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Kenneth Steven Dahl
(Ph.D. thesis)

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STRUCTURAL EFFECTS ON THE CIRCULAR DICHROISM
OF ETHIDIUM-NUCLEIC ACID COMPLEXES

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Ph.D. Thesis

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ABSTRACT

Binding of the frameshift mutagen ethidium bromide to dinucleoside phosphates (dimers) of different base sequences was studied by optical methods, notably UV-visible spectroscopy, circular dichroism (CD), and fluorescence detected circular dichroism (FDCD). The ethidium ion intercalated between the base pairs of the minihelix formed by the complementary dimers; the stoichiometry of the complex was 2:1 dimer:dye. Equilibrium constants for complex formation showed a general preference for dye binding to complementary sequences in the order:

\[ \text{Py}(3'-5')\text{Pu} > \text{Pu}(3'-5')\text{Pu} \equiv \text{Py}(3'-5')\text{Py} > \text{Pu}(3'-5')\text{Py} \]

where \text{Py} = a pyrimidine base and \text{Pu} = a purine base. Complexes with ribodinucleoside phosphates had larger formation constants than their deoxyribo- analogues. Above 300 nm, where only the dye absorbs, the induced CD spectra of the complexes had bands at 375 nm, 330 nm, and near 307 nm. The magnitude of the 307 nm band per bound dye depended upon the base sequence in the dimers. The CD spectra of these complexes down to 220 nm were obtained by FDCD measurements; between 220 and 300 nm both positions and magnitudes of the CD bands were sequence dependent. A study of ethidium ion binding to \text{dCA}_5\text{G} + \text{dCT}_5\text{G} tested for any site preference of dye binding and for any correspondence be-

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between the dimer:dye FDCD spectra and oligomer:dye FDCD spectra.

Ethidium ion binding and optical activity in complexes with calf thymus DNA were studied as a function of NaCl concentration. The binding constant was dependent upon salt concentration, increasing as the ionic strength decreased. The magnitude of the induced CD band at 307 nm also increased as the ionic strength decreased. Possible mechanisms for this behavior and the previously observed (Houssier et al. (1974) Biopolymers 13, 1141-1160) induced CD dependence upon the dye binding ratio were presented. The available evidence favored a mechanism which considered the effects of the complete nucleic acid/bound dye/counterion system on the optical properties of the dye.
"Dr. Hoenikker used to say that any scientist who couldn't explain to an eight-year-old what he was doing was a charlatan."

"Then I'm dumber than an eight-year-old... I don't even know what a charlatan is."

Kurt Vonnegut, Jr.

Cat's Cradle
DEDICATION

To my grandparents

Vivian and Ernest Madson
Alma and Carl Dahl
who have enriched life in many ways

To my parents

Mari and Fredrick Dahl
whose love, support, and confidence
in my ability has been of inestimable aid
ACKNOWLEDGEMENTS

In setting out to acknowledge those people who have in some way helped me during the course of this work, I find that I could cite many more than I am prepared to and so add extra pages to this already long manuscript. Failure of a name to appear here does not imply an oversight of your welcome company and assistance, but rather a desire to wrap this thing up on my part.

Foremost among the people deserving special thanks during my time in Berkeley is my research director, Nacho Tinoco, who often must have wondered just what I was doing (as did I), but was there to provide guidance and encouragement during the roughest parts. His approach to research direction encourages self-direction and the ability to learn from mistakes; this was a most satisfactory arrangement for me.

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Marc Maestre and Charlie Reich provided assistance and advice during the design and construction of the FDCD. The oft-maligned Charlie was always a firm advocate of the technique's true worth, a
position best left in the memories of those who heard him expound upon it.

Two postdocs, Frank Martin and Bruce Johnson, were a stabilizing influence in the lab. Frank, with his myriad projects, willingly collaborated in the work with dCA₅G and dCT₅G and was always ready to discuss matters. Bruce furnished many moments of unintended mirth with his knowledge of practically everything. His solilquies on slugs and "gorgeous" days are classics and his exuberant personality is sorely missed.

Steve Winkle never ceased to amaze with his fertile imagination, which often carried over to his interpretation of data. It never took more than one suggestion to pull him away from his work to hit the bars.

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Jeff Nelson was the source of needed assistance during my many struggles with perfidious computers and programming; often, if he didn't know the answer, just explaining my problem to him served to
aid in its solution. He and Kathy Morden, a constant source of good cheer, were also the most frequent recipients of my barbed comments, which were often made without thought at the spur of the moment. That I could still count on their friendship after these incidents is a tribute to their patience.

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With thanks to all these people and more, I can look back upon four-and-a-half enjoyable years here.

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Chapter I

INTRODUCTION

1. Background

One of the more fascinating areas of scientific inquiry over the past thirty-five years has been focused on the structure and function of the nucleic acids: deoxyribonucleic acid (DNA) and ribonucleic acid (RNA). After the publication of the right-handed double helix structure for DNA by Watson and Crick (1953), studies in succeeding years gradually revealed the processes by which the genetic message is preserved from generation to generation through replication of DNA and how the message is transcribed and translated via RNA intermediates into proteins (for reviews, see Watson, 1976; Kornberg, 1980).

In light of the known mutagenic and carcinogenic properties of certain molecules, among them, polycyclic and heterocyclic aromatics, studies aimed at uncovering their means of producing changes in the genetic message were undertaken. Models specifying how these molecules interacted with nucleic acids were proposed, among them, the intercalation model of Lerman (1961). As shown schematically in Figure 1.1, the aromatic dye molecules, which are similar in thickness to the base pairs of the double-stranded nucleic acid, slip into the helix between adjacent base pairs, both lengthening the helix and distorting the regularity of the backbone by unwinding the helix (Waring, 1970). Many of these same intercalators were found to be frameshifters, that is, they somehow caused either an insertion or a deletion of bases in the genome during replication. One model for this process (Streisinger et al., 1966) proposed that the intercalat-
Figure 1.1. Schematic representation of the double helix of DNA. Left: DNA in the B form with the base pairs perpendicular to the helix axis. Right: B form DNA bound with intercalating dye molecules.
ing drugs stabilized intermediate looped structures (Figure 1.2) that may form during DNA replication, resulting in a new strand with bases inserted or deleted. One widely studied frameshift mutagen is ethidium bromide (Figure 1.3). It shows frameshifting activity in the Ames test (McCann et al., 1975) and intercalates into both DNA (LePecq & Paoletti, 1967) and double-stranded RNA (Douthart et al., 1973). Among the features which make ethidium bromide popular for intercalation studies are solubility in water, a low tendency to self-aggregate (Reinhardt & Krugh, 1978), an absorption shift in the visible spectrum upon binding to nucleic acids (Waring, 1965), a marked fluorescence intensity increase upon binding (LePecq & Paoletti, 1967), and acquisition of an induced optical activity when bound in nucleic acids (Aktipis & Martz, 1970). These spectroscopic features of the dye were not only seen upon binding to polymers, but were also present when it bound to complementary nucleic acid fragments as small as two base pairs long (Krugh & Reinhardt, 1975; Krugh et al., 1975; Reinhardt & Krugh, 1978). Sobell and co-workers (Tsai et al., 1977; Jain et al., 1977) were even able to obtain refined crystal structures at atomic resolution of ethidium ion complexes with 5-iodoUpA and 5-iodoCpG.

The induced CD spectrum with the dinucleoside mono- and di-phosphates, together with nuclear magnetic resonance chemical shifts of the dye protons, led to the conclusion that an ethidium ion was intercalated between the two base pairs of the minihelix in the complexes (Krugh & Reinhardt, 1975; Krugh et al., 1975). The structures obtained from the X-ray data confirmed that the phenanthridinium ring
Figure 1.2. The Streisinger model of frameshift mutagenesis.
FRAMESHIFT MUTATIONS

**Single-strand break**

**Melting** or **Nuclease action**

**Misannealing**

**Repair**

**ADDITION**

**DELETION**

XBL 782-7420A
Figure 1.3. Structure of ethidium bromide, a frameshift mutagen.
of a dye molecule was stacked between the base pairs in each minihe-lix; in addition, a second dye was stacked outside of one base pair. In the solid state the dimer:dye stoichiometry thus was 2:2. On the other hand, all evidence pointed to a 2:1 dinucleoside phosphate:dye complex in solution as long as an excess amount of dinucleoside phosphate relative to ethidium ion was maintained (Krugh & Reinhardt, 1975).

From an optical spectroscopist's point of view, perhaps the most interesting of the above observations is the induced optical activity above 300 nm acquired by the ethidium ion (EI) when bound in nucleic acids. In Figure 1.4, characteristic circular dichroism (CD) bands at 307, 330, and 375 nm are shown for the calf thymus DNA:EI complex; an additional negative band corresponding to the visible absorption band of the dye is found at 510 nm. The magnitude of $\Delta\varepsilon$ per bound ethidium ion in DNA at 307 and 330 nm strongly depends upon the ratio of bound dye to phosphate and increases as dye bound/phosphate ($r$) increases. This is charted for the 307 nm band in Figure 1.5 with data taken from several studies of the phenomenon (Dalgleish et al., 1971; Aktipis & Kindelis, 1973). This curve was reproducible, within error, for binding to nucleic acids of different base contents (Dalgleish et al., 1971; Aktipis & Martz, 1974; Williams & Seligy, 1974) and for dye binding through a wide range of added monovalent counterion concentrations (Aktipis & Kindelis, 1973; Houssier et al., 1974). Increases in $\Delta\varepsilon_{\text{bound}}^{\lambda_{\max}}$ with increasing dye binding were also seen for ethidium ion binding to calf thymus DNA that had been denatured by heating and recooling (Aktipis et al., 1975) and for the binding of ethidium ion analogues to DNA (Kindelis & Aktipis, 1978). The behavior of the
Figure 1.4. Induced circular dichroism spectra for ethidium bound to CpG (X) and calf thymus DNA at binding ratios of 0.20 dye/phosphate (□) and 0.10 dye/phosphate (◇). The molar CD (Δε) is calculated per bound dye. The salt content of the DNA solutions is ~80 mM NaCl.
Figure 1.5. Variation of the magnitude of the induced CD per bound dye at 307 nm with the extent of ethidium binding. The molar induced CD, $\Delta \varepsilon_{\text{bound}}$, is calculated on the basis of bound dye. The extent of binding, $r$, is defined as the moles of bound dye per mole of DNA residues (phosphate). Data were taken from Dalgleish et al. (1971) and Aktipis and co-workers (1973, 1974), and represent a synthesis of results in different salt concentrations and nucleic acid sequences.
Palgeish et al., 1971
Aktipis et al., 1975, 1974
longest wavelength CD band at 510 nm was different: it remained relatively constant with increasing binding ratios (Houssier et al., 1974).

Dalgleish and co-workers (1969), in a study of the induced optical activity for aminoacridine dyes bound to DNA, proposed qualitative explanations for the variations of $\Delta e_\text{bound}^\lambda$ with binding ratio:

Two possible explanations can be advanced. (1) The variation is the result of interaction between bound ligands, which naturally increases as the number of molecules in a given interacting group increases...

(2) The progressive binding of ligand molecules alters continuously the shape of the macromolecule, so that the environment of any bound ligand is determined by the number of bound ligands in its vicinity...

Mechanisms (five in all) in terms of the electronic properties of the dye and helix for the induced CD of the aminoacridines in DNA were presented by Jackson and Mason (1971). These mechanisms, together with the general picture presented by Dalgleish et al. (1969), were applied to the problem of the induced CD of ethidium ion bound in DNA. For the band at 510 nm, the asymmetry of the intercalation site in the macromolecule alone was advanced as the reason for the induced optical activity; such an interaction would remain unchanged as the extent of dye binding increased (Aktipis & Kindelis, 1973; Houssier et al., 1974). The behavior of the near UV band at 307 nm was attributed to two distinct mechanisms based on the general presentation of Dalgleish et al. (1969). The first held that at low binding ratios the induced CD per bound dye was due to the asymmetry of the binding site and thus was low. As more dye molecules intercalated in the DNA, direct interaction between transitions on two or more adjacently bound
(assuming neighbor exclusion) dye molecules gave rise to increasing magnitudes of the CD band. These interactions could either be between different transitions on the ligands (non-degenerate excitons) or the same transitions on the ligands (degenerate excitons); the latter were given greater credence because a second, roughly equal, negative CD band at 290 nm was seen under certain conditions (Aktipis & Kindelis, 1973; Houssier et al., 1974; Williams & Seligy, 1974). A second mechanism for the 307 nm band behavior was based upon symmetry arguments and attributed the induced CD in the dye to the static asymmetric perturbing field of the rest of the complex (Lee et al., 1973). In this mechanism, increased ethidium ion intercalation would alter this perturbing field and, in this case, the change in this field would increase the induced CD magnitude per bound dye.

The leveling off of the \( \Delta c_{\text{bound}}^{307} \) vs. \( r \) curve at higher \( r \) values (\( r > 0.25 \)) was attributed to saturation of the available intercalation sites under the neighbor exclusion model (Armstrong et al., 1970; Bresloff & Crothers, 1975). In this model, intercalation of a dye molecule between two base pairs rendered the immediately adjacent sites unavailable for dye binding. Thus, the limit of dye intercalation was at \( r = 0.25 \); any binding beyond this was "outside" binding due to electrostatic attractions between the charged dye and the DNA phosphates (Waring, 1965). Since only intercalated dye molecules exhibited an induced CD, the magnitude of these bands (307 and 330 nm) leveled off near this limit and then decreased since outside binding increased the amount of bound dye, but not the amount of intercalated dye (Houssier et al., 1974; Williams & Seligy, 1974).

In a series of experiments designed to more fully characterize
the interaction between the dinucleoside phosphates and the ethidium ion, Pardi (1980) obtained equilibrium constants and thermodynamic parameters which established the formation of the 2:1 complex in an excess of the self-complementary dimers CpG and dCpG. As had been done previously, measurements of the induced CD per bound dye were made for these complexes (see Figure 1.4 for CpG complex). The striking feature of these spectra were the bands at 307 nm, which were comparable in magnitude to those for DNA that was saturated with ethidium ion \((r = 0.25)\). This observation called into question the exciton mechanism proposed to explain the behavior of this band: if the greater magnitudes were simply due to a greater likelihood of dye-dye interactions at higher binding ratios in DNA, why should there be equally large magnitudes when no second dye molecule to interact with the intercalated drug was present in the dimer complex? Further work by Pardi uncovered a possible effect of salt concentration on this band: by lowering the salt concentration at a fixed binding ratio in DNA:EI complexes, \(\Delta \varepsilon_{307}^{\text{bound}}\) increased. Quite possibly the second mechanism, which invoked the effects of the static field of the helix on the dye transitions, was responsible for the induced CD's binding dependence instead.

2. Purpose and Scope of This Study

We intend to examine more fully the possible mechanisms for the induced CD spectrum of ethidium ion bound in nucleic acids and also to account for variations in the spectrum as the extent of binding changes. We employ both old and new methods for studying this binding.

We begin by looking at the interaction of ethidium ion with di-
nucleoside phosphates (loosely referred to here as dimers) of RNA and DNA (Figure 1.6), using sequences which are both self-complementary and non-self-complementary (see Figure 1.7 for the complementary Watson-Crick base pairs). We obtain equilibrium constants and thermodynamic parameters for the formation of 2:1 complexes, and also examine their induced CD spectra. We find evidence for some sequence dependence in the binding as well as sequence variations in both the thermodynamic parameters and induced CD. These are all discussed in Chapter II.

In Chapter III we use a technique which relies on both the enhanced fluorescence and induced optical activity of ethidium ion in the dimer:dye complexes to obtain their CD spectra down to 220 nm. This technique is fluorescence detected circular dichroism (FDCD), and the construction and operation of this instrument are also outlined in this chapter. One of the discoveries of Chapter III, the apparent sequence dependence of the FDCD spectra for the dimer:dye complexes, is used in Chapter IV to attempt to discern any binding site preference for ethidium ion in a longer sequence formed by the complementary heptamers dCA_5G and dCT_5G.

The dependence of DNA:EI complexes' induced CD spectra upon counterion concentration is further investigated in Chapter V. Here we find an effect similar to that seen by Pardi (1980): \( \Delta \varepsilon_{307}^{\text{bound}} \) increases as the counterion concentration decreases, all other things being equal. We use polyelectrolyte theory (Manning, 1978) to propose an explanation for this effect. Finally, in Chapter VI, we consider mechanisms for the induced CD of ethidium ion and its dependence upon binding ratios. We propose a possible mechanism for this behav-
ior in light of our new evidence and suggest further experiments to test its validity.
Figure 1.6. Structure of a ribo-dinucleoside phosphate, CpG.
Replacement of each 2' OH group by H results in the deoxy-ribo- dinucleoside phosphate, dCpG.
Figure 1.7. Watson-Crick complementary base pairs for DNA.
Replacement of the CH$_3$ group of thymine by H results in uracil, its RNA analogue.
WATSON-CRICK BASE PAIRS

ADENINE

H-8

N

N

N

N

H-2
deoxyribose

THYMINE

CH₃

H-6

deoxyribose

GUANINE

H-8

N

N

N

N

H-5
deoxyribose

CYTOSINE

H-6

deoxyribose

XBL 801-7722
Chapter II
DINUCLEOSIDE PHOSPHATE - ETHIDIUM ION INTERACTIONS

1. Introduction

Several optical studies of the binding of ethidium ion (EI) to the dinucleoside mono- and di-phosphates have previously been done. In these studies, the sequences of the dimers were both self-complementary and non-self-complementary (Krugh & Reinhardt, 1975) and both deoxyribo- and ribo-nucleosides were utilized (Krugh et al., 1975; Reinhardt & Krugh, 1978). Until the study of Pardi (1980), no quantitative conclusions had been reached on the strength of the binding, although pyrimidine (3'-5') purine sequences were known to bind the dye more readily than purine (3'-5') pyrimidine sequences. Furthermore, the complexes' stoichiometry was 2:1 dimer:dye when an excess of the dimer(s) was present (Krugh & Reinhardt, 1975; Reinhardt & Krugh, 1978).

Pardi (1980) established the 2:1 stoichiometry of the dinucleoside phosphate:ethidium ion complex when the dye was present in an excess of the self-complementary dimer CpG and showed that aggregates of this complex did not form under his experimental conditions. Equilibrium constants for the complexes of ethidium ion with CpG, dCpG, and UpA were obtained, as well as enthalpies and entropies of formation for the first two dimers' complexes. In addition to these results, he also examined the induced circular dichroism of CpG:CpG:EI and found the magnitude of the CD band at 307 nm per bound dye ($\Delta \varepsilon_{307}^{\text{bound}}$) was as large ($-22 \text{ L/mol-cm}$) as it was for DNA saturated
with ethidium ion (Aktipis & Kindelis, 1973).

In this chapter we extend the quantitative binding study of ethidium ion with dimers to include five new base sequences, among them, non-self-complementary dimers. Our results are consistent with Pardi's conclusions and further bolster his contention that dye-dye interactions are not necessarily responsible for the increase in the 307 nm CD band as the extent of binding increases.

2. Experimental:

A) Materials

Ethidium bromide was purchased from Sigma Chemical. The ribo-dinucleoside (3'-5') phosphates: CpG, UpA, CpA, UpG, ApA, UpU, ApG, CpU, ApU, and GpC, were purchased from Sigma. The deoxyribo-dinucleoside (3'-5') phosphates dCpG and dTpA were purchased from Collaborative Research. All dimers except UpA and dTpA were used without further purification. UpA and dTpA displayed several bands under UV viewing of thin layer chromatograms developed in 70:30 v/v ethanol:1 M ammonium acetate. Each dimer was spotted on Whatmann 3M chromatography paper (previously developed in ethanol to remove any impurities) and developed in 80:20 v/v ethanol:water. The dimer bands were cut out, moistened with doubly distilled water, and eluted from the paper by analytical centrifugation. The fractions were pooled, filtered through a 0.45 μ Millipore filter, frozen, and lyophilized. This procedure was repeated with the substitution of a 50:50 v/v ethanol:water solvent for the second development. Upon completion of the process, each dimer displayed only one band on a thin layer chromatogram.

The buffer used in all cases was composed of 0.18 M NaCl, 8 mM
Na$_2$HPO$_4$, 20 mM NaH$_2$PO$_4$, and 0.1 mM Na$_2$EDTA and had a measured pH of 7.0.

B) Methods

Ethidium bromide was dissolved in doubly distilled water, followed by freezing and lyophilization. This procedure was repeated twice. Stock solutions of this material were prepared with doubly distilled water and were kept cool and in the dark. Stock solutions of the dimers were prepared by dissolving each in doubly distilled water and were kept in the refrigerator.

Concentrations were monitored optically using a Cary 118 spectrophotometer. The molar extinction coefficient of ethidium ion was taken as $\varepsilon_{480} 5600$ (Haring, 1965). The extinction coefficients for the ribo- dimers at pH 7 and 25°C were taken from Warshaw (1966); the extinction coefficients for the deoxyribo- dimers at pH 7 were taken from P-L Biochemicals Reference Guide and Price List 105 (p. 27, 1977).

The molar extinction coefficients on a per dimer basis were $\varepsilon_{255}$ 19,800 for CpG, $\varepsilon_{259}$ 24,600 for UpA, $\varepsilon_{261.5}$ 21,000 for CpA, $\varepsilon_{255}$ 20,000 for UpG, $\varepsilon_{257.5}$ 27,400 for ApA, $\varepsilon_{261}$ 19,600 for UpU, $\varepsilon_{255}$ 25,000 for ApG, $\varepsilon_{265}$ 16,200 for CpU, $\varepsilon_{260}$ 24,000 for ApU, $\varepsilon_{255.5}$ 18,000 for GpC, $\varepsilon_{254}$ 19,700 for dCpG, and $\varepsilon_{260}$ 20,800 for dTpA.

Solutions containing variable amounts of the dimers and a constant dye concentration were diluted to a fixed volume with buffer. Ethidium ion concentrations were typically around 0.04 mM, while the total dimer concentration was in excess of this by 8-fold up to 250-fold, depending upon the ease of complex formation. For complexes with the non-self-complementary dimers, roughly equal amounts (within 10%) of the two dimers were added to each solution. Complex for-
mation was monitored by measuring the shift of the band in the visible spectrum (Waring, 1965) and spectra were measured either on the Cary 118 or a Gilford 250 spectrophotometer with scanning option. Spectra were digitized and stored for later use via interfaced Pet minicomputers and software partially provided by Mr. Jeff Nelson. Temperatures of the sample cells were maintained to ± 0.4°C by an external bath (Neslab Instruments) on the Cary 118 and to ± 0.1°C by a Gilford 2527 thermoelectric temperature programmer on the Gilford 250. All spectra were run in 1 cm path length quartz micro cells (Precision Cells).

Binding studies were carried out at 0°C in all cases. Binding was also studied at 5 and 10°C for some dimers. The equilibrium constants at different temperatures were used to determine ΔH° and ΔS° for complex formation by van't Hoff plots of ln K vs. T⁻¹. Additional points were provided for some plots by performing optical melts on solutions with known dimer:dye ratios. The melting temperature of the complex, Tm, was defined as the midpoint of the transition and, at this point, half the available dye was bound in the complex and half was free in solution.

Circular dichroism (CD) spectra of the complexes were run on a Cary 60 spectropolarimeter equipped with a Cary 6001 accessory. Cell temperature was maintained at 0°C (± 0.2°C) with a thermoelectrically cooled temperature jacket (Allen et al., 1972) connected to a Hallikainen Thermotrol. Quartz micro cells of 1.0 and 0.5 cm were used. Solutions prepared for the binding studies were also used in the circular dichroism study. A baseline spectrum on an equal amount of total dimer was deducted from each spectrum. Spectra were digitized and
stored using a PDP 8/E minicomputer and the revised Super Spectrum software (Appendix B).

Mixing and instrumental errors in the determinations of the equilibrium constants and $\Delta e_{\text{bound}}$ were calculated with the propagation formulae in Bevington (1969). The estimated errors in pipetting and mixing were $\sim 10\%$. Errors in the absorbance and CD spectra arose from baseline shifts and noise during measurement. The estimated errors for the spectra were $\sim 2\%$ of the chart's full scale value during a run. All analyses are discussed further in Appendix A.

3. Results

A) Optical Titrations

The shift of the 480 nm absorption band of a fixed amount of ethidium ion mixed with successively larger amounts of dTPa is shown in Figure 2.1. This shift is analogous to that seen when the dye binds to nucleic acids and can be used to determine the binding constant for the reaction if a stoichiometry is known or assumed. The greatest difference in absorbance between the dye with no dimer present and the same amount of dye in an excess of dimer occurs at 465 nm, and data at this wavelength are used in all these calculations.

Writing the general reaction for two dimer molecules combining with one dye:

\[ \text{NP} \text{Na} + \text{NPb} + \text{EI} \leftrightarrow \text{Complex} \tag{1} \]

the equilibrium expression is

\[ K = \frac{[C_{\text{Cpl}x}]}{[C_{\text{NPa}} - kC_{\text{Cpl}x}][C_{\text{NPb}} - kC_{\text{Cpl}x}][C_{\text{EI}} - C_{\text{Cpl}x}]} \tag{2} \]
Figure 2.1. Titration of 0.043 mM ethidium ion with increasing amounts of dTpA. Cell length is 1 cm and the temperature is 0°C.
Absorbance vs Wavelength (nm)

Ethidium
- 0.043 mM

dTpA
- 2.0 mM
- 2.9 mM
- 3.9 mM
- 7.8 mM
where \( k = 1 \) if the dimers are non-self-complementary (\( N_p N_a \neq N_p N_b \)) and \( k = 2 \) if the dimers are self-complementary (\( N_p N_a = N_p N_b \)). \( C_{\text{EI}}^0 \), \( C_{\text{dimer}}^0 \), and \( C_{\text{compl}}^0 \) are the total concentrations of dimer \( a \), dimer \( b \), and ethidium ion, respectively. \( C_{\text{complex}} \) is the equilibrium concentration of the complex. Analysis of the data was performed using the method of Benesi and Hildebrand (1949). For cell lengths of 1 cm we can write

\[
C_{\text{complex}} = \frac{A - \varepsilon_f C_{\text{EI}}^0}{\varepsilon_b - \varepsilon_f} \tag{3}
\]

where \( A \) is the measured absorbance of the solution of dimer(s) plus dye, \( \varepsilon_f C_{\text{EI}}^0 \) is the measured absorbance of the dye solution alone, and \( \varepsilon_f \) and \( \varepsilon_b \) are the molar extinction coefficients of the free and complexed dye, respectively. Substituting (3) into (2) and rearranging, we obtain the form

\[
\frac{C_{\text{EI}}^0}{A - \varepsilon_f C_{\text{EI}}^0} = \frac{1}{[C_{\text{dimer}}^0 - kC_{\text{complex}}][C_{\text{dimer}}^0 - kC_{\text{complex}}][\varepsilon_b - \varepsilon_f]} K
\]

\[
+ \frac{1}{[\varepsilon_b - \varepsilon_f]} \tag{4}
\]

which, by plotting the left-hand quantity vs. \( ([C_{\text{dimer}}^0 - kC_{\text{complex}}][C_{\text{dimer}}^0 - kC_{\text{complex}}])^{-1} \) will yield both \( [\varepsilon_b - \varepsilon_f] \) and \( K \) from the slope and intercept if the data are linear. The equilibrium concentrations of the dimers were initially unknown, but as a first approximation we set \( C_{\text{complex}} \) at zero and used only the initial concentrations of each (which were larger than any amount of complex which may have formed). Arriving at \( K \) via (4), we then obtained \( C_{\text{complex}} \) via (2). Restarting the process with this value in (4), we iterated to convergence of the e-
equilibrium constant to within 1%. Convergence typically occurred within four iterations (see Appendix A for the computer program used to perform this calculation).

