold</td>
<td>148</td>
<td>117</td>
<td>111</td>
<td>108</td>
<td>46</td>
</tr>
<tr>
<td>Purple</td>
<td>211</td>
<td>103</td>
<td>108</td>
<td>133</td>
<td>73</td>
</tr>
<tr>
<td>Blue</td>
<td>64</td>
<td>62</td>
<td>41</td>
<td>69</td>
<td>16</td>
</tr>
<tr>
<td>Total</td>
<td>754</td>
<td>560</td>
<td>473</td>
<td>568</td>
<td>273</td>
</tr>
</tbody>
</table>
APPENDIX D

Site maps of Temae public beach and Sheraton Hotel with coral reef boundary and individual coral.

Sheraton Hotel

Legend

- Coral Colonies

- Coral Reef Boundary

Temae Beach

Legend

- Coral Colonies

- Coral Reef Boundary
APPENDIX E

Various colormorphs of *S. giganteus corniculatus*. 