Benesi-Hildebrand plots for the formation of 2:1 dimer:dye complexes with ethidium ion and the dimers UpA; dTpA; CpA and UpG; ApA and UpU; ApG and CpU; and ApU are shown in Figures 2.2a - 2.2f. All plots exhibit good linear fits to the data, indicating the assumption of a 2:1 stoichiometry was valid. Table I lists the equilibrium constants and $[e_b - e_f]$ for each case, along with the results for the dimers CpG and dCpG from Pardi (1980). The data are grouped in related general sequences, i.e., all ribo-dinucleoside sequences of the pyrimidine (3'-5') purine type and so forth, to facilitate comparison of sequence similarities and differences. No equilibrium constant for GpC plus ethidium ion could be measured owing to the formation of a precipitate in the mixture, even at room temperature.

B) Optical Melts

Additional values for the equilibrium constants of complex formation were obtained by performing optical melts on solutions of known dimer:dye concentrations. Monitoring of the absorbance at 465 nm throughout the melt measured the amount of ethidium ion bound in the complex at any temperature. A typical melt is displayed in Figure 2.3. The same lower baseline was applied to all melts for a particular dimer:dye system. Melts were successfully run only with the CpA/UpG/EI system; dimer:dye ratios here ranged from 10:1 up to 90:1. For the ApA/UpU/EI and ApG/CpU/EI systems we were unable to obtain a lower baseline at similar dimer:dye ratios. The lower stability of these complexes left much of the dye unbound, even at low temperatures.
Figure 2.2a. Benesi-Hildebrand plot of UpA:UpA:EI complex. Line represents least squares fit to the data. Concentrations are 0.031 – 0.049 mM for ethidium ion and 0.59 – 4.0 mM for UpA. Temperature is 0°C.
Figure 2.2b. Benesi-Hildebrand plot of dTpa:dTpa:EI complex.

Line represents least squares fit to the data. Concentrations are 0.043 mM for ethidium ion and 1.5 - 7.8 mM for dTpa. Temperature is 0°C.
\[ \frac{\varphi[Ethidium]T}{A - A_f} \times 10^{-4} \text{ (M} \cdot \text{cm)} \]

\[ \frac{1}{[dTPA]_{eq}^2} \times 10^{-5} \text{ (M}^{-2}) \]
Figure 2.2c. Benesi-Hildebrand plot of CpA:UpG:EI complex. Lines represent least squares fit to the data. Concentrations are 0.039 mM for ethidium ion, 0.16 - 1.8 mM for CpA, and 0.15 - 1.8 mM for UpG.
Figure 2.2d. Benesi-Hildebrand plot of ApA:UpU:EI complex. Lines represent least squares fit to the data. Concentrations are 0.039 mM for ethidium ion, 1.5 - 4.9 mM for ApA, and 1.5 - 5.0 mM for UpU.
Figure 2.2e. Benesi-Hildebrand plot of ApG:CpU:EI complex. Lines represent least squares fit to the data. Concentrations are 0.042 mM for ethidium ion, 1.6 - 3.9 mM for ApG, and 1.6 - 4.0 mM for CpU.
Figure 2.2f. Benesi-Hildebrand plot of ApU:ApU:EI complex.

Line represents least squares fit to the data. Concentrations are 0.040 mM for ethidium ion and 4.4 - 11 mM for ApU. Temperature is 0°C.
TABLE I

BENESI-HILDEBRAND FITS OF ETHIDIUM ION BINDING TO MINIHELICAL SEQUENCES

<table>
<thead>
<tr>
<th>Complex</th>
<th>Temperature (°C)</th>
<th>K x 10⁻⁵ (M⁻²)</th>
<th>ε_b - ε_f (L/mol-cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CpG:CpG:EI</td>
<td>0</td>
<td>890 ± 200</td>
<td>-3400 ± 200</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>420 ± 150</td>
<td>-3200 ± 300</td>
</tr>
<tr>
<td>UpA:UpA:EI</td>
<td>0</td>
<td>7.2 ± 1.0</td>
<td>-3600 ± 200</td>
</tr>
<tr>
<td>CpA:UpG:EI</td>
<td>0</td>
<td>150 ± 20</td>
<td>-3500 ± 100</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>46 ± 7</td>
<td>-3500 ± 200</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>19 ± 5</td>
<td>-3200 ± 400</td>
</tr>
<tr>
<td>dCpG:dCpG:EI</td>
<td>0</td>
<td>65 ± 10</td>
<td>-3500 ± 200</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>15 ± 2</td>
<td>-3300 ± 100</td>
</tr>
<tr>
<td>dTpA:dTpA:EI</td>
<td>0</td>
<td>1.3 ± 0.3</td>
<td>-2900 ± 400</td>
</tr>
<tr>
<td>ApA:UpU:EI</td>
<td>0</td>
<td>3.0 ± 0.4</td>
<td>-2900 ± 100</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>1.9 ± 0.3</td>
<td>-2600 ± 200</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>1.4 ± 0.3</td>
<td>-2300 ± 200</td>
</tr>
<tr>
<td>ApG:CpU:EI</td>
<td>0</td>
<td>4.9 ± 1.2</td>
<td>-3000 ± 300</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>2.7 ± 0.6</td>
<td>-2800 ± 300</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>2.1 ± 0.6</td>
<td>-2400 ± 400</td>
</tr>
<tr>
<td>ApU:ApU:EI</td>
<td>0</td>
<td>0.5 ± 0.1</td>
<td>-2500 ± 200</td>
</tr>
</tbody>
</table>

Data from Pardi (1980).
Figure 2.3. Optical melt of CpA:UpG:EI complex monitored at 465 nm in 1 cm cell. Concentrations are 0.039 mM for ethidium ion, 3.5 mM for CpA, and 3.5 mM for UpG.
The melting temperature, $T_m$, is the temperature where one half of the total ethidium ion in solution remains complexed and the rest is free. This assumes two-state behavior for the dye during the melting process. The $T_m$ is then the midpoint of the optical melt transition. Using equation (2), we can write an expression for $K$ at this temperature:

$$K = \frac{1}{(a - k/2)(b - k/2)[C_{EI}^0]^2}$$

where $a$ is the concentration ratio of dimer $NpNa$ to total dye and $b$ is the same number for $NpNb$. Experimentally, for self-complementary dimers, $a = b$ and $k = 2$, while for non-self-complementary dimers, $a = b$, generally, and $k = 1$. Each pair of $K$ and $T_m$ values from these studies was used in the van't Hoff plots (below).

C) Thermodynamics of the Binding Reaction

Determinations of the enthalpy and entropy of the binding of ethidium ion to dimers were carried out using the equilibrium constants from both the binding studies and the optical melts. Van't Hoff plots of $\ln K$ vs. $T^{-1}$ for CpA/UpG/EI, ApA/UpU/EI, and ApG/CpU/EI are presented in Figure 2.4. $\Delta H^0$ and $\Delta S^0$ for each are listed in Table II, together with those obtained by Pardi (1980) for CpG and dCpG plus ethidium ion. $\Delta H^0$ for dCpG, -29 kcal, is comparable to that of dpCpG with ethidium ion, where $\Delta H^0 = -27$ to -30.6 kcal, depending upon the method of measurement (Davanloo & Crothers, 1976).

D) Induced CD of Dimer: Dye Complexes

Measurements of the induced circular dichroism from 370 to 290 nm for ethidium ion in all the solutions from the binding studies were
Figure 2.4. Van't Hoff plots for ethidium complexes with CpA/UpG (□), ApA/UpU (○), and ApG/CpU (X). Lines represent least squares fit to the data and estimated error associated with each point.
Ethidium with

CpA/UpG

ApG/CpU

ApA/UpU

$\ln K$

$\frac{1}{T} \times 10^3$

$K^{-1}$

XBL 814-9087
TABLE II
THERMODYNAMICS OF ETHIDIUM ION BINDING
TO MINIHELICAL SEQUENCES

<table>
<thead>
<tr>
<th>Complex</th>
<th>$\Delta H^0$ (kcal/mol)</th>
<th>$\Delta S^0$ (cal/mol)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CpG:CpG:EI&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-32</td>
<td>-84</td>
</tr>
<tr>
<td>CpA:UpG:EI</td>
<td>-32 ± 1</td>
<td>-86 ± 4</td>
</tr>
<tr>
<td>dCpG:dCpG:EI&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-29</td>
<td>-69</td>
</tr>
</tbody>
</table>

<sup>a</sup>Data from Pardi (1980).
attempted. An observable induced CD spectrum was obtained for all cases except for ApU plus dye, where no significant CD signal for the ApU/EI mixture above the ApU baseline was seen, even at the highest dimer:dye ratio (11 mM:0.040 mM). The molar CD per bound dye may be very low for this complex. For GpC plus ethidium ion we obtained the same spectrum as Krugh et al. (1975), but a precipitate was suspended in the cell. We believe the spectrum is largely due to scattering by this precipitate.

The induced CD spectra for UpA; dTpA; CpA plus UpG; ApA plus UpU; and ApG plus CpU plus ethidium ion are presented in Figure 2.5. The quantity $\Delta \varepsilon_{\text{bound}}$ is the molar CD per bound dye, i.e., per mole of complex. Values of $\Delta \varepsilon_{\text{bound}}$ were calculated using

$$\Delta \varepsilon_{\text{bound}} = \theta^0 / (32.98 [C_{\text{plx}}])$$

where $\theta^0$ is the measured ellipticity of the solution in degrees, $[C_{\text{plx}}]$ is the equilibrium concentration of the complex, and $\lambda$ is the path length in cm.

The largest induced CD band is between 300 and 310 nm and the maximum position varies with the base sequence. The wavelength maxima and $\Delta \varepsilon_{\text{bound}}$ are listed with those from ethidium ion with CpG and dCpG (Pardi, 1980) in Table III. The maximum values of $\Delta \varepsilon_{\text{bound}}$ remain quite constant throughout the range of dimer:dye ratios studied for each sequence of bases. Krugh & Reinhardt (1975), using solutions in which all ethidium ion present was bound in the complex (as judged by the shift in the visible absorption band), also obtained $\Delta \varepsilon_{\text{bound}}$ values for some of the same sequences. Their results compare favorably with ours in most cases (Table III).
Figure 2.5. Induced CD spectra per bound ethidium ion at 0°C for 2:1 dimer:dye complexes.
### TABLE III

**INDUCED CD PER BOUND ETHIDIUM ION IN MINIHELICAL COMPLEXES**

<table>
<thead>
<tr>
<th>Complex at 0°C</th>
<th>Wavelength (nm)</th>
<th>$\Delta\varepsilon_{\text{bound}}$ (L/mol·cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CpG:CpG:EI</td>
<td>307</td>
<td>22 ± 3 $^a$</td>
</tr>
<tr>
<td></td>
<td>307</td>
<td>20 $^b$</td>
</tr>
<tr>
<td>UpA:UpA:EI</td>
<td>301</td>
<td>23 ± 4</td>
</tr>
<tr>
<td>CpA:UpG:EI</td>
<td>305</td>
<td>22 ± 4</td>
</tr>
<tr>
<td></td>
<td>305</td>
<td>21.4 $^b$</td>
</tr>
<tr>
<td>dCpG:dCpG:EI</td>
<td>307</td>
<td>15 ± 3 $^a$</td>
</tr>
<tr>
<td>dTpA:dTpA:EI</td>
<td>305</td>
<td>10 ± 5</td>
</tr>
<tr>
<td>ApA:UpU:EI</td>
<td>306</td>
<td>3.7 ± 0.8</td>
</tr>
<tr>
<td></td>
<td>303</td>
<td>9.0 $^b$</td>
</tr>
<tr>
<td>ApG:CpU:EI</td>
<td>308</td>
<td>7.4 ± 0.7</td>
</tr>
</tbody>
</table>

$^a$ Data from Pardi (1980).

$^b$ Data from Krugh & Reinhardt (1975).
4. Discussion

A) Stoichiometry of the Complexes

Krugh and co-workers (Krugh & Reinhardt, 1975; Krugh et al., 1975) stated that with excess dimer to ethidium ion, any minihelical complex formed in solution was likely 2:1 dimer:dye. However, the crystalline complexes of ethidium ion with 5-iodoUpA and 5-iodoCpG were composed of two dimers and two dyes (Tsai et al., 1977; Jain et al., 1977), even though the mother liquor originally contained an excess of the dimer in each case (Krugh & Reinhardt, 1975). More recently, fluorescence lifetime measurements of ethidium ions (Reinhardt & Krugh, 1978) established the existence of only one bound species in solution with excess CpG and roughly equal bound populations in crystals with CpG, confirming the 2:1 and 2:2 stoichiometries, respectively. Pardi (1980) obtained optical and equilibrium sedimentation evidence which also established the solution stoichiometry as CpG:CpG:EI.

We assumed the 2:1 dimer:dye stoichiometry in all our analyses, primarily because we always worked with an excess of dimer in each solution. That this was indeed the stoichiometry can be substantiated with two experimental observations: 1) the linearity of the data in the Benesi-Hildebrand plots, and 2) the constancy of $\Delta E_{\text{bound}}$ for each complex through a wide range of dimer to dye concentration ratios.

Attempts to fit our data to either 1:1 or 2:2 complex stoichiometries (see Pardi, 1980, for methods) all failed to produce a better fit than for a 2:1 complex, even at lower dimer:dye concentration ratios where these other complexes would more likely form. Either of these two competing stoichiometries would also have caused significant
deviations from linearity in our Benesi-Hildebrand plots (Figures 2.2a - 2.2f), especially as the dimer concentration decreased. No such deviations occurred in our data.

The occurrence of either alternate stoichiometry (1:1 or 2:2) would also have caused decreases in $\Delta \varepsilon_{\text{bound}}$ at lower dimer to dye concentration ratios. A qualitative reason for this can be advanced. First, any bound ethidium ion, no matter what the stoichiometry, would have displayed a red-shifted visible absorbance band. Such shifts were seen by Pardi (1980) in binding studies with single non-self-complementary dimers where no double-stranded minihelices formed. LePecq and Paoletti (1967) also saw this shift when ethidium ion bound via electrostatic attraction to the polyanion polyvinyl sulfate. Second, any dye bound, but not intercalated, would not exhibit an induced CD spectrum. Pardi (1980) observed this for the 1:1 complexes, and any outside stacking of dye on a 2:1 complex would presumably contribute little to the CD also. In combination, both effects would cause $\Delta \varepsilon_{\text{bound}}$ to decrease at lower dimer to dye concentration ratios; the absence of such decreases in our data rules out these competing stoichiometries.

B) Sequence Preferences in the Complexes

Previous studies of ethidium ion binding to dimers (Krugh & Reinhardt, 1975; Krugh et al., 1975; Reinhardt & Krugh, 1978; Lee & Tinoco, 1978) emphasized its relative preference for binding to pyrimidine (3'-5') purine sequences. We can explore this observation on a quantitative basis with this work and Pardi's (1980).

The grouping of complexes in Table I reveals a general sequence dependence for ethidium ion binding with complementary dimers. At 0°C
this order is \((\text{Py} = \text{pyrimidine}, \text{Pu} = \text{purine}):\)

\[
\text{Py}(3'\rightarrow 5')\text{Pu} > \text{Pu}(3'\rightarrow 5')\text{Pu} \equiv \text{Py}(3'\rightarrow 5')\text{Py} > \text{Pu}(3'\rightarrow 5')\text{Py}.
\]

This is consistent with previous results which showed the preference for \(\text{Py}(3'\rightarrow 5')\text{Pu}\) over \(\text{Pu}(3'\rightarrow 5')\text{Py}\); we place the remaining general sequence \((\text{Pu}(3'\rightarrow 5')\text{Pu} \equiv \text{Py}(3'\rightarrow 5')\text{Py})\) in the picture also. A similar order of ethidium ion binding preference has been presented for \(\text{G}\cdot\text{C}\) base pairs in deoxytetranucleotides (Kastrup et al., 1978). The equilibrium constants for the strongest (CpG) and weakest (ApU) complexes differ considerably: by a full four orders of magnitude.

The equilibrium constants we have determined are for an overall equilibrium:

\[
\text{NpN}_a + \text{NpN}_b + \text{EI} \rightleftharpoons \text{NpN}_a : \text{NpN}_b : \text{EI}.
\]  \(\text{(7)}\)

For the purposes of discussion, this overall reaction can be broken down into three reactions:

\[
\frac{\text{NpN}_a + \text{NpN}_b}{K_1} \rightleftharpoons \text{NpN}_a : \text{NpN}_b \text{ (wound)} \quad \text{(8)}
\]

\[
\frac{\text{NpN}_a : \text{NpN}_b \text{ (wound)}}{K_2} \rightleftharpoons \text{NpN}_a : \text{NpN}_b \text{ (unwound)} \quad \text{(9)}
\]

\[
\frac{\text{NpN}_a : \text{NpN}_b \text{ (unwound)} + \text{EI}}{K_3} \rightleftharpoons \text{NpN}_a : \text{NpN}_b : \text{EI} \quad \text{(10)}
\]

where our overall equilibrium constant is the product \(K_1 K_2 K_3\). The actual mechanism for complex formation depends upon sequence and is different from this scheme (Davanloo & Crothers, 1976), but individual contributions to the formation of the minihelix complexes can be considered within this framework.
Equation (8) represents the formation of minihelices in solution in the absence of the dye. Equation (9) represents the unwinding and separation of the two minihelix base pairs to accommodate a dye molecule, while equation (10) represents the intercalation of ethidium ion into the opened minihelix.

Krugh and co-workers (Young & Krugh, 1975; Krugh et al., 1976) measured the equilibrium constant $K_1$ for the formation of a double-stranded minihelix with the complementary dimers dpcpg, dpGpC, Cpg, GpC, and GpU plus ApC between 0 and 5°C. All the equilibrium constants were on the order of $10^{-1}$ M$^{-1}$ or less. Since our overall constants range from 5000 on up, the contribution of minihelix formation to the overall free energy of dimer:dye complex formation is probably small.

The energetics of the remaining two reactions have been evaluated in calculations by Ornstein and Rein (1979a, 1979b). Their calculations showed that minihelix unwinding (9) was energetically unfavorable ($\Delta H > 0$), but very specific for the general base sequence. The loss of stronger base-base stacking interactions in Pu(3'-5')Py sequences vs. Py(3'-5')Pu sequences partially accounted for the observed preferences for dye binding. Base-phosphate interactions comprised the remainder of the preference and again favored the Py(3'-5') Pu sequences; this last contribution was lessened if the phosphates were electrically neutral (Ornstein & Rein, 1979a).

Other calculations showed that the dye:minihelix interactions in reaction (10) were enthalpicly favorable ($\Delta H < 0$), but the Pu(3'-5')Py sequences were preferred over their isomeric Py(3'-5')Pu sequences; this last factor was reversed by the greater contribution to the spec-
ificity from reaction (9) (Ornstein & Rein, 1979b). Thus, the sequence specificity is provided in (9) but the driving force for binding is from (10). Important contributions in reaction (10) were provided by the dye-base interactions (overlap) and the phosphate-dye interactions (hydrogen bonding between the DNA phosphates and the amino groups of the dye). In their X-ray studies, Sobell and co-workers (Tsai et al., 1977; Jain et al., 1977) also noted the importance of dye-base overlap, while Kindelis and Aktipis (1978) found that mono-amino derivatives of the ethidium ion formed conformationally different complexes with DNA from the diamino derivatives, possibly because their hydrogen bonding properties were different.

Our equilibrium constants and other thermodynamic data are all consistent with these previous studies: Py(3'-5')Pu sequences form stronger complexes with the ethidium ion than Pu(3'-5')Py sequences. The likely reasons for this preference are those proposed in the previous studies. Complexes with complementary Pu(3'-5')Pu + Py(3'-5')Py sequences fall between these two cases. Removal of the small reaction (8) contribution to reaction (7)'s large total free energy shows that the contributions from (9) and (10) constitute the major driving force for the overall reaction in our results.

Comparisons of complex stabilities for analogous ribo- and deoxyribo- sequences are possible with these results. In each case (CpG vs. dCpG and UpA vs. dTpA), the ribo-sequence forms the stronger complex with ethidium ion. Krugh and co-workers (1975) observed the same effect with CpG and dpCpG but cautioned that direct comparison was restricted by the extra phosphate group on the deoxyribo-dimer. Both our deoxyribo-sequences have equilibrium constants smaller than their
ribo- analogues by a factor of ~10. Double-stranded RNA with G·C contents greater than 20% is more stable than DNA (Bloomfield et al., 1974); this might partially account for the greater stability of the CpG:CpG:EI complex compared to dCpG:dCpG:EI. However, the results for UpA:UpA:EI vs. dTpA:dTpA:EI run contrary to this: the deoxyribo-sequence should be more stable than the ribo-analogue. Whether all deoxyribo-dimer:dye complexes are less stable than their ribo-analologues requires further study.

The applicability of these results, especially the binding constants, to dye binding with longer oligomers and polymers is an interesting question. The binding of ethidium ion to longer sequences usually occurs with some sizable population of extant double strands. Since our results give overall constants for complex formation from two single strands plus dye, direct comparison with the equilibrium constants usually obtained with longer sequences is invalid. Still, the relative magnitudes of the constants, e.g. the ~1000-fold difference between a CpG and an ApU complex, may remain intact in longer sequences. Thus, a single macroscopic binding constant for dye binding to DNA or double-stranded RNA may mask contributions from many classes of binding sites, each with their own microscopic, but experimentally indistinguishable (at this time) binding constants.

C) Induced CD of the Complexes

The induced CD spectra of ethidium ion intercalated in complementary sequences of both deoxyribo- and ribo-dimers are all similar to those with polymers: bands are observed at 375, 330, and near 307 nm (Aktipis & Martz, 1970; Douthart et al., 1973). The similarity ends, however, when the magnitudes at 307 and 330 nm per bound dye are com-
pared for the dimer complexes and the polymers. In polymers, the magnitudes of these bands are low when few dyes are bound but they rise steadily as the extent of binding increases (Dalgleish et al., 1971; Aktipis & Kindelis, 1973; Aktipis & Martz, 1974; Houssier et al., 1974; Williams & Seligy, 1974). In the dimers, on the other hand, the magnitudes of the bands are large. In fact, the dimer $\Delta \varepsilon_{\text{bound}}$ values more closely resemble those of the polymers at higher $r$ values where virtually every intercalation site is occupied, especially the dimers with Py(3'-5')Pu sequences (Table III).

One question that immediately comes to mind is whether the CD's of the dimer complexes serve as models for longer sequences. In the only reported CD study to date of ethidium ion binding with oligomers, Kas-trup et al. (1978) examined dye binding with pdC-dG-dC-dG (2 dC-dG sites), pdC-dC-dG-dG (1 dC-dG site), pdG-dG-dC-dC, and pdG-dC-dG-dC (1 dC-dG site). For dye plus pdC-dG-dC-dG, both dC-dG sites were occupied at an added EI/strand ratio of 1, and the measured molar CD per bound dye at 305 nm was $-15$ L/mol-cm, a value identical to ours for the dCpG: dCpG:EI complex where $\Delta \varepsilon_{\text{bound}}^{305} = 15 \pm 3$ L/mol-cm. Binding stoichiometries for the dye with the other three self-complementary tetramers were less well-defined; however, at added EI/strand ratios of 1, $\Delta \varepsilon_{\text{bound}}^{305}$ for each was $-11$ L/mol-cm for pdC-dC-dG-dG, $-10$ L/mol-cm for pdG-dG-dC-dC, and $-6$ L/mol-cm for pdG-dC-dG-dC. The lower magnitudes for $\Delta \varepsilon_{\text{bound}}^{305}$ in the two sequences with one (presumably) preferred dC-dG binding site indicates that bases beyond the nearest neighbors may influence the magnitude of the CD band or that the dye molecules are bound in other sites.

A commonly accepted explanation for increasing polymer $\Delta \varepsilon_{\text{bound}}$ val-
ues near 307 nm with increasing dye binding maintains that contributions from the interaction between bound dye molecules can be added to the small CD contribution from the inherent asymmetry of the site (Houssier et al., 1974). The interaction is more likely to occur as more dye binds (and the dyes are closer to each other, on average), so the induced CD per bound dye increases. This exciton interaction between transitions on the dyes then might account for the negative lobe in the induced CD at 290 nm which is seen under certain circumstances (Aktipis & Kindelis, 1974; Aktipis & Martz, 1974; Williams & Seligy, 1974; Balcerski & Pysh, 1976).

The objections raised to this theory by Pardi (1980) centered on the fact that the magnitudes of the induced CD band around 307 nm for ethidium ion bound with CpG, dCpG, and UpA all were as large as in a DNA sample fully bound with the dye, yet only one dye molecule was present in the 2:1 complex. Furthermore, his equilibrium sedimentation study of CpG:CpG:EI showed that it was not forming aggregates, so dye-dye interactions between stacked complexes were not occurring. Clearly, if exciton interactions between intercalated dyes were not responsible for the large CD bands near 307 nm with dimers, maybe such interactions did not explain the changes in the induced CD with polymers.

For the sake of clarity, the question of the ethidium ion's induced CD in nucleic acids should be divided into two parts. First, what is the contribution to the CD from the inherent asymmetry of the binding site, and second, why does the induced CD per bound dye between 300 and 350 nm increase as more dye binds? With the dimer:dye results we can answer the first question: the apparent induced CD associated with the binding site asymmetry is large, and probably sequence depen-
dent. Naturally, there is the question of an exact correspondence between the dimer:dye complexes and complexes of the dye with longer sequences. The relative orientations of the base pairs and the dye may be less constrained in the dimer:dye complexes than in longer sequences, where bases beyond the nearest neighbors may restrict dye:base orientations in the binding site, but at present this does not seem significant because $\Delta c^\lambda_{\text{bound}}$ remains large in tetramer:dye complexes (Kastrup et al., 1978). More work with oligomers is needed to resolve this question. If the inherent asymmetry of the site contributes a large magnitude to the induced CD, previous approaches to the second question were misleading. Rather than ask what possible interactions contribute additional intensity to the low inherent CD of the near UV bands for ethidium ion in polymers as more dye binds, perhaps it is better to ask what interactions could reduce the intensity of the CD due to the site asymmetry as dye binding decreases. This is the point of view we will take into the succeeding chapters.
1. Introduction

In any solution containing more than one optically active species, the circular dichroism (CD) spectrum is sensitive to local structure around each chromophore, but it also a sum of contributions from all. Resolution of a particular chromophore's contribution from the entire spectrum may be difficult, or impossible. On the other hand, a single fluorophore mixed with other absorbing species can be readily isolated by measuring its fluorescence excitation profile. A single technique which unites the sensitivity of fluorescence with CD's conformational information is fluorescence detected circular dichroism (FDCD).

In FDCD, a sample is excited with circularly polarized light and the intensity of emission is measured as a function of the incident beam's circular polarization sense (Turner et al., 1974). Thus, FDCD is analogous to CD since both techniques provide information about the ground state of the molecule (Tinoco & Turner, 1976).

The use of fluorescence detected CD to study the complexes of ethidium ion with nucleic acids was first performed by Turner and co-workers (unpublished results) who looked at the complex formed with dCpG. The advantages of FDCD in these systems stem from the fact that the fluorescence quantum yield of the dye is enhanced up to 20-fold upon binding in the dimer:dye complex (Reinhardt & Krugh, 1978) and the dye is in a chiral environment. Thus, any FDCD signal in these
systems is almost solely from the dimer:dye complex. This is most convenient because the large excess of dimer relative to dye obscures the conventional CD spectrum of the complex below 300 nm in many cases.

The results of FDCD studies on the dimer:dye complexes studied in Chapter II are presented in this chapter. They show that the CD spectrum of the complexes below 300 nm is sensitive to the dimers' base sequence.

2. Experimental
   A) Materials

   The dinucleoside phosphates, ethidium bromide, and buffer were prepared as in Chapter II. In several cases, solutions used in the determination of binding constants were employed in the FDCD studies also. Rhodamine B was obtained from Eastman Kodak.

   α-Naphthylamine was obtained from Sigma. The material was a deep red amorphous mass, so purification by steam distillation was necessary. The steam feeder line was passed through two traps to remove any particulate matter. The slightly translucent distillate was cooled and then filtered through a 0.2 μm polycarbonate filter (Bio-Rad). Baseline solutions for FDCD were prepared from this stock by dilutions with either doubly distilled water or buffer. The pH of these solutions was ~7 as measured with indicator strips (Merck). All solutions were kept in the dark and refrigerated when not in use. α-Naphthylamine is an OSHA-regulated carcinogen and restrictions on its use are in effect. Purification of the compound was performed in a restricted access area; all glassware was cleaned separately and the washings collected in a separate waste container. The fluores-
Variable Monochromator
Circular Polarizer
Cut-Off Filter
Linear Polarizer
Photomultiplier 2
Photomultiplier 1
Cut-Off Filter
of potassium dichromate in $10^{-2}$ M KOH (Dorman et al., 1973) and an adjustable mount kindly provided by Dr. Marc Maestre. Each tube exhibited an artifact of ±1 millidegree on the 40 millidegree scale in the regions of dichromate absorbance. All fluorescence cells were also tested in this fashion; none showed artifacts above the level present in the photomultipliers.

A preamplification circuit for the photomultipliers was constructed after Turner (1978). The operational amplifier was from Union Carbide Electronics (H7020A); this model is no longer available, see Turner (1978) for an alternate. Jacks from either photomultiplier tube could be connected to the preamp depending upon the type of measurement desired. Each phototube was shielded with mu metal.

ii) Manipulation of FDCD Data

Multiple FDCD scans (5 to 9) were necessary due to the large amount of noise. Acquisition, storage, averaging, smoothing, and other manipulations of the data were performed on the PDP 8/E computer (Digital) with the revised Super Spectrum software (Appendix B) and also on the CDC 6400/6600/7600 computer system at Lawrence Berkeley Laboratory (Appendix C). Signals at dynode voltages above 950 volts were judged unreliable and ignored in subsequent analyses.

The signal from an FDCD measurement at the chart recorder on the Cary 60 is given by

$$\theta^0_F = -14.32(\Delta \varepsilon_F/\varepsilon_F - R)$$

$$R = \Delta A \left( \frac{1}{A} - \frac{2.303}{10^A - 1} \right)$$

(1)  (2)
where $\phi_F^0$ is the ellipticity in degrees, $\Delta \varepsilon_F (= \varepsilon_{FL} - \varepsilon_{FR})$ is the molar circular dichroism of the fluorophore in L/mol-cm, $\varepsilon_F$ is the molar extinction coefficient of the fluorophore (also in L/mol-cm), $\Delta A (= A_L - A_R)$ is the circular dichroism of the sample, and $A$ is its absorbance (Tinoco & Turner, 1976). The factor $\Delta \varepsilon_F/\varepsilon_F$ is referred to as the Kuhn anisotropy or the Kuhn dissymmetry factor of the fluorophore.

Equations (1) and (2) are for the general case. When only one fluorescent, optically active species is present $\Delta A/A = \Delta \varepsilon_F/\varepsilon_F$, and these reduce to the form

$$\phi_F^0 = -32.98\Delta A/(10^A - 1) \quad (3)$$

(Tinoco & Turner, 1976). For calibration of the instrument, the CD spectrum of the standard, d-10-camphorsulfonic acid (Eastman Kodak), is calculated from the FDCD and absorbance spectra by rearranging (3) to obtain

$$\phi^0 = -\phi_F^0(10^A - 1) \quad (4)$$

where the relation $\phi^0 = 32.98\Delta A$ has been used.

iii) Calibration of the Instrument

After calibration of the 6001 CD accessory according to the manufacturer's instructions, the new elevator was mounted in the Cary 60. Since the larger elevator was used for both CD and FDCD measurements, it was first calibrated in the CD mode. Prior to calibration, the Pockels cell on this elevator was aligned by minimizing the CD artifact for a solution of potassium dichromate in $10^{-2}$ M KOH.

Calibration of the new elevator for CD proceeded by connecting the leads for the CD photomultiplier tube to the preamplifier, follow-
ed by rotation of the mount into the path of the excitation beam. Using visible light (540 nm) and slits opened to 1 mm, the beam was centered on the phototube by placement of an opaque card with a small hole in the beam. When light passed by the hole reflected off the tube and was centered on the hole, the tube was properly aligned. After the mount was immobilized, calibration of the CD proceeded as for the Cary 6001 accessory. A 1 cm cell containing 4.3 mM (1 mg/ml) d-10-camphorsulfonic acid in doubly distilled water was set in the cell holder. Scans of the CD spectrum from 340 to 240 nm were made and the trimpot (R12 in Figure 5, Turner, 1978) was adjusted on the preamplifier until a maximum ellipticity of 312 millidegrees was measured (Cassim & Yang, 1969).

To calibrate the elevator for FDCD, the CD phototube leads were disconnected and the CD tube was rotated out of the excitation beam. The FDCD tube leads were connected and a Schott KV 380 interference filter was mounted in front of this phototube. To obtain a sufficiently strong signal, 21.5 mM (5 mg/ml) camphorsulfonic acid in a 1 cm fluorescence cell was used to take the FDCD spectrum. For this and all FDCD measurements, the slit multiplier was set at the maximum (10.0), as opposed to the usual setting for conventional CD (5.0) to increase the incident light intensity. The baseline solution was α-naphthylamine in buffer; this solution had an absorbance (1 cm) of 0.67 at 305 nm. To check calibration of the FDCD, the CD spectrum of the standard from FDCD and absorbance spectra via equation (4) is compared with the conventional CD spectrum in Figure 3.2; the agreement between the two curves is within 10%.

iv) Measurement of FDCD Spectra
Figure 3.2. Calibration curves of the FDCD instrument for d-10-camphorsulfonic acid (5 mg/ml) from CD measurements alone (□) and from FDCD and absorbance measurements via equation (4) (X).
Quartz fluorescence cells (Precision Cells) of 2 mm and 3 mm path lengths were utilized for all FDCD measurements of the ethidium ion:di-nucleoside phosphate complexes. A Schott KV 408 interference filter excluded all scattered or emitted light below 408 nm. In the tests for photoselection, a linear polarizer (Polaroid, HN32, 0.030") was mounted in front of the cutoff filter. No photoselection was observed for any of the complexes (see Chapter IV). Solutions of α-naphthylamine, either in water (15 OD units at 305 nm), or in buffer (1.5 OD units at 305 nm), were used for baselines depending upon the optical density of the dimer/dye solution.

Measurements of the CD and absorbance spectra at 0°C were performed on the Cary 60 with Cary 6001 CD accessory, and either the Cary 118 or Gilford 250 spectrometers, respectively. The temperature was maintained as in Chapter II. Path lengths were selected to keep the absorbance below 2 at the maximum.

v) Fluorescence Measurements

Corrected excitation profiles of ethidium ion, both alone and in the presence of CpG, were run at 0°C on a Perkin-Elmer MPF-44B fluorescence spectrophotometer. A Perkin-Elmer DCSU-2 unit was used to correct the profiles for lamp and photomultiplier characteristics. These instruments were kindly made available by Dr. Alex Glazer. The temperature was maintained with an external bath (Neslab) to within ±0.2°C. Solutions in buffer of ethidium ion (0.046 mM) alone and with CpG (0.53 mM) were scanned in a 2 mm path fluorescence cell. Slit widths were 2.4 mm (8 nm bandwidth) at the emission monochromator and 1.8 mm (6 nm bandwidth) at the excitation monochromator. Emission was monitored at 590 nm and the excitation spectra scanned from 225 to 550 nm.
Constancy of the free dye quantum yield over the profile range was checked against Rhodamine B by calculating the ratio

\[(I_R/A_R) \cdot (A_E/I_E)\]

from 225 to 550 nm (LePecq & Paoletti, 1967). \(I_R\) and \(I_E\) are the fluorescence intensities for the Rhodamine B and ethidium ion, respectively, and \(A_R\) and \(A_E\) are the absorbances of the same.

The ratio of quantum yields for equal amounts of dye bound in the complex and free in solution is

\[Q = \frac{q_b}{q_f} = \frac{I_b}{I_f} \cdot \frac{A_f}{A_b}\]

(LePecq & Paoletti, 1967), where \(I_b\) and \(I_f\) are the fluorescence intensities for bound and free ethidium ion and \(A_b\) and \(A_f\) are the absorbances of the same. The excitation profile \((I_b)\) of the CpG/EI mixture was corrected for the small amount of fluorescence due to the free dye (~5% of total dye). Concentrations of free dimer, free dye, and complex were calculated from the equilibrium constant at 0°C and the total dye and dimer concentrations. Errors in the final concentrations were estimated at 25% and were due primarily to the equilibrium constant's error.

The absorbance of the bound dye, \(A_b\), was calculated by deducting contributions of free CpG and free dye from the mixture absorbance. This provided \(A_{\text{complex}}\), from which an estimated contribution from the bases in the complex equal to \(2C_{\text{complex}^{\text{CpG}}}\), where \(e_{\text{CpG}}\) is the extinction coefficient of CpG, was subtracted to yield \(A_b\). This assumed the CpG absorbance characteristics were not changed in the complex, an as-
sumption similar to that made with the DNA:EI complexes (LePecq & Paoletti, 1967). For wavelengths greater than 310 nm, the resulting absorbance profile showed little difference from that of the mixture alone as expected, owing to the small amount of free dye and the absence of dimer absorbance in this region.

3. Results

A) Sequence Dependence of FDCD Spectra

The Kuhn anisotropy as a function of wavelength is presented for different dimer:dye complexes in Figures 3.3a through 3.3d. Above 300 nm, all the complexes exhibit a positive anisotropy. This corresponds to the induced CD band seen in this region for all the complexes except ApU:ApU:EI. The fact we were able to measure a spectrum for this last complex underscores the usefulness of FDCD for obtaining a CD spectrum where conventional CD fails. It should be noted again that the FDCD signal is solely from ethidium ion bound in the dimer:dye complex.

A change from a ribonucleoside sequence to an analogous deoxyribo-nucleoside sequence has no effect on the complexes' band positions. In the CpG vs. dCpG complexes (Figure 3.3a) this is evident. For the UpA vs. dTpA complexes (Figure 3.3b), agreement is seen below 240 nm and above 290 nm (within error). The high optical density of the dTpA mixture may be responsible for the discrepancy between the two spectra between 240 and 290 nm, where the absorbance was greatest.

Below 300 nm, where the dimers themselves possess CD spectra, distinct differences in the Kuhn anisotropy exist from complex to complex. These spectral differences may reflect the individual optical properties of the surrounding bases in the complex, different relative ori-
Figure 3.3a. Kuhn anisotropy spectra for CpG:CpG:EI complex (top) and dCpG:dCpG:EI complex (bottom). Estimated errors are represented by lines. Maximum absorbances for FDCD measurements were 2.57 in a 3 mm cell at 254 nm for CpG complex and 2.96 in a 3 mm cell at 253 nm for dCpG complex.
Figure 3.3b. Kuhn anisotropy spectra for UpA:UpA:E1 complex (top) and dTpA:dTpA:E1 complex (bottom). Estimated errors are represented by lines. Maximum absorbances for FDCD measurements were 8.07 in a 3 mm cell at 259 nm for UpA complex and 31.1 in a 2 mm cell at 261 nm for dTpA complex.
Figure 3.3c. Kuhn anisotropy spectra for CpA:UpG:EI complex (top) and ApG:CpU:EI complex (bottom). Estimated errors are represented by lines. Maximum absorbances for FDCD measurements were 5.95 in a 3 mm cell at 259 nm for CpA/UpG complex and 21.3 in a 2 mm cell at 262 nm for ApG/CpU complex.
Figure 3.3d. Kuhn anisotropy spectra for ApA:UpU:EI complex (top) and ApU:ApU:EI complex (bottom). Estimated errors are represented by lines. Maximum absorbances for FDCD measurements were 44 in a 2 mm cell at 259 nm for ApA/UpU complex and 48 in a 2 mm cell at 260 nm for ApU complex.
KUHN ANISOTROPY X 1000

ApA:UpU:Ethidium


WAVELENGTH (NM)
entations of the dye in each complex, energy transfer from nearby bases to the dye, or combinations of these effects.

B) Assignment of Spectral Bands

The Kuhn anisotropy is related to the electronic and magnetic properties of the fluorophore through the equation

$$\frac{\Delta \varepsilon_F}{\varepsilon_F} = \frac{4R_{0A}}{D_{0A}} = \frac{4\text{Im} \; \mu_{0A} \cdot m_{AO}}{\mu_{0A} \cdot m_{0A}}$$

where the transition proceeded from the ground state $0$ to the excited state $A$, $R_{0A}$ is the rotational strength of the transition, and $D_{0A}$ is its dipole strength (Tinoco & Turner, 1976). $\text{Im}$ denotes the imaginary part, and $\mu_{0A}$ and $m_{AO}$ are the electric and magnetic dipole moments.

The anisotropy should approximately be flat for each transition, providing the transition is energetically removed from other transitions, because $R_{0A}$ and $D_{0A}$ are constant (Tinoco & Turner, 1976). Overlapping transitions in the dye or with other groups will make the spectrum more complicated. The FDCD spectrum thus provides a measure of the interaction of dye transitions with neighboring transitions.

In Figure 3.4, the Kuhn anisotropy of CpG:CpG:EI is centered between absorption profiles for the complex and for ethidium ion bound in DNA (Sutherland & Sutherland, 1970), and profiles for free dimer and dye. We have obtained a profile similar to Sutherland and Sutherland's for bound dye in the CpG:CpG:EI complex (Figure 3.6). Several approximately flat regions of the anisotropy can be assigned.

The positive lobe of the Kuhn anisotropy above 300 nm can be ascribed solely to dye transitions as mentioned earlier: only the dye absorbs light in this region. The transition responsible for the nega-
Figure 3.4. Top: Extinction profiles for CpG:CpG:EI (□) and ethidium ion bound in calf thymus DNA (X). Center: Kuhn anisotropy for CpG:CpG:EI from FDCD. Bottom: Extinction profiles for CpG (△) and ethidium ion (▽).
(CpG)$_2$: Ethidium

Bound Ethidium

Sutherland and Sutherland, 1970

Ethidium

CpG

Wavelength (nm)
tive lobe of the anisotropy centered on 290 nm is the dye's strongest (Hudson & Jacobs, 1975), which is apparently reduced in intensity and red-shifted when the dye is bound (see the fluorescence excitation profiles, below).

The next anisotropy band between 240 and 280 nm is assigned to interactions between dye and base transitions. In this region of the complex absorption profile much of the energy is absorbed by the bases in the complex. Assignments of the remaining bands in the anisotropy (below 240 nm) are much more difficult to perform with the evidence at hand.

Comparisons of Kuhn anisotropies with absorption profiles cannot be done with other complexes because the high dimer/low dye concentrations and the small amounts of complex render attempts to deduce the complexes' absorption profiles statistically indefensible. However, insofar as the anisotropies of the other complexes resemble that of CpG:CpG:EI, we can make the same assignments for them. Above 300 nm the positive bands correspond to the previously obtained induced CD's and originate on the dye. Most of the spectra show at least a relative minimum near 290 nm in the anisotropy; this band also arises from a dye transition. Bands between 250 and 280 nm arise from dye-base interactions, presumably. The UpA:UpA:EI anisotropy presents a problem: it does not possess the same general features below 300 nm as the others. The applicability of the assignments to this complex is open to question.

C) CD Spectrum of Complexes from FDCD

Up to this point, analyses of the FDCD, CD, and absorbance spectra by equations (1) and (2) to obtain $\Delta \varepsilon_F / \varepsilon_F$ have been "clean": no assumptions of stoichiometries or amounts of species present have been needed.
Experimentally, either a CD spectrum could be measured or not.

One useful quantity to obtain from the Kuhn anisotropy, $\Delta \varepsilon_F / \varepsilon_F$, is the CD of the fluorophore in the complex, $\Delta \varepsilon_F$. To do this we need to derive $\varepsilon_F$ for the complex in some way. Ideally, the best method for arriving at $\varepsilon_F$ is to measure a corrected excitation profile of the bound dye, normalize this profile to the absorbance profile of the complex above 300 nm (where only the dye absorbs), and use the normalized profile extending down to 225 nm for $\varepsilon_F$.

The corrected excitation profiles for equal amounts of ethidium ion, alone and complexed with CpG, are shown in Figure 3.5. Two facts are evident: the efficiency of fluorescence is enhanced considerably upon dye binding to the dimer, and the profile for bound dye is merely red-shifted from the free dye version. The ratio of quantum yields for bound vs. free ethidium ion at different wavelengths from equation (5) are presented in Table IV. The enhancement of fluorescence upon ethidium ion intercalation into nucleic acids has been observed previously (LePecq & Paoletti, 1967; Krugh & Reinhardt, 1975; Reinhardt & Krugh, 1978; Kastrup et al., 1978). Particularly noteworthy is its relative independence of wavelength, even down in the ultraviolet where the bases absorb. This last feature was unexpected; energy transfer from the bases to the dye had been observed in DNA (LePecq & Paoletti, 1967; Sutherland & Sutherland, 1970).

In Figure 3.6, the excitation profile for CpG:CpG:EI has been normalized to the absorbance profile of the complex at 490 nm and divided by the complex concentration to obtain $\varepsilon_F$. Comparison with the profile derived from the corrected absorbance of the mixture shows each are similar. The main absorbance band at 285 nm in the free dye has shifted
Figure 3.5. Corrected excitation profiles for ethidium ion alone (X) and bound in a 2:1 complex with CpG (○).
TABLE IV

RELATIVE QUANTUM YIELDS

FOR BOUND VS. FREE ETHIDIUM ION

<table>
<thead>
<tr>
<th>Wavelength (nm)</th>
<th>$(q_b/q_f)^a$ (0°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>546</td>
<td>21 b</td>
</tr>
<tr>
<td>500</td>
<td>17</td>
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<td>450</td>
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<tr>
<td>260</td>
<td>16</td>
</tr>
<tr>
<td>240</td>
<td>28</td>
</tr>
</tbody>
</table>

a Estimated errors for values above 300 nm are ±20%; below 300 nm they are larger (±35%) due to the assumptions made for $A_b$ derivation.

b Krugh & Reinhardt (1975) reported a value of ~18 at 25°C.
Figure 3.6. Absorbance profiles for CpG:CpG:EI from corrected fluorescence excitation profile normalized at 480 nm (■) and from complex absorbance profile less the dimer's contribution (X).
CpG : CpG : Ethidium

from absorbance

from fluorescence

Wavelength (nm)
out to ~300 nm in the complex and lost ~50% of its intensity. The lack of features below 270 nm is indicative of little or no energy transfer from the bases to the dye (compare with complex's absorbance profile in Figure 3.4).

In Figure 3.7, the CD spectrum of CpG:CpG:EI from the product of $|\Delta c_F|/\epsilon_F$ and $\epsilon_F$ is presented, along with the spectrum obtained by deducting the free CpG contribution from the CpG/EI mixture CD (these two curves are shown in Figure 3.8). Coincident bands occur throughout the two CD's of the complex, but they differ in magnitudes below 300 nm, particularly below 280 nm. By assuming that the excitation profile for the CpG complex is the same for the dCpG complex, a comparison of the CD spectra from FDCD and the mixture CD (less the free dimer contribution) can be made for dCpG:dCpG:EI. This is shown in Figure 3.9. Here again, relatively good agreement on band positions is attained, but band magnitudes differ. Above 300 nm, the magnitudes of $\Delta c_{\text{bound}}$ from both methods: ~22 L/mol-cm at 307 nm for the CpG complex and ~14 L/mol-cm at 305 nm for the dCpG complex, agree well with previous results (Chapter II).

4. Discussion

A) Sequence Dependence of Complexes' CD

We have measured the Kuhn anisotropy in six of the possible ten different nearest neighbor sequences for ethidium ion binding in double-stranded RNA and also, in DNA, assuming the spectral similarities seen for the CpG and dCpG complexes are true for all analogous sequences. All the spectra are significantly different from one another, particularly below 300 nm. In the absence of complicating factors such as interactions of the dye with bases beyond the nearest neighbors, or different
Figure 3.7. CD spectrum for CpG:CpG:EI from product of $\Delta \varepsilon_F/\varepsilon_F$ and $\varepsilon_F$ (normalized fluorescence excitation profile) (□) and from mixture CD less free dimer contribution (X). Errors in band magnitudes are 30%.
The graph represents the difference in molar ellipticity ($\Delta \varepsilon = \varepsilon_L - \varepsilon_R$) as a function of wavelength (nm) for CPG/Ethidium. The data points are labeled as follows:

- Squares: CD from FD/CD
- Xs: DIFFERENCE CD

The wavelength range is from 220.00 to 370.00 nm.
Figure 3.8. CD spectra of CpG (0.41 mM) plus ethidium ion (0.043 mM) mixture at 0°C (□) and for free CpG (0.33 mM) in mixture (X). Free CpG contribution was calculated from a CD spectrum of CpG alone (0.41 mM) and concentrations derived from the equilibrium constant. Path length is 3 mm. The CD spectrum of the CpG:CpG:EI complex was obtained from these by subtraction of the free CpG component from the mixture CD and taking into account the complex concentration (0.039 mM).
Figure 3.9. CD spectrum for dCpG:dCpG:EI from product of $\Delta \varepsilon_F/\varepsilon_F$ and $\varepsilon_F$ (normalized fluorescence excitation profile) (□) and from mixture CD less free dimer contribution (X). Errors in band magnitudes are 30%.
geometries, these spectra can be utilized in attempts to distinguish preferential binding of the ethidium ion in longer sequences.

Why each complex has a different spectrum is a difficult question to answer in the absence of further experiments. In the X-ray structures of ethidium ion complexed with 5-iodoUpA and 5-iodoCpG (Tsai et al., 1977; Jain et al., 1977), the intercalated dye molecules each overlap by roughly equal amounts with the base pairs, and the substituents on the phenanthridinium ring are in the minor groove of the mini-helix. Thus, the differences in the anisotropies of the CpG and UpA complexes are more likely due to the different electronic properties of the bases, assuming the structures in the crystals apply in solution.

The effect of switching the orientation of one base pair in a sequence is observable in Figure 3.3c, where an A·U base pair is rotated about its dyad axis. The primary effect is an intensifying of bands below 300 nm for CpU:ApG:EI relative to CpA:UpG:EI; no effect is seen on the positions of the relative maxima and minima. If the dye molecules are oriented identically in each complex, then this difference is attributable to the change in the neighboring electronic environment of the dye. A second comparison can be made with UpA:UpA:EI (Figure 3.3b, top) and ApA:UpU:EI (Figure 3.3d, top), where again an A·U base pair has been rotated about its dyad axis. In this case, the changes are drastic. For self-complementary dimers, the symmetry of the mini-helix leaves the environment of the dye unaltered after a rotation about its pseudo-\( C_2 \) axis through the phenanthridinium ring. On the other hand, with non-self-complementary dimers, this rotation does change the dye's environment. Thus, even if there is a similar extent of base-dye overlap in each complex, the orientation of the dye may be different and
contribute to differences in the CD spectra also. Calculations of the CD spectra for the complexes may aid in judging the relative importance of these different effects.

B) The "Exciton" Band

One of the pieces of evidence cited in support of the dye-dye exciton theory for the induced CD of ethidium ion:nucleic acid complexes was the negative CD band near 295 nm seen at high ionic strengths (Aktipis & Kindelis, 1973; Aktipis & Martz, 1974; Balcerski & Pysh, 1976). The absence of the band at lower salt concentrations was attributed to overlap of the large positive band in the nucleic acid's CD at 275 nm; this band's intensity decreased at higher ionic strengths (Aktipis & Kindelis, 1973; Balcerski & Pysh, 1976).

We observed a negative band at 290 nm in the Kuhn anisotropy of ethidium ion in complexes with CpG, dCpG, and CpU/ApG and a relative minimum in the complexes with CpA/UpG and ApA/UpU (Figures 3.3a-d). There is only one dye to two dimers in these complexes and stacks of the 2:1 complexes do not form, so the possibility of dye-dye excitons in these complexes does not exist: such interactions are not responsible for this band in the dimer complexes. We believe this band arises from transitions on the dye, most likely the transition at 285 nm in the free dye which is red-shifted in the complex. The appearance of this CD band in 2:1 complexes suggests it might be a manifestation of the asymmetry of the binding site, both in these complexes and in polymers. If this is so, overlap of the nucleic acid's CD band at lower ionic strengths could still mask this band; it would re-appear at high ionic strengths where the 275 nm band loses intensity.

C) Is There Energy Transfer in the Complexes?
In our analysis of the CD spectrum for the CpG:CpG:EI complex we originally assumed energy transfer from the bases to the dye occurred. We believed such transfer might take place because LePecq and Paoletti (1967) observed it in DNA. They found that about half the energy absorbed by the DNA bases was transferred to the dye at phosphate/dye binding ratios near 14 and, furthermore, the transfer originated from bases not more than five base pairs away. This last result was nearly duplicated by Sutherland and Sutherland (1970), who found that transfer originated from bases 3.5 base pairs away.

The corrected fluorescence excitation profile of bound ethidium ion was enhanced above the bound dye difference spectrum in calf thymus DNA (Sutherland & Sutherland, 1970). This was an indication that energy transfer from the bases to the dye occurred. In our study of the CpG:CpG:EI complex we saw no significant difference between the corrected excitation profile and the absorbance profile for bound dye only (Figure 3.6) below 300 nm. Energy transfer in the ethidium ion complexes with dimers is seemingly nonexistent. However, in the calculation of the complex's CD spectrum by two theoretically equal methods (Figure 3.7), differences in the results above any calculated errors are evident below 300 nm. In this case, the excitation profile is missing intensity below 300 nm, suggesting that energy transfer might occur after all. This question is difficult to resolve one way or another at present.

LePecq and Paoletti (1967) measured the ratio of ethidium ion fluorescence enhancement at 260 nm to the enhancement in the visible as a function of phosphate/dye. They found the enhancement ratio was constant above P/D = 20, but that it rapidly dropped by almost a full factor of 3 near P/D = 5. Under their method of analysis, the percentage
of energy transfer from DNA to the dye would also drop an equal amount, down to 10-15% transferred. Our 2:1 dimer:dye complex is effectively at $P/D = 4$, so we estimate that the efficiency of energy transfer in our complexes is only ~10%, a figure that is apparently too low for us to measure in the fluorescence excitation profiles, if true.

Why the bases 3.5 to 5 base pairs away should be the most efficient in transferring energy to the bound ethidium ion while those closer to the dye are less efficient is an interesting question. The bases several base pairs away might be oriented in the optimum relative position to the dye for energy transfer, even though they are at greater distances than the nearer base pairs. The orientation factor may be more important than distance to the dye for the transfer. Calculations of energy transfer in DNA:EI complexes have been done (Paoletti & LePecq, 1971; Genest et al., 1974; LeBret et al., 1977), but more work in this area might prove illuminating, particularly on the question of energy transfer from the nearest and next-nearest neighbors to the dye.
Chapter IV
ETHIDIUM ION BINDING WITH dCA₅G + dCT₅G

1. Introduction

Since the ethidium ion (EI) exhibited sequence dependence in binding to complementary dinucleoside phosphates, the next logical step was to see if such specificity applied in longer sequences. Two separate investigations of binding to deoxytetranucleoside triphosphates (Patel & Canuel, 1976) and deoxytetranucleotides (Kastrup et al., 1978) have been reported. In each, the binding of dye in a Py(3'-5')Pu site (if present) predominated over binding to other site types. Ethidium ion binding in ribo- and deoxyribo- oligonucleosides with the general sequences CAₙG, CUₙG, and CTₘG where n and m are 5 or 6 is under study in this lab; a preliminary report of results can be found in Tinoco et al. (1981).

We report some CD and FDCD measurements on dCA₅G + dCT₅G + EI. In these experiments the ratio of dye to each single strand was ~1/3, so most of the minihelices contained no ethidium ion; those that did, bound only one dye. Three types of binding site exist in these minihelices: Py(3'-5')Pu (dC-dA:dT-dG, 1 site), Pu(3'-5')Pu (dA-dA:dT-dT, 4 sites), and another Pu(3'-5')Pu (dA-dG:dC-dT, 1 site). Evidence from the dimer studies indicates that dC-dA:dT-dG is the preferred binding site. We test this hypothesis in a comparison of the FDCD spectrum of the heptamer complex with the spectra obtained from dimer complexes (Chapter II).

2. Experimental

A) Materials

The deoxyribo-heptanucleoside hexaphosphates dCA₅G and dCT₅G were
prepared by the diester chemical method using triisopropylbenzenesulfonfyl chloride as the condensing agent (Khorana, 1968); these compounds were provided by Dr. Frank Martin. The molar extinction coefficients were calculated from the dinucleoside phosphates' and mononucleotides' values using the nearest neighbor approximation (Handbook of Biochemistry, Selected Data for Molecular Biology, 3rd Edition, CRC Press, p. 586). Molar extinction coefficients per strand were $\varepsilon_{260} = 79,000$ for dCA$_5$G and $\varepsilon_{260} = 58,000$ for dCT$_5$G.

Ethidium ion solutions were prepared as in Chapter II. The buffer for these experiments was composed of 0.2 M NaCl, 10 mM phosphate, pH 7.0. Two solutions of dye: alone (0.026 mM) and mixed with dCA$_5$G and dCT$_5$G (0.070 mM in each strand), were prepared in buffer for all experiments.

B) Methods

Optical melts at 260 and 280 nm were run on the Gilford 250 with 2 mm cells as described in Chapter II. Absorbance spectra in the ultraviolet (2 mm cell) and the visible (1 cm cell) were also taken on this instrument. CD and FDCD spectra were run in a 2 mm cell on the Cary 60 as in Chapter III; the cutoff filter for FDCD was a Schott KV 408. Spectra were measured at 1, 5, 15, 25, 34, and 50°C for the absorbance and CD, and at 1, 25, and 34°C for the FDCD.

Photoselection in the FDCD was tested with a linear polarizer mounted in front of the cutoff filter (Tinoco et al., 1977) in one of two positions: polarization sense vertical ($\phi = 0^\circ$) and polarization sense horizontal ($\phi = 90^\circ$). In addition, the normal FDCD spectrum with no polarizer was taken. Each averaged spectrum was analyzed via equations (1) and (2) of Chapter II with the CD and absorbance
spectra to obtain $\Delta \varepsilon_F/\varepsilon_F$ (= $g_F$). The three Kuhn anisotropies were then run in the equations of Table V to evaluate any photoselection in the heptamer minihelix:EI complex. Photoselected behavior was assumed if $\Delta \varepsilon_{33,1}/\varepsilon_F$ did not randomly scatter around a value of zero across the spectrum. The average Kuhn anisotropy from Table V, equation (1) was plotted in each case.

3. Results and Discussion

A) Melting of dCA$_5$G:dCT$_5$G:EI

The optical melt of the dCA$_5$G/dCT$_5$G/EI mixture is shown in Figure 4.1. The absorbance at 280 nm, which is near an isosbestic point for minihelix absorbance, monitors the amount of dye bound at any temperature (Dr. Frank Martin, personal communication). On the other hand, at 260 nm, the dye absorbance is low and the melting of the helix can be followed. The melting temperature, $T_m$, for the helix alone is 25 ± 1°C, while in the presence of approximately 1 ethidium ion for every 3 helices it is 27 ± 1°C. The apparent melting temperature for the dye in the helix from the absorbance at 280 nm is 34 ± 1°C. The difference in the helix melting temperatures in the mixture can be rationalized with a model in which the dye molecules bind to extant double-stranded regions rather than remain free in solution, i.e. there is migration of an ethidium ion from a complex that is melting to other remaining minihelices (Dr. Frank Martin, personal communication). This preference for dye binding to double-stranded sites was also observed in de-natured DNA (Aktipis et al., 1975). Under this model, at 27°C, 50% (ca. 0.035 mM) of the helices have reverted to single strands while ~80% (0.020 mM) of the dye molecules remain bound; at 34°C, ~20% (0.014 mM) of the minihelices remain and 50% (ca. 0.013 mM) of the dye
### Table V
EQUATIONS FOR PHOTOSELECTED FDCD

<table>
<thead>
<tr>
<th>Measured Quantities$^a$</th>
<th>Kuhn anisotropy</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phototube, polarizer orientation</td>
<td>( g_F )</td>
</tr>
</tbody>
</table>
| \( \theta = 90^\circ \), no polarizer | \[
\frac{6\Delta \epsilon_F/\epsilon_F + 2\Delta \epsilon_{33,F}/\epsilon_F}{7 - \epsilon_{33,F}/\epsilon_F}
\]
| \( \theta = 90^\circ \), \( \phi = 0^\circ \) | \[
\frac{4\Delta \epsilon_F/\epsilon_F - 2\Delta \epsilon_{33,F}/\epsilon_F}{3 + \epsilon_{33,F}/\epsilon_F}
\]
| \( \theta = 90^\circ \), \( \phi = 90^\circ \) | \[
\frac{\Delta \epsilon_F/\epsilon_F + 2\Delta \epsilon_{33,F}/\epsilon_F}{2 - \epsilon_{33,F}/\epsilon_F}
\]

where

\[
\Delta \epsilon_F/\epsilon_F = 4R/D = \text{average Kuhn anisotropy}
\]

\[
\Delta \epsilon_{33,F}/\epsilon_F = 4R_{33}/3D = \text{Kuhn anisotropy along emission transition moment}
\]

\[
\epsilon_{33,F}/\epsilon_F = D_{33}/3D = \text{absorption along emission transition moment}
\]

$^a$Tinoco et al. (1977).
### Table V

**EQUATIONS FOR PHOTOSELECTED FDCD**

Solution for Unknowns :

\[
\frac{\Delta \varepsilon}{\varepsilon_F} = \frac{(2g_\perp g_N - g_\parallel g_N - g_\parallel g_\perp)}{M} \tag{1}
\]

\[
\frac{\Delta \varepsilon_{33}}{\varepsilon_F} = \frac{(3g_\perp g_\parallel - 2g_N g_\parallel - g_N g_\perp)}{M} \tag{2}
\]

\[
\varepsilon_{33} = \frac{(-3g_\perp - 4g_\parallel + 7g_N)}{M} \tag{3}
\]

Where \( M = g_\perp - 2g_\parallel + g_N \)

- \( g_N = g_F \) at \( \theta = 90^\circ \), no polarizer
- \( g_\perp = g_F \) at \( \theta = 90^\circ \), \( \phi = 0^\circ \)
- \( g_\parallel = g_F \) at \( \theta = 90^\circ \), \( \phi = 90^\circ \)
Figure 4.1. Optical melt of dCA₅G (0.079 mM), dCT₅G (0.079 mM), and ethidium ion (0.026 mM) mixture. Measurements at 260 nm track the nucleic acid components during the melt; those at 280 nm track the dye only. Melting temperatures are 25 ± 1°C for the helix in absence of the dye, 27 ± 1°C for the helix in the presence of the dye, and 34 ± 1°C for the dye in the presence of the helix.
0.070 mM dCA₅G, 0.070 mM dCT₅G,
0.026 mM ethidium, 2 M NaCl,
0.01 M phosphate, pH 7

helix, no dye, 260 nm
helix, dye, 260 nm
helix, dye, 280 nm

RELATIVE ABSORBANCE

TEMPERATURE (°C)
molecules are free in solution. Thus, up to \(-34^\circ\text{C}\), there is an average of one ethidium ion or less bound in the minihelices, a suitable situation to distinguish preferences for different intercalation sites.

Two binding curves are presented in Figure 4.2. In Figure 4.2a, the absorbance difference at 466 nm between ethidium ion alone and in the mixture is plotted vs. temperature; in Figure 4.2b, the absorbance $\lambda_{\text{max}}$ for dye alone and in the dCA$_5$G/dCT$_5$G mixture is plotted vs. temperature. Each mixture curve approaches an asymptotic limit at low temperature, indicating almost all the dye is bound there. The maximum wavelength for fully bound dye, 518 nm, agrees with previous results of dye complexes with dimers (Krugh & Reinhardt, 1975; Krugh et al., 1975; Reinhardt & Krugh, 1978), tetramers (Kastrup et al., 1978), and DNA (Waring, 1965).

B) Circular Dichroism Studies

The CD spectrum of dCA$_5$G:dCT$_5$G as a function of temperature is displayed in Figure 4.3; in Figure 4.4, ethidium ion has been added to the solution. The appearance of the induced CD band above 300 nm at low temperatures is indicative of dye intercalation into the minihelix. The negative CD lobe between 290 and 300 nm increases by roughly a factor of two when the dye binds in the heptamer at 1°C. The spectra before and after ethidium ion binding are relatively unchanged below 290 nm.

After subtraction of the minihelix baseline contribution to the CD, $\Delta \varepsilon$ per bound dye at 310 nm is $2.1 \pm 0.5 \text{ L/mol-cm at } 1^\circ\text{C}$, assuming all dye is bound. This value is lower than in complexes with the dimers (Table III). Even with the apparent decrease in $\Delta \varepsilon_{\text{bound}}$ when ethidium ion binds in a deoxyribo- sequence (compared to its ribo- an-
Figure 4.2. Binding curves for 0.026 mM ethidium ion in the presence of dCA₅G + dCT₅G (0.070 mM in each strand). (a) Difference in absorbance at 466 nm vs. temperature for ethidium alone and in the presence of the helix. (b) Wavelength of maximum absorbance vs. temperature for ethidium alone and in the mixture.
WAVELENGTH (NM)

TEMPERATURE (°C)

\[ 466 \text{ nm} \]

\[ \Delta \text{ ABSORBANCE} \]
Figure 4.3. Circular dichroism spectra of dCA₅G + dCT₅G mixture (0.070 mM in each strand) vs. temperature. Cell length is 2 mm.
\[ d\text{CA}_5\text{G}:d\text{CT}_5\text{G} \]

Graph showing the relationship between wavelength (nm) and angle \( \theta \times 100 \) for different degrees: 1, 15, 25, 34, and 50 degrees.
Figure 4.4. Circular dichroism spectra for ethidium ion (0.026 mM) in the mixture of dCA$_5$G + dCT$_5$G (0.070 mM in each strand). Cell length is 2 mm. Spectra at 1 and 15°C have been multiplied by 50 above 300 nm.
alogue), it is difficult to make a case for preferential binding of dye in the dC-dA:dT-dG site with this result. Contributions to the dye's induced CD from bases beyond the immediate base pairs may also be significant in longer sequences so that the dimer complexes' CD's are not adequate models for comparison.

C) FDCD Spectra of dCA₅G:dCT₅G:EI

The averaged FDCD signals at two temperatures for dCA₅G:dCT₅G:EI with and without a linear polarizer are shown in Figure 4.5. The noise level at 34°C is higher than at 1°C; this is because half the dye present is free in solution at the higher temperature (see Figure 4.1).

In the FDCD measurements of dimer:ethidium complexes, the spectra taken with a polarizer were virtually coincident with the spectrum taken without it, or at least they were equally offset from it. We see similar behavior in the dCA₅G:dCT₅G:EI complex at 34°C. This is a crude indication that photoselection is not occurring in the FDCD (Turner et al., unpublished results). The true test for photoselection comes in the analysis with the three anisotropies by equation (2) in Table V. The dimer:ethidium ion complexes showed no measurable anisotropy along the emission transition moment axis ($\Delta \epsilon_{33}, F/\epsilon_F$): all values were scattered randomly about an anisotropy of zero. The same result occurs for dCA₅G:dCT₅G:EI at 34°C; we conclude that there is no photoselection in these cases. For the heptamer:dye complex at 1°C, $\Delta \epsilon_{33}, F/\epsilon_F$ was skewed toward a nonzero anisotropy, indicating some degree of photoselection. Equilibrium sedimentation studies of dCA₅G + dCT₅G at low temperatures uncovered evidence of aggregation by the minihelices (Nelson et al., 1981). The longer rotational lifetimes of the aggregates are probably responsible for the photoselection at 1°C.
Figure 4.5. Averaged FDCD signals for the ethidium ion (0.026 mM)/dCA₅G (0.070 mM)/dCT₅G (0.070 mM) mixture in 2 mm cell at 1°C (top) and 34°C (bottom). Vertical lines represent 95% confidence limits.
The average Kuhn anisotropy of dCA₅G:dCT₅G:EI calculated via equation (1) (Table V) is presented in Figure 4.6. The two spectra are similarly shaped, only the magnitudes of the bands have changed. We have not made the instrumental corrections necessary for reducing the instrumental artifacts of the FDCD spectrometer which are exacerbated in photoselecting systems (Lobenstine & Turner, 1979, 1980). Thus, the results at 1°C contain some error of unknown magnitude, but we believe the main effect is to shift the relative position of the zero line for this spectrum.

Relying on similarities between the FDCD spectra for analogous deoxyribo- and ribo- dimer complexes with ethidium ion, the FDCD spectrum for dCA₅G:dCT₅G:EI at 1°C can be compared with the spectra for each of the binding sites in Figure 4.7. Here the FDCD spectra for CpA:UpG:EI, ApA:UpU:EI, and ApC:CpU:EI represent ethidium ion binding in the dC-dA:dT-dG, dA-dA:dT-dT, and dA-dG:dC-dT sites of the heptamer, respectively. There is no close agreement between the spectrum of the heptamer and one of the "site" spectra, even for the presumably preferred dC-dA:dT-dG site, although the general patterns for all are similar. A search for the reasons behind this lack of agreement poses more questions than it answers.

On an obvious level, whether the spectra for the ribo- dimers resemble those of their deoxyribo- analogues is open to further study. Only two cases to date have been studied, CpG vs. dCpG and UpA vs. dTpA, and any similarities between the latter two were restricted to some parts of the spectrum. One way to escape this problem altogether is to take FDCD spectra with the ribo- minihelix rCA₅G:rCU₅G, plus ethidium ion.
Figure 4.6. Kuhn anisotropy of a 1:1:1 complex of $dCA_G:dCT_G$: $II$ at $1^\circ C$ (top) and $34^\circ C$ (bottom). Vertical lines represent estimated errors.
Figure 4.7. Kuhn anisotropy of 1:1:1 complex of dCA₅G:dCT₅G:EI compared with those of dimer:dye complexes representing the binding sites in the heptamer minihelix. Vertical lines represent estimated errors.
Another questionable assumption is the maintenance of relative preferences for ethidium ion binding to the different types of sites. The relative preferences for binding from dimer studies are 50:1.6:1 for the dC-dA:dT-dG, dA-dG:dC-dT, and dA-dA:dT-dT sites, respectively (ignoring the deoxyribo- vs. ribo- question). Tinoco and co-workers (1981) found that the best fits to melting curves of ethidium ion in dCA_5G:dCT_5G or rCA_5G:rCU_5G were obtained by assuming the dye bound preferentially in the Py(3'-5')Pu site relative to any of the Pu(3'-5')Pu sites. This site's binding constant was only 10 times larger than those for the other sites, a preference less than that seen in the dimers. In any case, there is some dye bound in sites other than the dC-dA:dT-dG site. Thus, the FDCO signal is partly from each of the three types of sites. For more than one fluorescent species, the Kuhn anisotropy, \( g_F \), is given by

\[
g_F = \frac{\sum \phi_i C_i \Delta \varepsilon_i}{\sum \phi_i C_i \varepsilon_i} \tag{4}
\]

where \( \Delta \varepsilon_i \) is the molar CD of species \( i \), \( \varepsilon_i \) is the molar extinction coefficient of species \( i \), \( C_i \) is the molar concentration of species \( i \), and \( \phi_i \) is the quantum yield of species \( i \) (Tinoco & Turner, 1976). Even with the assumption of equivalent quantum yields for each site, this expression is not simply a concentration-weighted average of individual site anisotropies. Resolution of contributions from individual sites is an onerous task requiring more experimental evidence than is presently available.

It is apparent from this attempt to measure preferential dye binding in oligomers by FDCO that the sequence must be carefully chosen.
There must be a strong preference for ethidium ion binding to one of the sites and/or the number of different site types must be minimized. A further factor which may complicate the picture in the current case, or any other, is possible contributions to the CD spectrum from bases further away from the dye than those in the binding site. Dye binding to sequences such as $A_n + U_n$ where $n = 2$ to $-10$ would be useful in examining the effects of bases beyond the nearest neighbors and/or other bound dye molecules on the FDCD spectrum of the complex.
Chapter V
COUNTERION EFFECTS ON DNA:ETHIDIUM ION COMPLEXES

1. Introduction

The DNA molecule, a polyanion, is sensitive to cation concentration. In general, double-stranded nucleic acids are stabilized by higher counterion concentrations (see Bloomfield et al., 1974, for review), and large conformational changes in DNA with salt/solvent conditions have been observed, as in the B form to A form transition (Ivanov et al., 1973; Ivanov et al., 1974) and the B form to C form transition (Ivanov et al., 1974). Application of polyelectrolyte theory to DNA properties in different salt concentrations has been done; one of the more successful theories is that of Manning (1978, review).

The ethidium ion, a cation, is sensitive to the salt concentration when binding to DNA (LePecq & Paoletti, 1967; Aktipis & Kindelis, 1973; Houssier et al., 1974). Furthermore, in addition to the normal intercalation binding with DNA, there is a weaker, outside binding associated with ethidium-DNA electrostatic interactions (Waring, 1965). Such binding becomes predominant at high dye-to-phosphate ratios.

The induced circular dichroism (CD) of ethidium ion (EI) bound to DNA as a function of added salt has been studied previously. Aktipis and Kindelis (1973) observed no change with ionic strength in the $\Delta \varepsilon_{\text{bound}}^{307}$ vs. $r$ curve (Figure 1.5). In this study they increased the NaCl concentration from 0 to 5 M in a basal buffer of 0.04 M tris-HCl, pH 7.9 for the DNA/dye solutions. Houssier and co-workers
(1974), working in 0.1 M and 1 mM NaCl buffers, found coincident \( \Delta \varepsilon_{308}^{\text{bound}} \) vs. \( r \) curves for the induced CD of ethidium ion; however, their binding curve was at slightly higher values of \( \Delta \varepsilon_{\text{bound}} \) relative to that of Aktipis and Kindelis (1973). More recently, Pardi (1980) compared the magnitude of the induced CD at 307 nm for \( E. \text{coli} \) DNA/ethidium ion complexes at the same binding ratio, but in different salt solutions. He found that \( \Delta \varepsilon_{\text{bound}} \) was larger for the same complex in 0.6 mM Na\(^+\) than in 50 to 500 mM Na\(^+\), suggesting that there may actually be an ionic strength dependence for the bound dye's induced CD.

In a CNDO/S study of the optical properties of ethidium ion, LeBret and Chalvet (1977) found that the shift of the dye's visible absorption band upon intercalation in DNA was almost entirely due to interaction of the dye with the phosphates of the DNA backbone. These last two works suggest that electrostatic interactions between charged groups and the bound dye may be very important in determining its optical properties.

We have run a series of experiments designed to further investigate the ionic strength dependence of the ethidium ion's induced CD when bound in DNA. We use the approach of Pardi (1980), who first dialyzed DNA samples against doubly distilled water to remove excess counterions from the DNA. This enables us to achieve lower effective Na\(^+\) concentrations when the DNA is finally diluted in the buffer. Our results qualitatively support his: at lower counterion concentrations, \( \Delta \varepsilon \) per bound dye increases relative to measurements made in higher counterion concentrations, all other things being equal. Applying Manning's theory (1978) to the DNA/EI complexes, we calculate the effective counterion concentrations in the "bound" and "free" states and
discuss a possible correlation between these electrostatic properties and the induced CD of the dye.

2. Experimental

A) Materials

Calf thymus DNA was purchased from Worthington Biochemical. A stock solution was prepared with ~200 mg of the DNA in ~200 ml of buffer (100 mM NaCl, 1 mM tris, 1 mM EDTA, pH 7.5). This solution was kept in the cold. Ethidium bromide was prepared as in Chapter II. All dye stock solutions were in doubly distilled water; their concentrations were between 0.4 and 2.5 mM.

Buffer solutions of NaCl/tris (hydroxymethyl) aminomethane were all mixed with doubly distilled water in prerinsed glassware. A high salt solution of 100 mM NaCl, 1.0 mM tris was prepared in a volumetric flask by dilution of an aliquot of 0.10 mM tris-HCl buffer, pH 7.6 (Sigma) and a weighed amount of NaCl (Sigma). The pH of the solution was 7.6 as measured with a glass electrode (Sigma) attached to a pH meter (Radiometer). A second buffer (1.0 mM NaCl, 10 μM tris) was prepared in the same fashion; the low salt solution of 0.1 mM NaCl, 1.0 μM tris was made by dilution of this stock with doubly distilled water. The pH of this solution was ~6.5.

B) Methods

i) Dialyses

Calf thymus DNA was dialyzed to remove excess counterions following the approach of Record (1975). Preliminary phenol extractions on the DNA showed no contaminating protein was present; the $A_{260}/A_{280}$ ratio for the DNA was 1.9. Aliquots (5-10 ml) of DNA stock solution were sheared by repeated passage through a 30-gauge Teflon needle, af-
ter which they were tied in dialysis tubing (VWR Scientific) cleaned by the method in Brewer et al. (1974).

All dialysis solution volumes were ~40x the original volume of DNA in each bag. Each sample was dialyzed in the cold for 24 hours against the original buffer (100 mM NaCl, 1 mM tris, 1 mM EDTA, pH 7.5), followed by another run vs. the same buffer without the EDTA. Next, four successive 24 hour dialyses vs. doubly distilled water were run to remove excess sodium ions. Samples were removed from the tubes and stored in the cold in sealed flasks.

Possible DNA denaturation during the dialysis was tested with optical melts on the Gilford 250. Samples were first degassed by bubbling with helium. In Figure 5.1 the melting curves for dialyzed and undialyzed samples at 1:5 dilution in the high salt solution are displayed; the dialyzed sample possesses a higher pre-melt baseline because of dilution of the DNA stock during dialysis. The melting temperature for each sample is around 83°C. The expected melting temperature in 200 mM Na⁺ is 86°C (Marmur & Doty, 1962), based upon the 42% G+C content of calf thymus DNA (Chan et al., 1979); since the salt content in our experiments is only 80 mM Na⁺, the actual melting temperature is slightly lower than the prediction, as expected. The breadth of the transition, defined as the inverse of the transition slope (Record, 1975), was 11°C for each.

To more accurately compare any differences between the two samples, the hypochromicity, h, defined as 100(1 - A_{260}^{25°C}/A_{260}^{90°C}), was calculated for each sample. For the undialyzed case, h_{260} = 20%, while for the dialyzed case, h_{260} = 16%. These values are both less than the commonly reported values for double-stranded nucleic acids.
Figure 5.1. Melts of calf thymus DNA stocks before (□) and after (X) dialysis to remove excess Na$^+$ ions. Aliquots of stocks were diluted to 5x their volume with 100 mM NaCl, 1.0 mM tris. Actual NaCl concentration is ~80 mM for the dialyzed sample, 100 mM for the undialyzed sample. Absorbances at 260 nm and 25°C are 0.69 for the dialyzed sample and 0.94 for the undialyzed sample; cell length is 2 mm.
Calf Thymus DNA in 100 mM NaCl, 1.0 mM TRIS dialyzed, Tm~83°C
undialyzed, Tm~83°C

TEMPERATURE (°C)
(Bloomfield et al., 1974), indicating some denaturation of the DNA sample might have occurred before the dialysis. The decrease in the hypochromicity after dialysis indicates further denaturation of the stock occurred. Nonetheless, the fact that there is a large population of double-stranded structures in the dialyzed stock, as witnessed by the melting curve, and that the dialysis procedure is reproducible allows us to prepare and use DNA samples by our method with confidence.

ii) Spectral Studies

The concentration of calf thymus DNA after dialysis was measured by diluting aliquots with 100 mM NaCl, 1 mM tris, pH 7.6 and taking the absorbance spectrum. For the DNA at 260 nm, an extinction coefficient per residue of 6600 L/mol·cm was used (Mahler et al., 1964). Concentrations of DNA in subsequent measurements were calculated from the dilution factor of the stock. DNA and dye mixtures were prepared with micropipets (Pipetman) and volumetric flasks. All volumes were checked by weighing the flask after each addition.

Scatchard analyses for the determination of ethidium ion binding constants with DNA were performed as in Waring (1965). Absorbance spectra in the visible region were taken in 1 cm quartz cells, digitized, and stored as in Chapter II. The temperature in the sample compartment was 25.0 ± 0.5°C during the measurements. CD spectra of the DNA/EI solutions at either constant dye or constant DNA concentrations were run on the Cary 60 in 1 cm or 2 cm cells. The temperature was maintained at 25.0 ± 0.2°C by thermoelectric cooling. Spectra were acquired, digitized, and stored as in Chapter II, also. The small contribution from the DNA to each spectrum was subtracted be-
fore the spectra were converted to the molar CD per bound dye basis.

(iii) Theory

The theory of polyelectrolyte solutions and counterion condensation has been actively developed over the past fifteen years. We use the work of Manning (review, 1978) in our analyses.

A linear polyelectrolyte is characterized by a regular arrangement of charged groups along the length of the molecule. In DNA these groups are the phosphates and each possesses a formal -1 charge. The strong electrostatic repulsion forces between the groups are eased by condensation of a counterion on the macromolecule; incomplete binding of counterions in the equilibrium state strikes a balance between the maximization of entropy by counterion dissociation and minimization of charge-charge repulsion energy by binding. The condensed counterions migrate freely along the macromolecule and are in equilibrium with the uncondensed counterions; no site binding is invoked.

In Manning's treatment, the linear charge density of the polyelectrolyte, \( b \), is the parameter governing the extent of counterion condensation through the dimensionless parameter \( \xi \):

\[
\xi = \frac{q^2}{\varepsilon k T b} (= 7.1/b \text{ in water at } 25^\circ C)
\]

where \( q \) is the electronic charge, \( \varepsilon \) is the bulk dielectric constant of solvent, \( k \) is Boltzmann's constant, \( T \) is the Kelvin temperature, and \( b \) has the units of Ångstroms/charge. Condensation of counterions will occur if \( \xi > 1 \), thus decreasing the linear charge density, and stop when \( \xi = 1 \). The effective charge per group is then \( (N \xi)^{-1} \) where \( N \) is the absolute value of the counterion valence. Once the polyions are stabilized via the condensation, the interactions between them
and the remaining ions are amenable to the Debye-Hückel analysis. Manning's theory thus separates the counterions in a polyelectrolyte solution into two classes: 1) those which are condensed or "bound" to the polion to ease the charge-charge repulsion and 2) those which are "free" in the remainder of the solution and interact with the partially charge-neutralized polion according to the Debye-Hückel approximation.

For native DNA in the B form, the rise along the helix axis per base pair is 3.4 Å and there are two phosphate groups with -1 charges in this distance. Therefore, b is 1.7 Å and ξ is 4.2 (Manning, 1972). Condensation of Na⁺ ions occurs in this case and the fractional charge per phosphate is reduced to ξ⁻¹ = 0.24 by the "binding" of 0.76 Na⁺ ions per phosphate.

One consequence of this theory is important for our purposes: if the density of charge on the polyelectrolyte is changed, the extent of counterion condensation will change. The ethidium ion carries a +1 charge and intercalates between base pairs of the DNA for r < 0.25 in the neighbor exclusion model. Thus, intercalation of the dye reduces the polion charge density in two ways: by a formal neutralization of one of the -1 phosphate charges and by lengthening the helix 3.4 Å for each bound dye. With this simple model of dye binding, an expression for the average axial distance per charge as a function of the binding ratio r (up to the neighbor exclusion limit of r = 0.25) is

\[ b = [1 + 3r/(1 - r)] \]  

where b is in Angstroms. This relation is derived in Appendix D.
Several other quantities of interest are also derived by Manning. The number of counterions associated per fixed charge (in a 1:1 salt like NaCl) is

$$\theta_1 = 1 - \xi^{-1}.$$  (3)

The volume surrounding the polyelectrolyte within which the counterions are considered "bound" is

$$V_p = 41.1(\xi - 1)b^3$$  (4)

where $V_p$ is in cm$^3$/mole phosphate if $b$ is in Ångstroms. The radius of a cylinder with volume $V_p$ and length $b$ aligned axially along the polyelectrolyte length is

$$a = \left(\frac{V_p}{\pi b L_{\text{Avog}}}\right)^{1/2}$$  (5)

where $a$ is in Ångstroms and $L_{\text{Avog}}$ is Avogadro's number. The local effective counterion concentration within $V_p$ is

$$C_{\text{loc}}^1 = 1000 \frac{\theta_1}{V_p}$$  (6)

where $C_{\text{loc}}^1$ is in moles/liter. These equations are all valid for native DNA and a total counterion concentration (1:1 salt) under 100 mM; we will apply them to DNA bound with intercalated dye.

3. Results

A) Stability of DNA

The stability of the dialyzed DNA in the Scatchard and CD experiments' conditions was tested via melts of representative solutions made with each of the two buffers. In Figure 5.2 the melts for 0.31 mM calf thymus DNA prepared by dilution of the dialyzed stock with
Figure 5.2. Melts for dialyzed calf thymus DNA (0.31 mM) prepared with solutions of 0.1 mM NaCl/1.0 μM tris (X) and 100 mM NaCl/1.0 mM tris (□). The T_m is 45 ± 1°C in low salt (~0.45 mM Na^+ ) and 80 ± 1°C in high salt (80 mM NaCl). Absorbances (2 mm cell) at 25°C and 250 nm are 0.41 and 0.39 for the low and high salt solutions, respectively.
either 100 mM NaCl/1.0 mM tris and 0.1 mM NaCl/1.0 μM tris are presented. The $T_m$ for the DNA in the high salt case is $80 \pm 1^\circ C$, while in the low salt case it is $45 \pm 1^\circ C$. At the temperature of the Scatchard and CD experiments, $25^\circ C$, the DNA remains double-stranded according to these results.

A point requiring clarification in this chapter is the value of the total sodium ion concentration in the DNA/dye solutions. Dialysis of the DNA vs. doubly distilled water removed a large number of sodium ions from the solution, but the Donnan effect requires some to remain with the nucleic acid above the amount needed to maintain charge neutrality. For example, Record (1975), in the dialysis of T4 DNA vs. doubly distilled water, found that $2.5 Na^+$ ions per phosphate remained in the solution after 18 hours. In a preliminary experiment, we analyzed for $Na^+$ with atomic absorption spectroscopy. Our results indicated that roughly $1.2 Na^+$ ions were present per DNA phosphate. The important point is that the total $Na^+$ concentration in solution is not that of the buffer because of these counterions associated with the DNA and also because of dilution of the buffer (80% or more of the final solution volume is from the salt solution). With the constant DNA concentration (0.31 mM) employed in the low salt CD studies, the $Na^+$ concentration is at least 0.45 mM, depending upon the reliability of the $Na^+$ analyses. In the high salt solutions the $Na^+$ concentration is 80 mM or greater and the $Na^+$ contribution from DNA is minimal.

B) Binding of Ethidium Ion to DNA

Constant amounts of ethidium ion were titrated with DNA in the two salt solutions at $25^\circ C$. Absorbance spectra at each salt level are
shown in Figures 5.3 and 5.4. The total dye concentration is near 0.03 mM and all is bound in ~0.5 mM DNA. The isosbestic points near 394 and 510 nm are indicative of two states for ethidium ion: free in solution and bound to the nucleic acid. For fully bound dye, the visible wavelength of maximum absorbance is 520 nm, a result which is in agreement with previous studies (Waring, 1965).

For ligands binding independently to equivalent sites on a macromolecule, the equilibrium between bound and free ligands is given by the Scatchard (1949) equation:

$$\frac{r}{c_f} = K_n - K_r$$

(7)

where $r$ is the ratio of bound ethidium ion per DNA phosphate, $c_f$ is the free dye concentration in moles/liter, $n$ is the total number of binding sites per DNA phosphate, and $K$ is the binding constant. The concentration of free dye is calculated from the total concentration of dye and the quantity $A_{mix} - A_{dye}$ measured at 465 nm; this last quantity gives a measure of the amount of dye bound in a two-state analysis. The DNA concentration is calculated from the dilution factor as mentioned earlier.

The Scatchard plot for calf thymus DNA/EI mixtures prepared with the 100 mM NaCl/1.0 mM tris buffer is shown in Figure 5.5. Again, because the salt solution is diluted in mixing with DNA/dye, the Na$^+$ concentration is less than 100 mM; the median value for the counter-ion concentration is 86 mM Na$^+$.

A least squares fit to the data yields an equilibrium constant of $9.7 (\pm 0.7) \times 10^5$ M$^{-1}$ and $0.20 \pm 0.02$ binding sites/phosphate. This result is in fair agreement with LePecq and Paoletti (1967), who ob-
Figure 5.3. Titration of 0.032 mM ethidium ion (D) with
dialyzed calf thymus DNA. DNA concentration as phosphate:

(☐) 0.015 mM
(X) 0.030 mM
(▽) 0.045 mM
(+) 0.076 mM
(0) 0.50 mM

Solutions were prepared with 0.1 mM NaCl, 1.0 mM tris; the
actual Na\(^+\) concentration depends upon DNA concentration.
Figure 5.4. Titration of 0.033 mM ethidium ion (□) with dialyzed calf thymus DNA. DNA concentration as phosphate:

- (◇) 0.054 mM
- (X) 0.11 mM
- (▽) 0.14 mM
- (+) 0.19 mM
- (0) 0.53 mM

Solutions were prepared with 100 mM NaCl, 1.0 mM tris; the actual Na⁺ concentration depends upon DNA concentration.
Figure 5.5. Scatchard plot for ethidium ion binding in dialyzed calf thymus DNA at 25°C. Buffer used is 100 mM NaCl/1.0 mM tris; the median Na⁺ concentration is ~86 mM for points fitted by least squares routine. Vertical lines represent estimated errors. Least squares fit to all points except for those above \( r = 0.19 \) is represented by sloping line. For this fit, \( K = 9.7 (\pm 0.7) \times 10^5 \text{ M}^{-1} \) and \( n = 0.20 \pm 0.02 \).
Calf Thymus DNA
plus Ethidium
100 mM NaCl
tained a dye binding constant with calf thymus DNA of $6.6 \times 10^5 \text{ M}^{-1}$ at $23^\circ\text{C}$ in 90 mM Na$^+$. 

In the mixture prepared with the low salt solution (0.1 mM NaCl, 1.0 μM tris), all solutions with total dye/DNA phosphate < 0.25 display coincident absorbance spectra that are characteristic of fully bound ethidium ion. The equilibrium constant in this case is too large to be measured by this method. Previous workers (LePecq & Paoletti, 1967; Aktipis & Kindelis, 1973; Houssier et al., 1974) found that the equilibrium constant for ethidium ion binding to DNA increased as the counterion concentration decreased. For our purposes, it is sufficient to say that all dye is bound to the nucleic acid at dye/phosphate ratios below 0.25 in our low salt solutions. For the CD solutions with a constant DNA concentration and variable dye concentrations in the low salt mixtures, the measured absorbance spectra all resemble that of fully bound ethidium ion.

C) Induced CD of DNA/Ethidium Ion Complexes

The induced CD spectra of ethidium ion bound to calf thymus DNA in the two different salt solutions are shown in Figure 5.6. Two features are apparent upon inspection and comparison of the spectra. The first is the relative constancy of $\Delta_{\epsilon_{\text{bound}}}^s$ at 375 nm, especially in the high salt mixtures. Comparison of spectra at the same binding ratio in the two salt solutions turns up the second feature: $\Delta_{\epsilon_{\text{bound}}}^s$ is greater in the lower salt concentration at either 307 or 330 nm relative to the high salt case.

The increase in $\Delta_{\epsilon_{\text{bound}}}^{307}$ with r, the binding ratio, is shown in Figure 5.7. The curve assembled from the previous studies of Dalgleish et al. (1971) and Aktipis and co-workers (1973, 1974), which
Figure 5.6. Induced CD spectra of DNA/EI complexes at different binding ratios and salt concentrations. Na\(^+\) concentrations are ~0.86 mM (median value, top) and ~0.45 mM (bottom).
100 mM NaCl

$x = 0.05$
$三人行 = 0.10$
$+$ $r = 0.15$
$\square r = 0.20$

0.1 mM NaCl

$x = 0.05$
$三人行 = 0.10$
$+$ $r = 0.15$
$\square r = 0.20$
Figure 5.7. Variation of the magnitude of the induced CD per bound dye at 307 nm with the extent of ethidium ion binding and counterion concentration. Spectra run in solutions prepared with 0.1 mM NaCl/1.0 μM tris (X) have actual Na⁺ concentration of ~0.45 mM. Spectra run in solutions prepared with 100 mM NaCl/1.0 mM tris (○) have median Na⁺ concentration of ~0.86 mM. Vertical lines represent estimated errors; solid line represents a synthesis of data from previous studies of Dalgleish et al. (1971) and Aktipis and co-workers (1973, 1974).
were all at counterion concentrations of 40 mM or greater is also shown. Our data in the high salt case agree with previous results, although we generally obtain slightly higher values of $\Delta \varepsilon_{\text{bound}}$ ($\sim 1 \text{ L/mol-cm}$) throughout the range of our mixtures; Houssier et al. (1974) obtained slightly higher $\Delta \varepsilon_{307}^{308}$ values also. Our low salt curve is in direct conflict with these previous studies: we find a change in the ionic strength of the mixture affects the magnitude of $\Delta \varepsilon_{\text{bound}}$. In the low salt solution, the induced CD at 307 nm is larger than in high salt, all things being equal. Furthermore, the difference between the two cases grows as the extent of binding increases.

We believe past failures to observe an effect on $\Delta \varepsilon_{\text{bound}}$ from different salt concentrations is attributable to the conditions used to vary $r$: constant dye and variable DNA concentrations. Variable DNA concentrations necessarily alter the effective counterion concentration; at lower salt concentrations these alterations can be sizable, often increasing the effective counterion concentration manyfold, as in this study. When we measured the induced CD of ethidium ion in the DNA/dye mixtures from the Scatchard analyses at low salt, the $\Delta \varepsilon_{307}^{307}$ vs. $r$ curve (data not shown) closely resembled that of the high salt solution. This was especially true at lower $r$ values where the amount of added DNA (and hence, added Na$^+$) was larger. Only when we kept the DNA concentration constant could we be sure of maintaining a fixed Na$^+$ concentration throughout the range of $r$ values. The fact that the dye is completely bound to the DNA under our experimental conditions allowed us to take this approach. At higher salt concentrations, 10 mM or greater, the variable DNA concentration in solutions from Scatchard analyses is less of a problem: the
Na⁺ contribution from the DNA aliquot is a smaller fraction of the total counterion concentration.

D) Counterion Changes with Dye Binding

The ability to alter the magnitude of Δε₃₀⁷ bound in DNA/ethidium ion complexes by simply lowering the counterion concentration is an intriguing observation. This phenomenon suggests that charged groups, either the counterions or the phosphate groups, or both in concert, may be responsible in part for these variations, and quite possibly, the increase in Δε₃₀⁷ bound with increasing r. We employ polyelectrolyte theory to examine this question. We begin our analysis from the viewpoint that electrostatic interactions between the ionic groups and the dye are solely responsible for variations in the CD spectrum with binding ratio. We will discuss other effects later.

The average axial distance per charge calculated via equation (2), and the effective charge/phosphate, ξ⁻¹, from equation (1), as a function of the extent of ethidium ion binding in DNA are presented in Figure 5.8. The combined effects of helix lengthening and charge neutralization result in a doubling of the effective phosphate charge in fully loaded DNA (r = 0.25) compared with unaltered DNA (r = 0.00). The values of b and ξ⁻¹, together with other quantities of interest, are presented at discrete values of r in Table VI.

According to these calculations, the interaction of ethidium ion causes a decrease in the condensed charge/phosphate ratio and an expansion of the volume within which the counterions are considered "bound". The net result of these two effects is a steep drop in the effective local concentration of the counterion, C₁₀C, as the extent of binding increases. The counterions are divided into two classes
Figure 5.8. Calculated distance per phosphate (b) and fractional charge per phosphate ($\xi^{-1}$) vs. binding ratio for ethidium ion intercalation in DNA. Equations (1) and (2) were used to create curves.
<table>
<thead>
<tr>
<th>Parameter</th>
<th>0.00</th>
<th>0.05</th>
<th>0.10</th>
<th>0.15</th>
<th>0.20</th>
<th>0.25</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bound dye/phosphate (r)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Axial distance/phosphate (b)</td>
<td>1.70 Å</td>
<td>1.97 Å</td>
<td>2.27 Å</td>
<td>2.60 Å</td>
<td>2.98 Å</td>
<td>3.40 Å</td>
</tr>
<tr>
<td>Charge/phosphate ($\zeta^{-1}$)</td>
<td>0.239</td>
<td>0.277</td>
<td>0.319</td>
<td>0.366</td>
<td>0.419</td>
<td>0.479</td>
</tr>
<tr>
<td>Counterions/phosphate ($\delta_1$)</td>
<td>0.761</td>
<td>0.723</td>
<td>0.681</td>
<td>0.634</td>
<td>0.581</td>
<td>0.521</td>
</tr>
<tr>
<td>Bound ion volume/mole phosphate ($V_p$)</td>
<td>643 cm$^3$</td>
<td>820 cm$^3$</td>
<td>1026 cm$^3$</td>
<td>1251 cm$^3$</td>
<td>1508 cm$^3$</td>
<td>1757 cm$^3$</td>
</tr>
<tr>
<td>Bound ion radius (a)</td>
<td>14.1 Å</td>
<td>14.8 Å</td>
<td>15.4 Å</td>
<td>15.9 Å</td>
<td>16.3 Å</td>
<td>16.5 Å</td>
</tr>
<tr>
<td>Local counterion concentration ($C_{\text{loc}}$)</td>
<td>1.18 M</td>
<td>0.88 M</td>
<td>0.66 M</td>
<td>0.51 M</td>
<td>0.38 M</td>
<td>0.30 M</td>
</tr>
</tbody>
</table>
in Manning's approach to polyelectrolyte properties: 1) those condensed on the macromolecule and 2) those remaining free in solution. In the current case, consideration of our results using these two classes is very helpful. The locally condensed counterions (and the phosphate groups) should exert a large influence upon the electronic properties of the bound dye molecule: they are physically closer to the binding sites and their concentration is much larger than those free in solution (and even the total counterion concentrations for our experiments). The atmosphere of uncondensed counterions, which extends outward from the outer radius of the condensed ion volume (a), should exert smaller effects upon dye electronic properties, because of both its greater distance from the binding sites and also its lower effective concentration.

One of the main tenets of Manning's theory is the strict requirement for counterion condensation to relieve the charge-charge repulsions in the helix backbone of DNA; condensation will occur even to the point of virtual depletion of the counterion in the Debye-Hückel atmosphere (Manning, 1977). For the Na\(^+\) concentrations employed in our study, \(\sim 0.45\) mM and \(\sim 86\) mM, this requirement applies and its consequence is simple: in both solutions the local concentrations of condensed counterions are those calculated in Table VI. Thus there is no difference between the two salt solutions at the level of the strongest electrostatic effect from the counterions on the dye, despite the vastly different total counterion concentrations.

A possible correlation between the local counterion concentration and the magnitude of \(\Delta_{\text{bound}}^{307}\) exists: the local counterion concentration decreases as both the extent of binding rises and, by extension
to the experimental results, $\Delta \varepsilon_{\text{bound}}^{307}$ rises. The locally bound counterions, together with the phosphates, constitute the source of a perturbing field on the bound dye. This field may be responsible for the variations in $\Delta \varepsilon_{\text{bound}}$ with $r$ between 300 and 350 nm since the local counterion concentration varies with $r$. To restate and generalize the correlation: higher counterion concentrations cause lower magnitudes in the induced CD between 300 and 350 nm of ethidium ion bound in DNA.

To qualitatively examine the possible effects of the Debye-Hückel atmosphere on the induced CD, we estimated the total Na$^+$ concentration in the low (~0.45 mM) salt and high (~86 mM) salt solutions. Using the binding data for DNA/EI complexes in Table VI, we calculate the amount of Na$^+$ ions removed from solution during condensation by multiplying the counterion/phosphate ratio, $e_1$, by the total DNA concentration (0.31 mM). Subtraction of this quantity from the total Na$^+$ concentration provides an estimate of the concentration of ions in the Debye-Hückel atmosphere that interact with the partially neutralized polyions. These values are presented at each $r$ value in Table VII.

The immediately obvious effect is the absence of any significant change at high salt in the Debye-Hückel atmosphere Na$^+$ concentration from the bulk Na$^+$ concentration in solution. This is due to the low concentration of DNA relative to the salt concentration. Conversely, the removal of Na$^+$ ions by condensation in low salt causes significant (30% to 50%) reductions in the concentration of free Na$^+$. We said earlier that any field due to these free counterions will be a weaker perturbant of the bound dyes' optical properties than the con-
<table>
<thead>
<tr>
<th></th>
<th>Bound/phosphate (r)</th>
<th>Counterions/phosphate ($\theta_1$)</th>
<th>Condensed counterions$^a$</th>
<th>Free counterions, low salt$^b$</th>
<th>Free counterions, high salt$^c$</th>
<th>Local counterion concentration ($C_{1}^{loc}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0.00</td>
<td>0.761</td>
<td>0.24 mM</td>
<td>0.21 mM</td>
<td>0.21 mM</td>
<td>1.18 M</td>
</tr>
<tr>
<td></td>
<td>0.05</td>
<td>0.723</td>
<td>0.22 mM</td>
<td>0.23 mM</td>
<td>0.23 mM</td>
<td>0.88 M</td>
</tr>
<tr>
<td></td>
<td>0.10</td>
<td>0.681</td>
<td>0.21 mM</td>
<td>0.24 mM</td>
<td>0.24 mM</td>
<td>0.66 M</td>
</tr>
<tr>
<td></td>
<td>0.15</td>
<td>0.634</td>
<td>0.20 mM</td>
<td>0.25 mM</td>
<td>0.25 mM</td>
<td>0.51 M</td>
</tr>
<tr>
<td></td>
<td>0.20</td>
<td>0.581</td>
<td>0.18 mM</td>
<td>0.27 mM</td>
<td>0.27 mM</td>
<td>0.38 M</td>
</tr>
<tr>
<td></td>
<td>0.25</td>
<td>0.521</td>
<td>0.16 mM</td>
<td>0.29 mM</td>
<td>0.29 mM</td>
<td>0.30 M</td>
</tr>
</tbody>
</table>

$^a$DNA concentration (0.31 mM) x $\theta_1$

$^b$Total counterion concentration = -0.45 mM Na$^+$.  

$^c$Total counterion concentration = -86 mM Na$^+$.  

TABLE VII  
DISTRIBUTION OF COUNTERIONS IN DNA/ETHIDIUM ION MIXTURES
densed ions because of both their greater distance and lower concentration. Comparison of the effective concentrations for the local and free counterions shows a tremendous difference between the two domains of up to four orders of magnitude.

The Debye-Hückel atmosphere and its potential field may be responsible for the difference between the $\Delta e_{\text{bound}}^{307}$ vs. r curves in the different salt solutions (Figure 5.7). In this case, the low salt curve, for which the free counterion concentration is lower than in the high salt solution, has consistently higher values for $\Delta e_{\text{bound}}^{307}$ at equal values for r. This free counterion field seemingly affects the dye CD just like the local counterion field: lower counterion concentrations cause larger magnitudes in the induced CD between 300 and 350 nm, all other things being equal. However, because these effects are weaker for the counterions in this atmosphere, the local counterion field remains the predominant perturbant. Thus, the shape of the $\Delta e_{\text{bound}}^{307}$ vs. r curve remains generally the same in each case; the effect of the weaker field is to displace the curve in low salt to higher $\Delta e_{\text{bound}}^{307}$ values.

4. Discussion

From the viewpoint of electrostatic effects, we have only considered interactions between the dye transitions and the potential fields of the counterions, both condensed and free. Possible significant correlations between the magnitude of the induced CD in the near UV and the strength of these fields (as measured by the ion concentrations) are evident in our results. The fields attributed to these counterions are but a part of the total electrostatic picture and the ions are only well-characterized along a radial direction.
from the helix axis; they are not bound in any specific fixed site (Manning, 1978) relative to the intercalated dye.

We have ignored the geometrically "fixed" charge groups in the nucleic acid/dye complex. These groups are the phosphates, each with a \(-1\) charge (the nucleic acid plus condensed ions constitute the species of interest for the polyelectrolyte theory, but the formal charges on each ionic group have not been removed), and other intercalated ethidium ions, each with a \(+1\) charge. Each phosphate (two in all) connecting the nucleosides comprising the two nearest neighbor base pairs of the binding site and probably each phosphate (four more) in the immediately adjacent sites excluded from binding are geometrically invariant relative to the dye, no matter what the binding ratio, and so contribute to the CD as part of the asymmetry of the site. Phosphates farther away than two base pairs, and other dye molecules, constitute a possible binding ratio-dependent electrostatic contribution to the total induced CD of the ethidium ion because the relative position of these groups to a bound dye will depend upon how far they are from the dye and the extent of binding. This contribution to the total electrostatic potential field, together with that of the counterions, represents an attractive mechanism for the binding ratio dependence of the near UV induced CD in DNA/EI complexes. This theory was proposed by Lee and co-workers (1973), who recognized the possible role of other dye molecules intercalating near previously bound dye. We have taken it a step further and introduced the counterions' contributions to the perturbing field.

The geometry of DNA in aqueous solution (Bram & Beeman, 1971;
Bram, 1971) is very similar to the B form geometry obtained in fiber diffraction studies (Arnott & Hukins, 1972). The solution geometry of the DNA was also independent of ionic strength between the values of 50 to 150 mM NaCl in wide-angle X-ray scattering experiments (Bram, 1971) and between the values of 1.3 to 200 mM NaCl in viscosity measurements (Rosenberg & Studier, 1969). These types of measurements are not as sensitive to small changes in the double helix geometry as the CD spectrum is. Recently, Johnson et al. (1981) reported a study in which magnitude changes in the CD band at 275 nm for nucleic acids were correlated with changes in the helix winding angle and the propeller angle (twist) between the two bases in a base pair.

We measured the CD of our dialyzed DNA in the two salt solutions used in the induced CD study (data not shown). There was a decrease of ∼10% in the magnitude of the long wavelength CD band when measured at lower salt concentrations. Pardi (1980) saw a comparable effect in his study with E. coli DNA. These decreases are due to conformational changes in the DNA: most likely a slight decrease in the propeller angle and/or a slight increase in the winding angle (Johnson et al., 1981). These different geometries of the nucleic acid can affect the induced CD of the dye in other ways besides the effects due to different geometries for the charged groups relative to the dye. Any significant interactions between the bound dyes and bases farther from the base pairs of the binding site would also be affected. This is a second possibility for the origin of the difference between the $\Delta \varepsilon_{\text{bound}}^{307}$ vs. $r$ curves we measured earlier (Figure 5.7). The difficulty in trying to decide between the relative importance of different geometries vs. charge effects as the source of induced CD
variations lies in the interconnection of the two: a change in one necessitates a change in the other. CD calculations offer the best chance for resolving the question. We discuss this further in the next chapter.
Chapter VI

CONCLUSIONS

1. The Problem Revisited

We began this dissertation with the intention of settling two questions about the induced circular dichroism (CD) of ethidium ion in nucleic acids. First, what is the contribution of the inherent asymmetry of the intercalation site to the induced CD of the ethidium ion between 300 and 350 nm? Second, what possible mechanism(s) account for the increase in the molar CD per bound dye ($\Delta_\text{e}_{\text{bound}}$) at 307 nm as the extent of binding increases? The two hypotheses presented up to the time of this writing address the second question more than the first. One of them, the dye-dye exciton mechanism (Aktipis & Kindelis, 1973; Houssier et al., 1974), holds that the site asymmetry CD contribution is small and the greater likelihood of dye-dye interactions at higher binding ratios accounts for the increase in $\Delta_\text{e}_{\text{bound}}$. The second mechanism links increases in the induced CD of the dye with the changes intercalation produces in the potential field or environment of the bound dye(s) (Lee et al., 1973). In the course of this study we have obtained new evidence bearing upon each of these hypotheses. We will review these discoveries and their implications, after which we will propose a model which may be the answer to the questions above. Finally, we propose some experiments which will test our model or provide some useful information.

2. New Evidence Bearing on the Problem

One of the first clues that the dye-dye exciton mechanism might not be responsible for the changes in the induced CD between 300 and
350 nm of bound ethidium ion was the large magnitudes of the induced CD for 2:1 dimer:dye complexes. These magnitudes per bound dye at 307 nm were of comparable size to the spectrum from fully loaded DNA (Par-di, 1980). For example, $\Delta \varepsilon_{\text{bound}}^{307}$ for CpG:CpG:EI is 22 L/mol-cm, while in calf thymus DNA at $r = 0.25$, $\Delta \varepsilon_{\text{bound}}^{307}$ is ~19 L/mol-cm (see Figure 1.5). Since only one dye molecule is present in the complex with the dimers, and it is intercalated between the base pairs of the minihelix, the large magnitude of its induced CD is not attributable to dye-dye interactions.

We measured the induced CD of ethidium ion intercalated in several more sequences of complementary dimers. The results all reflect one fact: the induced CD per bound dye is quite large between 300 and 350 nm. These complexes represented the simplest unit of the bound drug in complexes with the nucleic acids, and as such, their CD spectra are attributable to only one thing: the asymmetry of the binding site. This result directly contradicts one of the basic assumptions of the dye-dye exciton mechanism, namely, that the induced CD due to the asymmetry of the site is low.

Another piece of evidence cited in favor of the dye-dye exciton mechanism was the occurrence of a negative lobe in the CD spectrum centered near 295 nm at high ionic strengths (Aktipis & Kindelis, 1973; Balcerski & Pysh, 1976). This band, of roughly equal size with the positive lobe centered near 307 nm, is masked at lower ionic strengths by the larger, positive lobe of the nucleic acid's CD near 270 nm, according to these authors. A conservative CD spectrum such as this is characteristic of an exciton interaction between identical chromophores (Tinoco, 1963).
In our studies of the 2:1 dimer:dye complexes via fluorescence detected circular dichroism, we obtained the CD spectrum of these complexes well below 300 nm, where the large excesses of dimers present in the mixtures usually mask the complexes' CD spectra. We found either true negative lobes or relative minima in the CD spectra near 295 nm for most of the 2:1 complexes studied. In all cases, a positive CD lobe occurs above 300 nm. Since only one dye is present in each complex, dye-dye excitons are not responsible for this pattern of bands. This suggests that such bands in the CD spectra of DNA/EI complexes need not solely be attributed to dye-dye excitons either. Another exciton mechanism involving transitions on both the DNA bases and the dye was presented by Houssier et al. (1974). Such interactions might be operating here, although the magnitude of the bands would not change with the binding ratio. This type of spectral contribution would be grouped with others from the asymmetry of the site.

In our experiments on the binding behavior and circular dichroism of ethidium ion in calf thymus DNA mixtures of different salt concentrations, we found that the concentration of the counterions affected the magnitude of $\Delta \varepsilon_{307}^{\text{bound}}$ for solutions at the same binding ratio, $r$. The fact that an environmental effect, such as the counterion concentration, can produce changes in the CD spectrum is reminiscent of the mechanism of Lee et al. (1973), which connects changes in the bound dye's optical properties with changes in the perturbing potential field at the binding site as other sites fill. Using electrostatic theories of DNA's polyelectrolyte behavior (Manning, 1978), we examined possible sources of the change in the field: the atmosphere of counterions, both condensed and free, which associate with the nucleic acid. Other con-
ceivable sources of change in the field are the relative positions of other charged groups with the dye: the phosphates of the helix backbone and other intercalated dye molecules, and the relative orientations of base pairs beyond the binding site with the dye.

3. The Model

We have discovered some facts which suggest that the variation of $\Delta E_{\text{bound}}^{307}$ with the binding ratio, $r$, is not the result of an increased chance of dye-dye exciton interactions at higher binding ratios. Before proposing a model based upon the alternate mechanism, we realize we have by no means uncovered an overwhelming amount of evidence to prove or disprove either mechanism. Instead, we believe the facts are more consistent with an explanation based upon a mechanism incorporating the perturbing potential field as the source of the spectral behavior.

The main points of the model are as follows: 1) the spectral properties (CD) of the 2:1 dimer:dye complexes best represent those of the nucleic acids at high binding ratios; 2) the potential field arising from charged groups contributes a component to the induced CD which reduces the magnitude of $\Delta E_{\text{bound}}^{307}$; 3) at the lowest binding levels, this last component is dominant, so $\Delta E_{\text{bound}}^{307}$ is reduced to its lowest levels; and 4) $\Delta E_{\text{bound}}^{307}$ increases as $r$ increases because the combined geometric and electrostatic changes associated with the intercalation of dye diminish the contribution of the perturbing field, and the CD contribution from the asymmetry of the site (i.e., the dimer:dye spectrum) gains ascendancy in the sum of all effects.

In selecting the CD spectrum of fully bound DNA/EI complex as the analogue of the 2:1 dimer:dye complexes' spectra, we recognize that
The arrangement of bases and dye in the binding site may be freer to assume energetically favorable conformations in the dimer:dye complexes than in the longer sequences, where the adjacent bases and the backbone may restrict the number of possible conformations. Alternatively, the other perturbants of the dye's electronic properties (electrostatic fields, etc.) may serve to make the values for $\Delta \varepsilon_{\text{bound}}^{307}$ in each site more uniform in the polymer. We feel compelled to use the dimer/dye complexes as models for the fully loaded DNA because they display large magnitudes for $\Delta \varepsilon_{\text{bound}}$, because the spectra for the DNA/dye complexes maintain shapes similar to the dimer/dye spectra throughout the range of binding ratios, and because the magnitude of $\Delta \varepsilon_{\text{bound}}^{305}$ for ethidium ion bound in both dC-dG sites of pdC-dG-dC-dG minihelices is virtually identical with $\Delta \varepsilon_{\text{bound}}^{305}$ for the dCpG:dCpG:EI complex (Chapter II).

Once we select the asymmetry of the binding site as the basis for the large magnitudes in $\Delta \varepsilon_{\text{bound}}^{307}$ at the high binding ratios, the question of variations in the CD with the binding ratio can be approached from a new point of view. Rather than ask why $\Delta \varepsilon_{\text{bound}}^{307}$ starts off at low values and increases with $r$, we now want to know why it decreases as more dye molecules return to solution ($r$ is decreasing). If we make use of the perturbing field mechanism, we can answer this: the strength of the perturbation varies with the binding ratio. Whether the electrostatic field, the relative geometry of more distant bases and the bound dyes, or other sources constitute the sole source of the field, or all contribute to it is difficult to say just at this time.
condensed counterion concentrations, an electrostatic effect, but whether this plays any role in altering the bound dyes' induced CD requires further study. Whatever the source of the perturbing field, its contribution to the total CD of the dye at 307 nm is negatively signed and so subtracts from the (assumed virtually constant) CD contribution from the site asymmetry.

4. Further Experiments

We have arrived at a model for the behavior of the induced CD between 300 and 350 nm of ethidium ion bound in DNA. The evidence supporting this model is certainly scanty at this point, but with the following experiments, support and refinement (and possibly overthrow) of the model may be forthcoming.

One of the weak links in the model is our reliance upon data from dimer/dye studies and DNA/dye studies only: the two are linked in the model even though there is very little evidence that anything true at one extreme holds at the other. Further studies of ethidium ion binding with complementary oligomers will aid in determining the validity of the correspondence. In particular, the question of whether a dimer/dye complex spectrum (CD) adequately represents longer sequences can be answered with CD measurements at different dye:helix ratios with complementary sequences like \( rA_n + rU_n \), where all potential intercalation sites have the same sequence, or \( r(C-A)_n + r(U-G)_n \), where two types of intercalation site exist, but one is presumably preferred over the other because it is \( Py(3'-5')Pu \) instead of \( Pu(3'-5')Py \). Both types of sequences could be studied as polymers, also, and the deoxyribo-analogues could be compared with the ribo-sequences.

The effect of environment and/or perturbing field on the induced
CD of ethidium ion in DNA could be tested further by measuring the CD spectrum of the dye in the presence of competing intercalating species, such as proflavine and methylene blue (Lee et al., 1973), which are also cations, or 4-nitroquinoline-1-oxide (4-NQO) (Winkle, 1979), which is uncharged. In each case the charge density of the DNA would change as both the ethidium ion and the competing species bound. If the binding ratio of ethidium ion to the DNA, $r_{EI}$, and the binding ratio of the competing species, $r_{comp}$, could be obtained in some manner, then, by comparison of the magnitude of the ethidium ion's induced CD at 307 nm in this system with that of the ethidium ion alone with DNA at the same $r_{EI}$ value, one could determine the validity of the environment/perturbing field model. The model predicts that $\Delta\varepsilon_{307}^{bound}$ in the ethidium ion/competitor/DNA mixture would be larger than $\Delta\varepsilon_{307}^{bound}$ in the ethidium ion/DNA mixture because the total of occupied sites is greater in the former. The two cationic dyes would work well because their induced CD bands are at wavelengths greater than 400 nm; 4-NQO would work well because it apparently does not acquire an induced CD spectrum when intercalated in nucleic acids (Winkle, 1979).

Finally, computer calculations of the CD spectrum for ethidium ion bound in different dimer sequences or in a polymer would be useful, at least for predicting changes in the spectrum as the nearby bases changed. The coordinates for ethidium ion bound in 5-iodoUpA and 5-iodoCpG from the X-ray studies (Tsai et al., 1977; Jain et al., 1977) would be a good starting point for the dimer/dye calculations. Sobell and co-workers (1977) have also published coordinates for ethidium ion bound in a DNA sequence by juxtaposing the dimer coordinates (representing the binding site) with the B form coordinates (representing
the remainder of the molecule). Beginning with the dimer complex and adding succeeding base pairs to each end of the structure, one could obtain a clearer picture of the role of the base pairs outside the binding site on the induced CD of the bound dye with calculations on each new structure. Similarly, one could investigate the effect of the electrostatic fields of the phosphates and the counterions on the spectrum using the larger structure. The methods of calculation employed by Johnson et al. (1981) seem particularly useful for this problem.
BIBLIOGRAPHY

New Jersey.

Kastrup, R. V., Young, M. A., & Krugh, T. R. (1978) *Biochemistry* 17,


Record, M. T., Jr. (1975) Biopolymers 14, 2137-2158.


Tinoco, I., Jr. (1963) Radiation Res. 20, 133-139.


*Die Makromolekulare Chemie*, in press.


Appendix A

COMPUTER PROGRAMS FOR ERROR ANALYSIS AND DATA FITTING
IN BENESI-HILDEBRAND PLOTS AND \( \Delta c_{\text{bound}} \)

1. Benesi-Hildebrand Plots

A) Description

The programs BHFIT1, BHFIT2, and BHFIT3 fit dimer/dye titration data to 1:1, 2:1, or 1:1:1 binding stoichiometries, calculate the errors associated with the binding constant and \( \epsilon_b - \epsilon_f \), and prepare plots of the data using the general graphics package in OS/8. The programs are written in FORTRAN, compiled with .R FORT under the /I and /O options, and saved as core image files with .SAVE SYS BHFITN, where \( N = 1, 2, \) or 3. All listings are available on paper tape.

The data consists of absorbance differences between the dye alone and the mixture of dimer and dye at some wavelength (DABS), the initial dinucleoside phosphate concentrations ([NUC1] and [NUC2]), and the estimated errors (standard deviations) in each, if desired. These values correspond to \( A - \epsilon^0_{E1}, C^0_{NpN_a}, \) and \( C^0_{NpN_b} \) in equation (4) of Chapter II, respectively. The first fitting does not weight the data in any fashion; the second fits the data by weighting each point according to its associated error.
B) Listing

i) BHFIT1

C THIS PROGRAM FITS DIFFERENCE IN ABSORBANCE VERSUS ADDED NUCLEOTIDE CONCENTRATION DATA IN A BENESI-HILDEBRAND PL...
C ENTER DATA. OBSERVABLE IS ABSORBANCE DIFFERENCE NORMALIZED TO 1 CM PATH. CHUC IS ADDED NUCLEOTIDE CONCENTRATION. OCHER IS ERROR IN ADDED NUCLEOTIDE CONCENTRATION, AND YER IS ERROR IN Y VALUE AS DERIVED FROM THE ERROR IN DELTA ABSORBANCE.

15      WRITE(1,510)
      READ(1,520)IDYE
      LIMLO=0
      LIMHI=0
      READ(1,530)IER
      DO 29 I=1,IDYE
         WRITE(1,510)
         READ(1,540)NPTS(I)
         READ(1,550)CDYE(I)
         LIMLO=LIMHI-1
         LIMHI=LIMHI+NPTS(I)
         WRITE(1,560)
      WRITE(1,565)
      DO 28 M=LIMLO,LIMHI
         READ(1,600)CN1(M)
         WRITE(1,700)CHF1T2)
      20     CONTINUE
         WRITE(1,710)CN1(M),CN2(M),DABS(M),ER1(M),ER2(M),YER(M)
         ALF1(M)=0
      35     CONTINUE

C AT THIS POINT. THE PROGRAM CONTINUES ON THE FILE 'SHFIT2.SY'.

      CALL CHAIN('SHFIT2')
      END
C THIS PROGRAM FITS DIFFERENCE IN ABSORBANCE VERSUS ADDED
C NUCLEOTIDE CONCENTRATION DATA IN A BENESI-HILDEBRAND PLOT.

COMMON FNPT,FNDA,NSIZE,NTHETA,NSYM,NCUR,
  NTIT,TITLE,XTIT,TITLE,XTIT,XTIT,XTIT,XTIT,XTIT,XTIT,XTIT,XTIT,XTIT,XTIT,
  XMAX, XMIN, XMAX, XMIN, XMAX, XMIN, XMAX, XMIN, XMAX, XMIN,
  NCH, XFACT, XSIGN, NFACT, YFACT, YSIGN, YMAX, NMAX,
  NSLAB, NORM, NORYL, NSITIT, NORMT, NORYT,
  XFACT, YSIGN, NMAX, YFACT, YSIGN, YMAX,
  DABS,YEP, CN1, ER1, CN2, ER2, X, SOX, Y, SOY, ALF1,
  N, SGX, SGY, NPTST, CDYE, IDY, K, RK, ERNRM, IER
DIMENSION TITLE(7), XTIT(7), YTITLE(7)
DIMENSION DABS(15), CN1(15), CN2(15), SGX(15), SGY(15)
DIMENSION ALF1(15), X(15), Y(15), NPTST(5), CDYE(5)
DIMENSION ER1(15), ER2(15), YER(15)

530 FORMAT(2000H)
540 FORMAT(/'FIRST RUN DOES NOT WEIGHT FOR ERRORS;'/)
550 FORMAT(/'SECOND RUN BEGINS WITH FIRST RUN VALUES, BUT WEIGHTS FOR ERRORS.'/
560 FORMAT(/)

C FIRST RUN DOES NOT WEIGHT FOR ERRORS,
C SECOND RUN BEGINS WITH FIRST RUN VALUES, BUT WEIGHTS FOR ERRORS.
C A IS Y INTERCEPT, B IS SLOPE.
C IFLAG IS RUN NUMBER FLAG: 0=1ST, 1=2ND.

A=1.
B=1.
SGX=1.
SGY=1.
IFLAG=0
WRITE(1,500)
WRITE(1,510)
40 EQUKO=0
C CALCULATE X AND Y GRAPH VALUES AND THEIR ASSOCIATED ERRORS.
C Y ERROR = ENTERED Y ERROR + PROPAGATED X ERROR.
C SGX AND SGY ARE VARIANCES; SGA AND SGB ARE STANDARD DEVIATIONS.
C ERRNRM IS THE LOWEST Y ERROR FOR NORMALIZING ALL ERRORS IN WEIGHING.
C YVALN IS AMOUNT OF FREE DIMER.

42    ERNRM=0
    LIMLO=0
    LIMHI=0
    DO 55 I=1, IDVE
        LIMLO=LIMHI+1
        LIMHI=LIMHI+NPTS(I)
    DO 50 M=LIMLO,LIMHI
        CALL FREE(VALNEV,FRE1,FRE2,M,I)
        X(M)=1./VALNEV
        Y(M)=CDEV(I)/DABS(M)
        CN1ER=(ER1(M)/FRE1)**2.
        CN2ER=(ER2(M)/FRE2)**2.
        SGX(M)=X(M)**2.*CN1ER+CN2ER
        SGY(M)=VER(M)**2.
        $B+SGX(M)+X(M)*X(M)*SGB+SGA+SGA
        WRITE(1,520)X(M),Y(M),ALF1(M)

C ASSIGN FIRST ERROR AS NORMALIZING VALUE TO START WITH;
C THEREAFTER, SUBSTITUTE NEW ERROR VALUE IF SMALLER.
C THIS SIMPLY KEEPS NUMBERS FROM OVERFLOWING IN FLTSQ.

        IF(M-1) 47.48.47
        IF(ERNRM-SGY(M)) 50.50.48
        ERNRM=SGY(M)
    50    CONTINUE
    55    CONTINUE

C FIT LINE.

    CALL FLTSQ(IFLAG)
C CALCULATE FRACTION OF DYE BOUND IN COMPLEX TO WITHIN .001 ALPHA.

ECK=A/B
KK=0
LINLO=0
LINHI=0
DO 66 M=1, IDYE
   LINLO=LINHI+1
   LINHI=LINHI+NPTS(M)
DO 65 J=LINLO, LINHI
   DO 62 I=1,100
      CALL FREE (VALNEW, FRE1, FRE2, J, M)
      ALPH=ECK/VALNEW
      ALPH=ALPH/(1. +ALPH)
      DIFF=ABS(ALPH-ALF1(J))
      ALF1(J)=ALPH
      IF (DIFF-.001*ALF1(J)) 64,62,62
   CONTINUE
   WRITE(1,530)J
   KK=1000
   CONTINUE
66 CONTINUE
C IF CONVERGANCE OF ALPHA DID NOT OCCUR, ABORT RUN.
   IF (KK=1000) 66,62,66
C CALCULATE EQUILIBRIUM CONSTANT TO WITHIN .01 K.
68 DIFF=ABS(ECK-EQUKO)
   EQUKO=ECK
C IF CONVERGANCE OF K DID NOT OCCUR, RECALCULATE WITH NEW VALUES.
   IF (DIFF-.01*ECK) 70,42,42
   IF (IFLAG) 80,72,80
C CHECK TO SEE IF CALCULATION WITH WEIGHING ERRORS IS POSSIBLE.
72 IF (IER=1) 80,74,80
   WRITE(1,500)
   READ(1,540)I
   IF (I) 80,76,80
   WRITE(1,550)
   IFLAG=1
   GO TO 40
76 WRITE(1,560)
80 GO TO 40
C AT THIS POINT, THE PROGRAM CONTINUES ON THE FILE 'BFIT3.SV'.
   CALL CHAIN('BFIT3')
C RETURN TO START IF ALPHA DID NOT CONVERGE.
82 CALL CHAIN(FNAET)
   END
a) Subroutine FREE

This program fits difference in absorbance versus added nucleotide concentration data in a Benesi-Hildebrand plot.

SUBROUTINE FREE (VALNEW, FRE1, FRE2, M, I)
COMMON FRET, FNDATA, N, NSIZE, NTHETA, NSYM, NCOR,
   NT1T, TITLE, XNT1T, XTITLE, YNT1T, YTITLE, NNEW,
   XMIN, XMAX, XINC, YMIN, YMAX, YINC,
   XST, XTIC, XINC, YST, YTIC, YINC, LTIC,
   NSLAB, NORM, N0RL, N0SLT1T, N0RT, NORT, NORYT,
   NFACT, YSIGN, NMAX, YFACT, YSIGN, YMAX,
   DABS, YER, CN1, ER1, CN2, ER2, K, SGX, Y, SGY, ALF1,
   GN, SG, B, SGF, NPTS, NFST, CDVE, IDVE, K, RK, ERNRM, IER

DIMENSION TITLE(7), XTITLE(7), YTITLE(7)
DIMENSION DABS(15), CN1(15), CN2(15), SGX(15), SGY(15)
DIMENSION ALF1(15), X(15), Y(15), NPTS(5), CDVE(5)
DIMENSION ER1(15), ER2(15), VFR(15)

C calculate the amount of dimer free at equilibrium

FRE1=CN1(M)-RK*ALF1(M)*CDVE(I)
FRE2=CN2(M)-RK*ALF1(M)*CDVE(I)
IF(K-2)<10, 20, 30
10 VALNEW=FRE1
20 FRE2=1.
GOTO 40
30 RETURN
40 RETURN
END
b) Subroutine FLTSQ

This program fits difference in absorbance versus added nucleotide concentration data in a Benesi-Hildebrand plot.

SUBROUTINE FLTSQ (IFLAG)
COMMON FNDEF, FNDATA, N, NSIZE, NTIT, NSYM, NCOR,
2 NTIT, NTIT, NTIT, NTIT, NTIT, NTIT, NTIT, NTIT, NTIT, NTIT,
3 MAXMIN, XKMIN, XKMAX, YKMIN, YKMAX, YKMAX, YKMIN, YKMAX,
4 XST, XTEX, XINC, YST, YTEX, YINC, LITIC,
5 NSILAC, NSILB, NSILC, NSILD, NSIZE, NMAX, NSYM, NMIN,
6 XPARAM, XSYM, XMIN, XMAX, XNORM, YNORM, NPARAM,
7 DOES, YER, CN1, ER1, CN2, ER2, X, SGX, Y, SGY, ALF1,
8 A, SGB, B, SGB, NPTS, NPTT, CDVE, IDVE, IRE, RK, ERNRM, IER
DIMENSION TITLE (7), NTITLE (7), NTITLE (7)
DIMENSION XDATA (15), YDATA (15), XDATA (15), YDATA (15), YDATA (15)
DIMENSION ALF1 (15), XDATA (15), YDATA (15), NDATA (5), CDVE (5)
DIMENSION ER1 (15), ER2 (15), VER (15)

C CALCULATE THE LEAST SQUARES FIT TO A LINE FOR A DATA SET.
C MODIFIED FROM THE PROGRAM 'LINFIT' IN 'DATA REDUCTION
C AND ERROR ANALYSIS FOR THE PHYSICAL SCIENCES', BY
C P. R. BEVINGTON, McGRAW-HILL, 1969. PP. 104-5.

500 FORMAT ('Y INTERCEPT = 'E12.5', SLOPE = 'E12.5')
510 FORMAT ('SIGMA B = 'E12.5', SIGMA M = 'E12.5')
520 FORMAT ('R = 'F8.5')
530 FORMAT ('K = ( 'E10.5', ')
SUM=0
SUMX=0
SUMY=0
SUM2=0
SUM2Y=0
SUMXY=0
DO 50 L=1, NPTST
C IF NO ERRORS ENTERED, WEIGHT = 1. (ONLY WILL RUN ONCE).
XI=X(L)
YI=Y(L)
WEIGHT=ERNRM/SGY(L)
IF (IFLAG) 49, 49, 49
C IF NOT WEIGHING FOR ERRORS, WEIGHT = 1.
48 WEIGHT=1.
49 SUM=SUM+WEIGHT
SUMX=SUMX+WEIGHT*XI
SUMY=SUMY+WEIGHT*YI
SUM2=SUM2+WEIGHT*XI*XI
SUM2Y=SUM2Y+WEIGHT*YI*YI
SUMXY=SUMXY+WEIGHT*XI*YI
CONTINUE
DELTA=SUM2-2.*SUMX*SUMY/DELTA
A=(SUMXY-SUM2)*SUM2Y/DELTA
B=(SUMY-SUMX)*SUM2X/DELTA
C=NPTST-2.
VARNCE=(SUM2+2.*SUMX*SUMY/SUM2X)/C
C IF WEIGHING FOR ERRORS, VARNCE=SMALLEST Y ERROR.
IF (IFLAG) 51, 52, 51
51 VARNCE=ERNRM
52 SGN=SQRT(VARNCE+SUM2)/DELTA
SGB=SORT(VARNCE+DELTA)
R=(SUMXY-SUM2)*SORT(DELTA+(SUM2+2.*SUMX*SUMY))/C
WRITE (1, 500) A, B
WRITE (1, 510) SGA, SGB
WRITE (1, 520) R
END
iii) BHFIT3

C THIS PROGRAM FITS DIFFERENCE IN ABSORBANCE VERSUS ADDED NUCLEOTIDE CONCENTRATION DATA IN A BENESI-HILDEBRAND PLOT.

COMMON FINT, FDATA, N, NSIZE, NTHETA, NWM, NCOR,
NTIT, TITLE, NTITX, TITLX, NTITY, TITLY, NEW,
XMIM, XMAX, XINCH, YMIN, YMAX, YINCH,
XST, XTIC, XINC,YST, YTIC, YINC, YTIC,
NSLAB, NORML, NORMR, NSITIT, NORXT, NORYT,
XFACT, XSIGN, YNAM, YFACT, YSIGN, YNAM,
DABS, YER, CNI, ER1, CN2, ER2, X, SQX, Y, SQY, ALF1,
A, SQA, B, SQB, NPTS, NPTST, CDVE, IDVE, K, RK, ERNRM, IER
DIMENSION TITLE(7), XTITLE(7), YTITLE(7)
DIMENSION DATS(15), CN1(15), CN2(15), SQX(15), SQY(15)
DIMENSION ALF1(15), X(15), Y(15), NPTS(5), CDVE(5)
DIMENSION ER1(15), ER2(15), YER(15)
DIMENSION XX(15), YY(15)

C PRINT OUT FINAL VALUES FOR K, DELTA EPSILON, X, Y [COMPLEX], AND ERRORS IN EACH [NOT [COMPLEX]].
C PROCEED TO PLOTTING ROUTINE, IF DESIRED.

209 FORMAT('J=', '12', 'X(J)=', 'E13.5', 'Y(J)=', 'E11.3', 'ALPHA(J)=', 'F7.4')
209 FORMAT('J=', '12', 'X(J)=', 'E12.4', 'FINAL DELTA EPSILON=', 'E12.4')
210 FORMAT('J=', '12', 'X(J)=', 'E12.4', 'FINAL DELTA EPSILON=', 'E12.4')
211 FORMAT('J=', '12', 'X(J)=', 'E12.4', 'DXY(J)=', 'E10.2' [COMPLEX]= 'E12.4')
250 FORMAT('')
250 FORMAT('')
250 FORMAT('')
600 FORMAT('NORMALIZER TO MAKE X AXIS INTEGRAL = ', 'E12.4')
610 FORMAT('NORMALIZER TO MAKE Y AXIS INTEGRAL = ', 'E12.4')
620 FORMAT('NORMALIZER TO MAKE X AXIS INTEGRAL = ', 'E12.4')
620 FORMAT('NORMALIZER TO MAKE X AXIS INTEGRAL = ', 'E12.4')
700 FORMAT('NEW GRAPH? (NO=0, YES=1)', '11')
710 FORMAT('NEW PARAMETERS?', '11')
720 FORMAT('XMIN=0.0, XST=XMIN')
730 FORMAT('XMAX=', 'E12.4')
735 FORMAT('XINCH=', 'E12.4')
740 FORMAT('XTIC=XINC=', 'E12.4')
750 FORMAT('YMAX=0.0, YST=YMIN')
760 FORMAT('YMIN=', 'E12.4')
765 FORMAT('VINCH=', 'E12.4')
770 FORMAT('VTIC=VINC=', 'E12.4')
780 FORMAT('SYMBOL #=', '11')
785 FORMAT('SYMBOL SIZE=', '11')
790 FORMAT('CALCOMP(0) OR TEKTRONIX(1)', '11')
800 FORMAT('ALL OK?', '11')
810 FORMAT(2(16) -
DO 2 J=1,NPTST
   WRITE(1,208)J,XX(J),YY(J),ALF1(J)
2 CONTINUE
DELEP=L/4
EDK=A/V
AE2=SGA*SGA/(A+A)
BE2=SGB*SGB/(B+B)
ERK=ABS(EDK+SORT(AE2+BE2))
ERD=ABS(DELEP+SORT(AE2))
WRITE(1,209)EDK,DELEP
WRITE(1,210)ERK,ERD
LIMLO=0
LIMHI=0
DO 4 J=1,1DYE
   LIMLO=LIMHI+1
   LIMHI=LIMHI+NPTS(I)
4 CONTINUE
DO 3 J=LIMLO,LIMHI
   ERN=SORT(SGH(J))
   ERV=SORT(SGY(J))
   CNNUC=ALF1(J)+CDYE(I)
   WRITE(1,211)J,ERN,ERV,CNNUC
3 CONTINUE
WRITE(1,250)
READ(1,260)
!F(I)8,6,8
CALL CHAIN(FNRET)

C SET VARIABLES THAT ARE UNCHANGED

3 NTHETA=0
NCOR=100
NTIT=0
NXTIT=0
LT1C=0
NSILAB=2
NSITIT=2
NORKL=0
NORKL=0
NORKT=0
NORKT=1
C ENTER "NORMALIZING" POWERS OF TEN

10 WRITE(1,250)
READ(1,600)XXMNM
READ(1,610)YYMNM
DO 20 I=1,NPTST
XX(I)=XX(I)/XXMNM
YY(I)=YY(I)/YYMNM
WRITE(1,620)I,XX(I),YY(I)
20 CONTINUE
READ(1,800)INP
IF(INP)22,10,22

C PREPARE FOR PLOTTING

22 WRITE(1,250)
READ(1,700)NEW
READ(1,710)INP
IF(INP)25,30,25
25 XMIN=0
WRITE(1,720)
READ(1,730)XMAX
READ(1,735)XINCH
XST=XMIN
READ(1,740)XTIC
XINC=XTIC
YMAX=0
WRITE(1,750)
READ(1,760)YMIN
READ(1,765)YINCH
YST=YMIN
READ(1,770)YTIC
YINC=YTIC
30 READ(1,780)NSYM
READ(1,785)NSIZE
READ(1,790)INP
IF(INP)35,40,35
35 NCR=150
40 READ(1,900)INP
IF(INP)50,22,50
50 CALL OPEN("DSK",FNDATA)
DO 55 I=1,NPTST
WRITE(4,810)XX(I),YY(I)
55 CONTINUE
H=NPTST
CALL CLOSE

C AT THIS POINT, THE PROGRAM TRANSFERS CONTROL TO THE GENERAL
C PLOTTING SYSTEM. RETURNING AFTER PLOTTING TO 'BHF11.FT'.

CALL CHAIN('GENPNT')
END
C) Sample Run

.R B6F11
1:1(#1), 2:1(#2), OR 1:1:1(#3) COMPLEX? 2

HOW MANY DIFFERENT [DYE]? 1
ENTER ERRORS? (1=YES, 2=NO) 1

# POINTS= 6
[DYE]= .043E-03

<table>
<thead>
<tr>
<th>[NUC1]</th>
<th>[NUC2]</th>
<th>DABS</th>
<th>ERROR</th>
<th>ERROR</th>
<th>ERROR</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.5E-03</td>
<td>- .022</td>
<td>.05E-03</td>
<td></td>
<td></td>
<td>22E-03</td>
</tr>
<tr>
<td>2.0E-03</td>
<td>- .040</td>
<td>.06E-03</td>
<td></td>
<td></td>
<td>14E-03</td>
</tr>
<tr>
<td>2.0E-03</td>
<td>- .040</td>
<td>.06E-03</td>
<td></td>
<td></td>
<td>14E-03</td>
</tr>
<tr>
<td>2.0E-03</td>
<td>- .063</td>
<td>.03E-03</td>
<td></td>
<td></td>
<td>63E-04</td>
</tr>
<tr>
<td>2.0E-03</td>
<td>- .390</td>
<td>.02E-03</td>
<td></td>
<td></td>
<td>42E-04</td>
</tr>
<tr>
<td>7.0E-03</td>
<td>- .112</td>
<td>.16E-03</td>
<td></td>
<td></td>
<td>24E-04</td>
</tr>
</tbody>
</table>

[DYE]= .0430E-04 # POINTS= 6

<table>
<thead>
<tr>
<th>[NUC1]</th>
<th>[NUC2]</th>
<th>DABS</th>
<th>DELTA</th>
<th>ERROR</th>
<th>ERROR</th>
<th>ERROR</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.150E-02</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.15E-01</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.500E-04</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.50E-03</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.200E-02</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.20E-01</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.500E-04</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.50E-03</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.200E-02</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.20E-01</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.500E-04</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.50E-03</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.250E-02</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.25E-01</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.500E-04</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.50E-03</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.780E-02</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.78E-01</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.160E-03</td>
<td>0.00E-00</td>
<td>0.00E-00</td>
<td>-0.16E-00</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
FIT W/O ERRORS

J= 1 X(J)= 0.44E+06 Y(J)= -0.134E-02 ALPHA(J)= 0.0000
J= 2 X(J)= 0.258E+06 Y(J)= -0.107E-02 ALPHA(J)= 0.0000
J= 3 X(J)= 0.258E+06 Y(J)= -0.107E-02 ALPHA(J)= 0.0000
J= 4 X(J)= 0.112E+06 Y(J)= -0.65E-03 ALPHA(J)= 0.0000
J= 5 X(J)= 0.657E+05 Y(J)= -0.52E-03 ALPHA(J)= 0.0000
J= 6 X(J)= 0.154E+05 Y(J)= -0.38E-03 ALPHA(J)= 0.0000
Y INTERCEPT= -0.49529E-03 SLOPE= -0.23243E-08
SIGMA B= 0.56996E-04 SIGMA M= 0.24341E-09
R= -0.9727
K= 0.174E+06

J= 1 X(J)= 0.459E+06 Y(J)= -0.134E-02 ALPHA(J)= 0.2754
J= 2 X(J)= 0.259E+06 Y(J)= -0.107E-02 ALPHA(J)= 0.4025
J= 3 X(J)= 0.259E+06 Y(J)= -0.107E-02 ALPHA(J)= 0.4025
J= 4 X(J)= 0.123E+06 Y(J)= -0.68E-03 ALPHA(J)= 0.6681
J= 5 X(J)= 0.679E+05 Y(J)= -0.53E-03 ALPHA(J)= 0.7198
J= 6 X(J)= 0.168E+05 Y(J)= -0.38E-03 ALPHA(J)= 0.9123
Y INTERCEPT= -0.40473E-03 SLOPE= -0.22509E-08
SIGMA R= 0.57385E-04 SIGMA M= 0.22799E-09
R= -0.9727
K= 0.180E+06

J= 1 X(J)= 0.459E+06 Y(J)= -0.134E-02 ALPHA(J)= 0.2814
J= 2 X(J)= 0.259E+06 Y(J)= -0.107E-02 ALPHA(J)= 0.4097
J= 3 X(J)= 0.259E+06 Y(J)= -0.107E-02 ALPHA(J)= 0.4097
J= 4 X(J)= 0.123E+06 Y(J)= -0.69E-03 ALPHA(J)= 0.5924
J= 5 X(J)= 0.679E+05 Y(J)= -0.53E-03 ALPHA(J)= 0.7259
J= 6 X(J)= 0.168E+05 Y(J)= -0.38E-03 ALPHA(J)= 0.9147
Y INTERCEPT= -0.40479E-03 SLOPE= -0.22492E-08
SIGMA R= 0.57389E-04 SIGMA M= 0.22855E-09
R= -0.9727
K= 0.180E+06
TYPE -CR- TO FIT W/ ERRORS; -1-, -CR- TO SKIP

FIT W/ ERRORS

J= 1 \( X(J)= 0.458E+06 \) \( Y(J)= -0.134E-02 \) ALPHAC(J)= 0.2816
J= 2 \( X(J)= 0.259E+06 \) \( Y(J)= -0.107E-02 \) ALPHAC(J)= 0.4099
J= 3 \( X(J)= 0.259E+06 \) \( Y(J)= -0.107E-02 \) ALPHAC(J)= 0.4099
J= 4 \( X(J)= 0.123E+06 \) \( Y(J)= -0.633E-03 \) ALPHAC(J)= 0.5926
J= 5 \( X(J)= 0.672E+05 \) \( Y(J)= -0.518E-03 \) ALPHAC(J)= 0.7261
J= 6 \( X(J)= 0.156E+05 \) \( Y(J)= -0.331E-03 \) ALPHAC(J)= 0.9147
Y INTERCEPT= -0.354E+03 SLOPE= -0.259E-03 SIGMA B= 0.529E-04 SIGMA M= 0.426E-09
R= -0.99874
K= 0.138E+06

J= 1 \( X(J)= 0.457E+06 \) \( Y(J)= -0.134E-02 \) ALPHAC(J)= 0.2326
J= 2 \( X(J)= 0.258E+06 \) \( Y(J)= -0.107E-02 \) ALPHAC(J)= 0.3493
J= 3 \( X(J)= 0.258E+06 \) \( Y(J)= -0.107E-02 \) ALPHAC(J)= 0.3493
J= 4 \( X(J)= 0.122E+06 \) \( Y(J)= -0.633E-03 \) ALPHAC(J)= 0.5299
J= 5 \( X(J)= 0.677E+05 \) \( Y(J)= -0.518E-03 \) ALPHAC(J)= 0.6713
J= 6 \( X(J)= 0.166E+05 \) \( Y(J)= -0.331E-03 \) ALPHAC(J)= 0.8219
Y INTERCEPT= -0.3504E-03 SLOPE= -0.2609SE-03 SIGMA B= 0.535E-04 SIGMA M= 0.466E-09
R= -0.98974
K= 0.124E+06

J= 1 \( X(J)= 0.456E+06 \) \( Y(J)= -0.134E-02 \) ALPHAC(J)= 0.2274
J= 2 \( X(J)= 0.258E+06 \) \( Y(J)= -0.107E-02 \) ALPHAC(J)= 0.3427
J= 3 \( X(J)= 0.258E+06 \) \( Y(J)= -0.107E-02 \) ALPHAC(J)= 0.3427
J= 4 \( X(J)= 0.122E+06 \) \( Y(J)= -0.633E-03 \) ALPHAC(J)= 0.5226
J= 5 \( X(J)= 0.677E+05 \) \( Y(J)= -0.518E-03 \) ALPHAC(J)= 0.6648
J= 6 \( X(J)= 0.166E+05 \) \( Y(J)= -0.331E-03 \) ALPHAC(J)= 0.8690
Y INTERCEPT= -0.3492E-03 SLOPE= -0.2617E-03 SIGMA B= 0.5295E-04 SIGMA M= 0.4803E-09
R= -0.98994
K= 0.134E+06
\[ J = 1 \quad X(J) = 0.45626E+06 \quad Y(J) = -0.134E-02 \quad \text{ALPHA}(J) = 0.2265 \]
\[ J = 2 \quad X(J) = 0.23754E+06 \quad Y(J) = -0.107E-02 \quad \text{ALPHA}(J) = 0.3416 \]
\[ J = 3 \quad X(J) = 0.25754E+06 \quad Y(J) = -0.107E-02 \quad \text{ALPHA}(J) = 0.5416 \]
\[ J = 4 \quad X(J) = 0.13265E+06 \quad Y(J) = -0.631E-03 \quad \text{ALPHA}(J) = 0.5213 \]
\[ J = 5 \quad X(J) = 0.57717E+05 \quad Y(J) = -0.531E-03 \quad \text{ALPHA}(J) = 0.6636 \]
\[ J = 6 \quad X(J) = 0.13764E+05 \quad Y(J) = -0.351E-03 \quad \text{ALPHA}(J) = 0.8835 \]

FINAL K = 0.1336E-06 FINAL DELTA EPSILON = -0.286E+04
SIGMA K = 0.3205E+05 SIGMA DELTA EPSILON = 0.413E+03

\[ J = 1 \quad DX(J) = 0.395E+05 \quad DY(J) = 0.22E-03 \quad \text{[COMPLEX]} = 0.0736E-05 \]
\[ J = 2 \quad DX(J) = 0.158E+05 \quad DY(J) = 0.20E-03 \quad \text{[COMPLEX]} = 0.146E-04 \]
\[ J = 3 \quad DX(J) = 0.158E+05 \quad DY(J) = 0.20E-03 \quad \text{[COMPLEX]} = 0.146E-04 \]
\[ J = 4 \quad DX(J) = 0.687E+04 \quad DY(J) = 0.10E-03 \quad \text{[COMPLEX]} = 0.223E-04 \]
\[ J = 5 \quad DX(J) = 0.317E+04 \quad DY(J) = 0.75E-04 \quad \text{[COMPLEX]} = 0.285E-04 \]
\[ J = 6 \quad DX(J) = 0.634E+03 \quad DY(J) = 0.39E-04 \quad \text{[COMPLEX]} = 0.332E-04 \]

PLOT DATA? (0=NO,1=YES) 1

NORMALIZER TO MAKE X AXIS INTEGRAL = 1.E+05
NORMALIZER TO MAKE Y AXIS INTEGRAL = 1.E-04

\[ I = 1 \quad \text{NORMALIZED X(I)} = 4.562 \quad \text{NORMALIZED Y(I)} = -13.437 \]
\[ I = 2 \quad \text{NORMALIZED X(I)} = 2.575 \quad \text{NORMALIZED Y(I)} = -10.750 \]
\[ I = 3 \quad \text{NORMALIZED X(I)} = 2.575 \quad \text{NORMALIZED Y(I)} = -10.750 \]
\[ I = 4 \quad \text{NORMALIZED X(I)} = 1.227 \quad \text{NORMALIZED Y(I)} = -6.825 \]
\[ I = 5 \quad \text{NORMALIZED X(I)} = 0.677 \quad \text{NORMALIZED Y(I)} = -5.375 \]
\[ I = 6 \quad \text{NORMALIZED X(I)} = 0.168 \quad \text{NORMALIZED Y(I)} = -2.905 \]

ALL OK? 1

NEW GRAPH? (0=NO,1=YES) 1
NEW PARAMETERS? 1
XMIN=0, YST=XMIN
XMAX=5, YMAX=0, YST=YMIN
YMIN=-16, YINC=8
XTIC=0, YTIC=4
SYMBOL # 1
SYMBOL SIZE 2
CALCOMP(0) OR TEKTRONIX(1)

ALL OK? 1

PLOT IS BOUND FOR CALCOMP

PLOT EVERY NTH POINT, N = 1
RE-FIRST? 1

1:1(#1), 2:1(#2), OR 1:1:1(#3) COMPLEX?
2. Equilibrium Concentrations/$\Delta \varepsilon_{\text{bound}}$

A) Description

The program EQ calculates the equilibrium concentration of dimer(s) and dye for 1:1, 2:1, or 1:1:1 stoichiometries and also calculates $\Delta \varepsilon^\lambda_{\text{bound}}$ and its associated error. The program is written in FORTRAN, compiled with .R FORT, and saved as a core image file with .SAVE SYS EQ. All listings are available on paper tape.

The data consists of the complex's equilibrium constant, the initial concentrations of dimer(s) and dye, and their errors for the calculation of the equilibrium concentrations. To calculate $\Delta \varepsilon_{\text{bound}}$, enter the measured induced CD ($\varepsilon^0$ x 100) and its error. Equations (2) and (6) of Chapter II are used in the calculations.
B) Listing 

i) EQ

COMMON WAVE, TH100, ERTH, NHAVE, CN1, CN2, CB, PLEX,
B, K, RK, ALPHA, ERPLX, EC1, EC2, CSUM, SUM
DIMENSION WAVE(6), TH100(6), ERTH(6)

700 FORMAT(' THIS PROGRAM CALCULATES EQUILIBRIUM CONCENTRATIONS, ')
710 FORMAT(' CD DELTA EPSILONS, AND ERRORS FOR EACH. TO CALCULATE ')
720 FORMAT(' EQUILIBRIUM VALUES ALONE, ENTER 2 FOR EACH ERROR AND ')
730 FORMAT(' NUMBER OF WAVELENGTHS. ')
740 FORMAT(')
750 FORMAT('1:1(=1), 2:1(=2), OR 1:1:1(=3) COMPLEX? ')
760 FORMAT(' EQUILIBRIUM CONSTANT (E10.4) = 'E10.4)
770 FORMAT(' ERROR K (E11.5) = 'E11.5)
780 FORMAT(' INITIAL [DYE] (E10.4) = 'E10.4)
790 FORMAT(' ERROR [DYE] (E11.5) = 'E11.5)
800 FORMAT(' INITIAL [DIMER 1] (E10.4) = 'E10.4)
810 FORMAT(' ERROR [DIMER 1] (E11.5) = 'E11.5)
820 FORMAT(' INITIAL [DIMER 2] (E10.4) = 'E10.4)
830 FORMAT(' ERROR [DIMER 2] (E11.5) = 'E11.5)
840 FORMAT(' PATH LENGTH (CM) = 'F7.4)
850 FORMAT(' NUMBER OF DIFFERENT WAVELENGTHS (11) = '11)
860 FORMAT(' WAVELENGTH (F5.1) = 'F5.1)
870 FORMAT(' THETA X 100 (E11.4) = 'E11.4)
880 FORMAT(' ERROR IN THETA X 100 (E12.5) = 'E12.5)
890 FORMAT(' NO CONVERGENCE OF ALPHA AFTER 50 STEPS ')
900 FORMAT(' K = 'E10.4)
910 FORMAT(' COMPLEX: ')
920 FORMAT(' INITIAL = 'E10.4, ERROR = 'E11.5)
930 FORMAT(' INITIAL = 'E10.4, FINAL = 'E11.5)
940 FORMAT(' ALL ERRORS USED ')
950 FORMAT(' CONVERGENCE OF DYE, THETA ERRORS ZEROED ')
960 FORMAT(' DYE, THETA ERRORS ZERED ')
970 FORMAT(' NEW [DYE] OR SYSTEM? (0=NO, 1=YES) ')

C
WRITE(1,700)
WRITE(1,710)
WRITE(1,720)
WRITE(1,730)
10 CN2=0
EC2=0
WRITE(1,740)
11 RK=1.
READ(1,750)K
IF(K<2)10,15,11
12 IF(K<3)10,15,10
13 RK=2.
14
C ENTER CONCENTRATION DATA

15  READ(1,800)CA, 
    READ(1,810)ERCA, 
    READ(1,820)C0, 
    READ(1,830)ERC 
17  READ(1,840)CN1, 
    IF(K-2)=25,20.25 
20  READ(1,850)CN2, 
25  READ(1,860)EC1, 
    IF(K-2)=35,30.25 
30  READ(1,870)EC2

C ENTER CD DATA

35  READ(1,875)D, 
    READ(1,880)NWEAVE, 
    DO 40 I=1,NWEAVE 
    WRITE(1,740) 
    READ(1,885)WAVE(I), 
    READ(1,890)TH100(I), 
    READ(1,895)ERTH(I) 
40  CONTINUE

C CALCULATE EQUILIBRIUM CONCENTRATIONS

C
ALPHA=0
CSUM=(ERCAY/C0)**2.+(ERC/C0)**2.
DO 50 J=1,50 
    CALL FREE (FRE1,FRE2,YALNEW) 
    ALF=CA-YALNEW 
    ALFA=ALF/(1.0*ALF) 
    DIFF=ABS(ALFA-ALPHA) 
    ALPHA=ALFA 
    PLEX=C0*ALPHA 
    CALL FREE (FRE1,FRE2,YALNEW) 
    IF(DIFF+.001*ALPHA)60,60,50 
50  CONTINUE
    WRITE(1,900) 
    GO TO 99

C PRINT EQUILIBRIUM CONCENTRATIONS

60  ERPLX=SQRT(PLEX*PLEX*SUM) 
    WRITE(1,910)CA, 
    WRITE(1,920)PLEX,ERPLX 
    WRITE(1,930)C0,C,ERC 
    WRITE(1,940)1, CN1,FRE1,EC1 
    IF(K-2)=70,65,70 
65  WRITE(1,945)2, CN2,FRE2,EC2 
70  IF(NWAVE)39,39,75
C CALCULATE DELTA EPSILON, IF DESIRED
C
75 WRITE(1,950)
    CALL THETA
    DO 80 I=1,NWAVE
        ERTH(I)=3
    80 CONTINUE
    WRITE(1,960)
    CALL THETA
    ERPLX=ABS(PLEX*ERCAY/CAY)
    WRITE(1,970)
    CALL THETA
79 WRITE(1,740)
    READ(1,980)I
    WRITE(1,740)
    IF(I) 10,17,10
END
ii) Subroutine THETA

```fortran
SUBROUTINE THETA
COMMON WAVE, TH100, ERTH, NAVE; CN1, CN2, C0, PLEX,
B, K, RK, ALPHA, ERPLX, EC1, EC2, CSUM, SUM
DIMENSION WAVE(6), TH100(6), ERTH(6)
C FORMAT(F5.1* NM, THETA * 100 = 'E11.4' ERROR = 'E12.5')
C FORMAT(F5.1* NM, DELTA EPSILON = 'E10.3' ERROR = 'E11.4')
DO 10 I=1, NAVE
   DELEP = TH100(I)/(3238. * PLEX) + B
   SUM = ERTH(I)/ABS(TH100(I)) +2. + (ERPLX/PLEX)**2.
   ERDEL = SQRT(DELEP + DELEP + SUM)
   WRITE(1, 710) WAVE(I), TH100(I), ERTH(I)
   WRITE(1, 720) WAVE(I), DELEP, ERDEL
10 CONTINUE
RETURN
END
```

iii) Subroutine FREE

```fortran
SUBROUTINE FREE (FRE1, FRE2, VALNEW)
COMMON WAVE, TH100, ERTH, NAVE; CN1, CN2, C0, PLEX,
B, K, RK, ALPHA, ERPLX, EC1, EC2, CSUM, SUM
DIMENSION WAVE(6), TH100(6), ERTH(6)
C FRE1 = CN1 - RK * ALPHA + C0
FRE2 = CN2 - RK * ALPHA + C0
IF (K-2)52, 54, 56
52 VALNEW = FRE1
   SUM = CSUM + (RK * EC1/CN1)**2.
   FRE2 = 1.
   RETURN
54 VALNEW = FRE1
   SUM = CSUM + (RK * EC1/CN1)**2.
   FRE2 = 1.
   RETURN
56 VALNEW = FRE1 + FRE2
   SUM = CSUM + (RK * EC1/CN1)**2. + (RK * EC2/CN2)**2.
   RETURN
END
```
C) Sample Run

R EQ

THIS PROGRAM CALCULATES EQUILIBRIUM CONCENTRATIONS, CD DELTA EPSILONS, AND ERRORS FOR EACH. TO CALCULATE EQUILIBRIUM VALUES ALONE, ENTER 0 FOR EACH ERROR AND NUMBER OF WAVELENGTHS.

1:1<1> = 1), 2:1<2> = 2), OR 1:1:1<3> = 3) COMPLEX? 2
EQUILIBRIUM CONSTANT (E10.4) = 1.2E+05
ERROR K <E11.5> = .3E+05
INITIAL (DYE) <E10.4> = .04E-03
ERROR (DYE) <E11.5> = .002E-03
INITIAL (DIMER 1) <E10.4> = 2.9E-03
ERROR (DIMER 1) <E11.5> = .01E-03
PATH LENGTH <CM> = 1.
NUMBER OF DIFFERENT WAVELENGTHS <11> = 1

WAVELENGTH <F 5.1> = 305.
THETA X 100 <E11.4> = .712
ERROR IN THETA X 100 <E12.5> = .014

K = 0.1390E+06

[COMPLEX].

FINAL = 0.22128E-04 ERROR = 0.52293E-05
(DYE): INITIAL = 0.4760E-04 FINAL = 0.20372E-04 ERROR = 0.20000E-05
(DIMER): INITIAL = 0.2990E-03 FINAL = 0.20357E-02 ERROR = 0.16000E-04

ALL ERRORS USED

305.0 NM: THETA X 100 = 0.7120E-00 ERROR = 0.24000E-01
305.0 NM: DELTA EPSILON = 0.976E-01 ERROR = 0.2352E+01

THETA ERRORS ZEROED

305.0 NM: THETA X 100 = 0.7120E-00 ERROR = 0.24000E-00
305.0 NM: DELTA EPSILON = 0.976E+01 ERROR = 0.2306E+01

DIMER, DYE, THETA ERRORS ZEROED

305.0 NM: THETA X 100 = 0.7120E-00 ERROR = 0.20000E-00
305.0 NM: DELTA EPSILON = 0.976E+01 ERROR = 0.2251E+01

NEW (DYE) OR SYSTEM? (0=NO, NZ=YES) 1

1:1<1> = 1), 2:1<2> = 2), OR 1:1:1<3> = 3) COMPLEX?
Appendix B

SUPER SPECTRUM DATA SYSTEM

The Super Spectrum system of programs is a lineal descendant of the system described by Tomlinson (1968) and is used to acquire and process data from the Cary 60 and Cary 118 spectrometers with a PDP 8/E computer (Digital) and a RK-8E disk drive. The updates and corrections to the program have been extensive; the main differences are modification of the pen averaging routine to use the exact "stable averaging" algorithm (Savitsky & Golay, 1964), the addition of plotting routines, and the creation of an overlay to transmit data to the Lawrence Berkeley Lab computer system.

Complete operating instructions and listings of the programs are on the enclosed microfiche. All programs are stored on punched card decks and on GSS tape 10515 at LBL. A handbook describing operation of the system and assembly of the programs has been prepared from the OPERATE program - this is available in the lab.
1. Introduction

The programs PREPARE, SMOOTHS, and PROCESS translate, average, smooth, and perform other manipulations with spectra from Super Spectrum. The programs are designed to run in succession: the output from one is the input to the next. These programs are descended from the programs described in Borer (1972). More information on the ID requirements and formats of the spectra for proper operation in this system is listed in the OPERATE program of Super Spectrum (Appendix B).

2. Program PREPARE

A) Operation

i) Input

The hexadecimal spectra files on PSS created by the transmission overlay from Super Spectrum.

ii) Output

The file OUTPUT contains a summary of the data translation. Misread lines are listed here. This is usually placed in the HOLDOUT queue for immediate viewing.

The file TAPE10 contains intermediate listings and plots of the data. This is usually deleted, but may be DISPOSEd to microfiche, if necessary.

The file TAPE30 contains the translated spectra in a form acceptable for SMOOTHS. This file is carried over to the next program when the two programs are run in tandem (below).
iii) Errors

Stray or missing bits in the PSS file will always prevent SMOOTH from starting after translation of all files is completed (if possible). The line in error will be listed as read with an error message on the file OUTPUT. The original listing of hexadecimal lines will contain the correct information for comparison. The entire data file may not be read if there is an error in the parameter line (1st line) of a new spectrum; correction of this error is necessary before the rest of the file will be read.

B) Listing

i) Program PREPARE

ii) Subroutine UNLOAD

iii) Function UNLOAD

iv) Subroutine REDO

v) Subroutine PRNPLT

vi) Subroutine PLSCAL
PROGRAM PREPAPE (INPUT, OUTPUT, TAPE13, TAPE5 INPUT, TAPE10, TAPE30)

C*** THIS VERSION OF -PREPAPE- IS COMPATIBLE WITH THE LATEST
C*** UPDATE OF -SUPER SPECTRUM- ONLY (1980). NO PREVIOUS LIST-
C*** THIS IN THIS VERSION TAPE IS SC SSTRUCUTED.
C*** THIS PROGRAM READS DATA OUT OF PSS AND TRANSLATES IT FROM HEXA-
C*** SPECTRAL RULES OF DATA WAS READ IN THROUGH MARE I OVERLAY (NUMBER 2) OF
C*** SPECTRUM AND STORED AS HEXADECIMAL CHARACTERS; THREE CHARAC-
C*** TERS/WORD, 25 WORDS/LINE, PLUS CHECKSUM. TAPE5 FILE CONTAINS THE
C*** DATA OUT OF PSS. TAPE10 FILE WILL CONTAIN PRELIMINARY TRANSMIS-
C*** SION OF THE DATA, AND WILL BE FURTHER USED IN THE PROGRAM SMOOTH-
C*** TAPE30 FILE IS THE TRANSLATED DATA AND WILL ALSO GO TO THE PROGRAM
C*** SMOOTH. TAPE30 CAN ALSO BE STORED ON TAPE ANY ERRORS IN TRANS-
C*** MISSION HALT WRITING ON TAPE10 AND TAPE30 AND DELETE THEM. TAPE13
C*** CONTAINS THE FILE CONTROL AND IS REWRITTEN IF TRANSMISSION ERRORS
C*** OCCUR TO halt THE PROGRAM SEQUENCE, A TRANSMISSION SUMMARY IS
C*** PRINTED ON THE FILE OUTPUT, ALONG WITH ANY ERROR MESSAGES.

COMMON/CD/CD(300)
COMMON/DAWA/DAWA(300)
COMMON/IDATA/IDATA(300)
COMMON/IDATA/IDATA(300)
COMMON/SUM/SUM(30)

REAL CD,DAWA
INTEGER IDATA, SUM
DIMENSION PARAM(6)

3* SMOOTH DATA *A3)
701 FORMAT(3A3)
3* =, 5X, *SCALE=, 3X, *NOPTS /, 64X, *X(0,001), /*)
710 FORMAT(*, *INTER =, 10, * SPECTRA READ FROM FILE*)
740 FORMAT(I2, 63)
741 FORMAT(*, 14, 4X, 12, 1X, *START =, f7.2, 1X, *NM =, 12, 1X, *END =, f7.2, 
3* =, NM, INCREMENTING BY 1, F7.2, * NM, 1X, DATE FACT =, f9.4, 3*
3* =, EPSILON =, f9.4, 3*, SCALE(1,001) =, f9.4, 3*, 1X, 63,
3* =, DATA POINTS /, /, (1H, 12404, 1X, 041)
742 FORMAT(*, 14, 4, 12, 1X, 3, f9.2, 2X, 1, f10, 2, 2X, 1, f10, 2, 2X, 1, /)
743 FORMAT(14, 3, 2, 3, f10, 2, 1, f10, 2, 2, 1, f10, 2, 2, 1, f10, 2, 2, 1, /)
744 FORMAT(14, 4, 2, 3, f10, 2, 2, 1, f10, 2, 2, 1, f10, 2, 2, 1, /)
745 FORMAT(14)
746 FORMAT(*)
747 FORMAT(*, *UNPACKING CF DATA HALTED; UNABLE TO SET NUMPTS*)
748 FORMAT(*, 14, 4, 12, 1X, 3, f9.2, 2X, 1, f10, 2, 2X, 1, f10, 2, 2X, 1, f10, 2, 2X, 1, /)
749 FORMAT(14, 2)
750 FORMAT(*, *ONE OR MORE TRANSMISSION ERRORS HAVE OCCURRED AND MUST 
3* BE CORRECTED, BEFORE COMPLETE WORKUP OF DATA IS POSSIBLE*)
REIND IC
REIND 3C
READ, 701 IUPNPR
PRINT 700, 1UPNPR
WRITE(701, 701) IUPNPR
PRINT 702
ANSCI=0
ITREV=0
ITRNS=0

C++++ TRANSLATE SPECTRUM CONTROLS

READ(5,740) (I0ATA(I), I=1,25), I5UM(I)
IF(I0ATA(5).NE.C) GO TO 49
WRITE(11,740)
CALL ULOAD(1,1,ITRNS)
CO 413 I=1,7
(HEAD(1)=10ATA(1),AND,178).

410 CONTINUE
ID1=HEAD(I)*100+HEAD(2)*100+HEAD(3)*10+HEAD(4)
CO 411 I=5,8
INT=IDCODE(IDATA(1))
(HEAD(I)=INT)

411 CONTINUE
I02P=HEAD(5)*100+HEAD(6)
I03P=HEAD(7)*100+HEAD(8)
LECODE(I)=749,102P IGA,102
LECODE(10,103) IGA,103
PARA(1)=FLOAT(I0ATA(11))
PARA(2)=FLOAT(I0ATA(12))
PARA(3)=FLOAT(I1,12,10),AND,7777811/10.
NPTS=1,4 (1,12,101,102,103,PARA,1PTS,DATA1,1PTS)
CC 43 I=1,3
J=2*(4+1)
IFEXP(I)=I0ATA(J-1)
IFAM(I)=I0ATA(J)*10000+I0ATA(J+1)
IF(IEXP(I).AND.400000.EQ.0) GO TO 42
IFEXP(I)=-11(I,12,101,EXP(I),AND,777781)

42 CONTINUE
IF(SPECT.T.EQ.,1,AND,ITRNS,2,0) NUMPTS=NPTS
IF(SPECT.T.EQ.,1,AND,ITRNS,3,0,AND,NUMPTS,NE,NPTS) GC TO 48
IF(SPECT.T.EQ.,1,AND,ITRNS,3,0,AND,NUMPTS,NE,NPTS) GC TO 48

C++++ TRANSLATE SPECTRUM POINTS

READ(5,740) ((I0ATA(J+1-11),I=1,25),(SUM(J/25+21),J=1),NUMPTS,152)
LACNT=(NUMPTS+25)/25+1
CALL ULOAD (2,1ACNT,ITRNS)
SPECT=SPECT+1
IF(1TRNS,NE,C) GO TO 430
PRINT 748,101,102,103,(PARA(I),I=1,6),NPTS

43G ITREF=ITREF+ITRNS
IF(ITREF,NE,0) GC TO 4G

C+++ TAPE* WILL GC TO MICROFICHIE WITH TRANSMISSION DATA, RAW DATA
C+++ TABLES, AND PLOTS
WRITE(10,741) ID1,102,103,PARA,NPTS,(I0ATA(I),I=1,NUMPTS)
CC 44 I=1,NPTS
WRITE(I0,741) ID1,102,103,PARA,NPTS,(I0ATA(I),I=1,NUMPTS)
CC 44 I=1,NPTS
IF(I0ATA(I).EQ.,1,AND,400000.EQ.0) GO TO 431
I0ATA(I)=I0ATA(I)*777777777777777

431 CONTINUE
WRITE(10,742) ID1,102,103,PARA,NPTS
11=NPTS/3+1
CC 45 I=1, II
J=J+1
K=J+1

WRITE(10,743) HAVE(1), CD(1), HAVE(J), CD(J), HAVE(K), CD(K)
CONTINUE
WRITE(10,742) 101, 102, 103, PARAM, NPTS

I0E=10
YMAX=PARAM(11)
YINC=(PARAM(11)-PARAM(2))/100.

TWICE=YINC/0.5

M=TWICE
IF(MEEE, TWICE) XINC=0.5*(M+1)

CALL PRTSCT(WAVE13,CC(1),XFAK,XINC,YMAX,YINC,SY,NPTS,DEV)

TAP30 WILL GC TO SMOOTHING PROGRAM AND TO STORAGE ON TAPE
WRITE(22,744) 101, 102, 103, PARAM, NPTS
WRITE(10,745) (ICATA(1),I=1,NPTS)

CC TO 40

49 PRINT 747

CALL REDO

49 PRINT 718,NSPECT

WRITE(10,733) [PUNPR

WRITE(10,718) NSPECT

ENDFILE 30

IF TRER.EQ.0 CALL EXIT

PRINT 750

CALL REDO

END
SUBROUTINE UNLOAD (LCLIM, LNCT, INTRNS)

C*** CONVERTS LINES OF HEXADECIMAL DATA (126 WORDS/LINE, 78 CHARACTERS)
C*** TO BINARY FORM
C*** LAST WORD (3 CHARACTERS) IN EACH LINE IS CHECKSUM
C
INTEGER NDATA, NSUM

751 FORMAT*,8X,CONTROL LINE OF SPECTRUM *,BRI,
$*, OCTAL ERROR IN NTOT= **,C4)
752 FORMAT*,8X,DATA LINE NUMBER *,12,* OF SPECTRUM *,BRI,
$, OCTAL ERROR IN NTOT= *,04)
753 FORMAT*,8X,*ERROR IN DATA TRANSMISSION*,/9X,
$*ERROR IN DATA TRANSMISSION*1
754 FORMAT*,8X,25(R3,LX),LX,R3)
INDEX=0
DC 57 I=LHLIM, LNCT

NTOT=0
DO 51 J=1,25
INDEX=INDEX+1
C*** SAVE LINE FOR POSSIBLE ERROR PRINTING
LODAT(J)=DATA(INDEX)
N(I)=NDATA(INDEX) AND 7778
N(1)=NDATA(INDEX) AND 7700/1000B
N(3)=NDATA(INDEX) AND 7700B/10000B
C*** DISPLAY MODE HEXADECIMAL TO BINARY CONVERSION
C*** DISPLAY MODE REPRESENTS D-9=33-448, A-F=61-068
DO 50 K=1,3
IF (N(K),AD,70B),EQ,01 N(K)=N(K)+448
N(K)=N(K)-338
CONTINUE
NTOT+NTOT+N(I)*K(2)+N(3)
NDATA(INDEX)=N(I)+16*K(2)+256*N(3)
51 CONTINUE
C*** CHECK LINE TRANSMISSION
N(I)=NSUM(I) AND 7778
N(1)=NSUM(I) AND 7700/1000B
N(3)=NSUM(I) AND 7700B/10000B
DC 52 J=1,3
IF (N(J),AND,70B),EQ,01 N(J)=N(J)+448
N(J)=N(J)-338
CONTINUE
NTOT+NTOT=N(I)+16*N(1)+256*N(3)
IF (LOLIM,NE,1) GO TO 54
C*** SAVE ID FOR POSSIBLE ERROR PRINTING
DO 53 J=1,8
NHEAD(J)=IC0DE(NDATA(J))
CONTINUE
54 IF (NCT.EQ,0) GO TO 57
C*** NOTE ERRORS THAT OCCURRED IN TRANSMISSION
NTOT=INTRANS+1
IF (LOLIM,NE,1) GO TO 55
PRINT 753
PRINT 751,NHEAD, NTOT
PRINT 754,(LODAT(I),L=1,25),NSUM(I)
GO TO 57
55 PRINT 753
PRINT 752,I-1,NHEAD, NTOT
PRINT 754,(LODAT(I),L=1,25),NSUM(I)
CONTINUE
RETURN
END
FUNCTION ICODE(ITY)
CHARACTER IN 8-BIT ASCII CODE TO 6-BIT DISPLAY CODE CONVERSION

DIMENSION ISPEC(27)

DATA (SPEC(I), I=1,27) /538,558,528,578,508,518,448,758,408,548,
568,348,338,158,728,4*438,368,2*408,768,3*438,738/
ITY=ITY, AND, 00778
IF (ITY .GE. 33) GO TO 60
ICODE=ITY
RETURN

60 IF (ITY .LT. 608, CODE. ITY .GT. 718) GO TO 61
ICODE=ITY-258
RETURN

61 GO 62 I=1,27
62 CONTINUE
RETURN
63 ICODE=I+448
RETURN
END

SUBROUTINE END

** WRITING THE CONTROL FILE TO END RUN **
801 FORMAT(*DELETE,TAPE1,TAPE30,*)
802 FORMAT(*DELETE,TAPE30,*)
803 FORMAT(*DELETE,TAPE30,*)
REWIND 13
WRITE(13,801)
WRITE(13,802)
REWIND 13
CALL EXIT
END
**SUPERCLINE PSCALE (V,VMAX,VINCR,IPTS,NDIVIS,DMAX,DMIN,ISCLER)**

**CC*** SCALING ROUTINE FOR PSCALE M.J. ITZKOVITZ MAY 1967

**CC*** TO BE AN INTEGRAL MULTIPLE OF 5.0*VINCR

**DIMENSION RLI(11),VIN(PNTS)**

**DATA (FLIM111,1,1,1,1,1,1,25,1,60,2,70,2,50,3,20,4,CC,5,00,8,40,**

**1,CC,1,CC,1,CC)**

**57** **FUNCTION** **= ,** **FULL** **SCALE** **CALLED** **TO** **SCALE** **ARRAY** **WITH** **ZERO** **RANGE***

**VMIN=V(1)**

**VMAX=V(1)**

**CC RC I=1,PNTS**

**IF(VI111, LT, VMIN) VMIN=V(I)**

**IF(VI111, G1, VMAX) VMAX=V(I)**

**80** **CONTINUE**

**CRANGE=VMAX-VMIN**

**IF(CRANGE, LE, 0) GO TO 91**

**CRANGE=Aalog(CRANGE)/Aalog0.01**

**IF(CRANGE, LE, 0) GO TO 92**

**CRANGE=CRANGE**

**GO TO 83**

**92** **CRANGE=-CRANGE**

**IPRANGE=IPRANGE-1**

**93** **CRANGE=CRANGE-FLOAT(CRANGE)**

**RANGE=IPRANGE**

**GO TO 79**

**C*** **PANCE** **IS** **BETWEEN** **1.0** **AND** **10.0**

**CC B4 I=1,9**

**IF(IPRANGE, LE, FLIM111) GO TO 84**

**RANGE=FLIM111**

**ILIM=1**

**GO TO 85**

**84** **CONTINUE**

**RANGE=10.0**

**ILIM=10**

**85** **TRANSE=ANGE=11.0**

**VINCR=TRANSE/FLOAT(NDIVIS)**

**IF(WX, LE, 0) GO TO 87**

**TX=VX/5.0*VINCR**

**VX=5.0*VINCR**

**GO TO 88**

**87** **VMAX=VMAX/5.0*VINCR**

**X*X=5.0*VINCR**

**GO TO 90**

**88** **IF(VMIN, GE, VXMAX-TRANSE) GO TO 90**

**ILIM=ILIM+11**

**ILIM=ILIM+1**

**RANGE=FLIM(1)ILIM**

**IF(IPRANGE, LE, 11, LT, 1) GO TO 85**

**RANGE=PANCE/10.0**

**IPRANGE=IPRANGE+1**

**GO TO 95**

**90** **EWMAX=VMAX**

**CMIN=VMIN**

**VXMAX=XMAX**

**RETURN**

**91** **PRINT 927**

**ISCLER=1**

**RETURN**

**END**
3. Program SMOOTH

A) Operation

i) Input

The translated spectra on the TAPE30 file from PREPARE.

ii) Output

The file TAPE10 contains intermediate listings and plots of the data. This is usually deleted, but may be DISPOSEd to microfiche if necessary.

The file OUTPUT (= TAPE7) contains listings and plots of all averaged spectra and difference spectra created with the ID codes ST, SU, US, SM, and MS.

The file PUNCH contains the averaged and smoothed spectra in a form acceptable for PROCESS. This file is punched and the deck serves as input in the next program (see note, below).

iii) Errors

The order of spectra in the file is important. Spectra with AV codes will be averaged with the next in line whether they are in the right order or not. Spectra with ID codes SU, US, SM, and MS require a second spectrum previously prepared by either one of these commands or the ST code. If this spectrum is not present, an error message is left and the next series of spectra are run.

B) Listing

i) Program SMOOTH

ii) Subroutine LETSEE

iii) Subroutine SHIFTS

iv) Subroutine WRAPUP

v) Subroutine PRNPLT and Subroutine PLSCAL (see PREPARE)
**THIS IS THE LATEST VERSION OF THE PROGRAM - SMOOTH**

**is fully compatible with any version of -SUPER SPECTRUM**

**THIS PROGRAM READS THE TRANSLATED SPECTRA FROM THE PROGRAM PREPARE**

**AV weekend, and calculates difference spectra. A punched deck of**

**other programs, the tapeio file is carried over from prepares and**

**will contain the lists and plots of AV type data. Tapeio should**

**be disposed to microfiche, and the tape file should be disposed**

**to the printer.**

**INPUT PRINTE MEANINGS**

(1) = YES, compute baseline shifts; NO, disregard sh commands

(2) = YES, punch data to be averaged; NO, punch results only

(3) = YES, smooth the data; AC, AC smoothing

**CONTROL MEANINGS**

(1) is maximum wavelength in 'NM'; (2) is minimum wavelength in 'NM';

(3) is AM per point; (4) is D factor; (5) is E factor, and (6) is

**SCALE TXG, GOI**

**USE OF AV, ST, SV, MS, SU, AND US COMMANDS**

**INPUT PRINTE SETTINGS**

(1) = YES, 2 = NC, 3 = YES.

**CONSIDER THE FOLLOWING SEQUENCE OF SPECTRA**

1001AVM, 1002STM, 1003AVM, 1004SMO2, WHERE NN IS ANY TWO LETTERS.

**IN THIS SEQUENCE, 1001 WILL BE SMOOTHED, LISTED, AND PLOTTED**

**1002 WILL ALSO BE SMOOTHED, LISTED, AND PLOTTED; 1001 AND**

**1003 WILL BE AVERAGED TOGETHER, LISTED, PLOTTED**

**PUNCH, AND THE RESULT STORED AS 1002. 1002 AND 1004 WILL BOTH**

**INDIVIDUALLY BE SMOOTHED, ETC.; 1003 AND 1004 THEN WILL BE**

**AVERAGED TOGETHER, ETC., AND THE RESULT STORED AS 1004. FINALLY,**

**THE DIFFERENCE SPECTRUM (1004-1002) XE/OD WILL BE CALCULATED**

**LISTED, PLOTTED, PUNCH, AND STORED AS 1005RS1T**

**REPLACING SMOUS WILL CALCULATE, ETC.**

**REPLACING SM BY SU WILL CALCULATE, ETC.**

**REPLACING SMOUS WILL CALCULATE, ETC.**

**DO IT AGAIN COMMANDS**

**ASSUME LAST OPERATION WAS 1004SMO2.**

**REPLACE PREVIOUS OPERATION TO 1004SMO2 WILL CALCULATE, ETC.**

**REPLACE PREVIOUS OPERATION TO 1004SM02 WILL CALCULATE, ETC.**

**1004SM02 CALCULATES, ETC.; 1007 = 1004-1002 XE/OD (SM)**

**1004SM02 CALCULATES, ETC.; 1007 = 1002-1004 XE/OD (MS)**

**1004SM02 CALCULATES, ETC.; 1007 = 1004-1002 (SU)**

**1004SM02 CALCULATES, ETC.; 1007 = 1002-1004 (US)**

**BASELINE SHIFT CORRECTIONS**

**100625C WILL CORRECT SPECTRUM 1006 FOR A POSSIBLE PARALLEL-BASE**

**LINE SHIFT AT 250 AM AND BELOW BEFORE SMOOTHING AND STORE RESULT**

**IN-1006 IN FURTHER WORKUP (SMOOTHING, AVERAGING, ETC.). THIS IS**

**TREATED AS AN AV COMMAND.**
COMMON/CD/CD(300)
COMMON/CVSN/CVSN(5), SVCTX(6), NPTS, NPTSSV
COMMON/PRCONF/PRCONF(300)
COMMON/FR(300)
COMMON/AVE/AVE(300)
COMMON/IDATA/IDATA(300)
COMMON/IP/R/IP/R(13)
REAL CO/CONTRAPRCONF/K, SVCTRL, WAVE
INTEGER IDATA/IP/R
DIMENSION TL(13), TL(10)
DIMENSION DIFFER(300), SIGMA(300)
DIMENSION A(50,200,AVSPECT(10,300), CNF ID(50,300)
INTEGER ID(10), SPCT(50), TOTNLM
INTEGER ET(7), SSH, SH, SS, US
INTEGER AVE
DATA AV/2HAV/
DATA MS/2HMS/
DATA SN/2HNS/
DATA ST/2HST/
DATA SU/2HSS/
DATA US/2HUS/
DATA TL(1), I(1,2,10)/12, 76, 4, 303, 3, 182, 2, 776, 2, 447, 2, 165,
  2, 306, 2, 262/
701 FORMAT(183)
702 FORMAT(14*, Z7, A2, Z7, F7.2, 21, G12.6, 13)
703 FORMAT(14*)
704 FORMAT(14*, 14, SMOOTH*)
705 FORMAT(*, 10(X, 5X, I0, 4X, VSTART*, 6X, VEND*, 5X, VINCR, 8X, OD*, 10X, 
  *F*, 10X, SCALE*)
706 FORMAT(*, T14*A2, Z7, 2X3, 31, F7.2, 2X1, 31, F10.3, 2X))
707 FORMAT(*, Z14*, A2, A2, S, 90, 10, 4)
708 FORMAT(*, 14, 14, * IS AVERAGE OF #, 10(14, 2X))
709 FORMAT(14, * SMOOTH*)
710 FORMAT(14, * IS AVERAGE OF #, 11(14, 1X))
711 FORMAT(14)
712 FORMAT(14)
713 FORMAT(*, 14, 14, * IS EQUAL TO (*, 14, * - *, 14, *) X(E = *, F8.4, 
  $4)/(CD = *, F8.4, *)
714 FORMAT(*, 14 = *, 14, * - *, 14, * X(E = *, F8.4, */(OD = *, 
  $8), 14, *))
715 FORMAT(*, 14, * IS EQUAL TO (*, 14, * - *, 14, *))
**C*** initialize parameters

C*** number of spectra to be averaged by av and similar controls

C*** total number of resulting spectra from averaging and other workup

TOTAL=1

CC 1C I=1,50

SPECTRUM

CC 10 J=1,300

CONTID(I,J)=100

10 CONTINUE

REWIND 30

READ(30,701) IPUNPR

1 NCSPCC=1

CC 12 I=1,300

R(I)=0

DIFFER(I)=0

SIGNAL(I)=0

PROCN(I)=100

12 CONTINUE

2 I06=0

108=G

CRCN=0

READ(30,702) I01, I02, I03, CONTRL, NPTS

IF(EQ(2C), NE, 0) CALL WRAPUP

ECC000(I)=I01, I02, I03

IF(I05.EQ.1H3, OR, I05.EQ.1HC, OR, I05.EQ.1HD) GO TO 400

IF(I05.EQ.1HF, OR, I05.EQ.1HF) GO TO 400

CC 13 I=1,6

SYCTRL(I)=CONTRL(I)

13 CONTINUE

MP; S1V=MPTS

READ(30,704) IEATA(I)

R(I)=CONTRL(6) IEATA(I)

14 CONTINUE

SCALE=CONTRL(6) I010, J

IF(I05.EQ.1(I)) CALL SHIFTS (CRCN)

IF(I02.EQ.1, OR, I03.EQ.1SH) I08=1

IF(I01EIPUNPR(I), EQ, 1HNC, OR, I03.EQ.2HVB) I08=0

IF(I05.EQ.1) CALL SHIFTS (CRCN)

NSMTH=1

IF(I01PLNPR(I), EQ, 2HNO) GC TC 18

C*** 13"point-smoothing"routine

C*** R=SMOOTHED DATA, CD=SMOOTHED DATA, T=TEMPORARY STORAGE

N*NPTS=12

CC 15 I=2,13

J=1

T(I)=R(J)

15 CONTINUE

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)

16 CONTINUE

DO 17 K=1, N

CC 17 I=1, N

J=1+12

DO 16 K=1,12

KK=K+1

T(K)=T(KK)
CONTINUE
T(13)=R(13)
SUM=25.*T(7)+24.*T(6)+T(8)+21.*T(5)+T(9)+16.*T(4)+T(10))
$=9.*T(13)+T(11)-11.*T(11)+T(13))$
L=I+6
CO(I)=SUM/143.
CONTINUE
NSMTH=NPTS+6
CO 19 I=1,6
CO(I)=R(I)
CONTINUE
DC 20 I=NSMTH,300
CO(I)=R(I)
CONTINUE
IF(D2.EQ.ST.CR.ID2.EQ.AV) GC TO 21
IF(D2.EQ.MS.CR.ID2.EQ.SM) GC TO 21
IF(D2.EQ.US.CR.ID2.EQ.SU) GC TO 21
C**** ALL OTHER 1D2 COMBINATIONS ARE INVALID GC TO 1
C**** SAVE DATA FOR AVERAGING
21 ID(NOSPEC)=ID2
CC 22 I=1,NPTS
AVSP(NT(NOSPEC),1)=CD(I)
SIGMA(I)=SIGMA(I)+CD(I)
CONTINUE
IF(ID2.NE.AV) NOSPEC=NOSPEC+1
NOSPEC=1
C**** PUT INDIVIDUAL COMPONENTS ON MICROFICHE FILE
1DEV=19
WRITE(DEV,705) 101
WRITE(DEV,706) 101
IF(CRNS.NE.0) WRITE(IDEV,700) 101,102,103,CRCN
IF(1.PCPR(1),EQ.3,HYES) PUNCH 709,101
C**** AVFLG FLAGS AV TYPE DATA FOR PROPER CSPCSAL (TO MICROFICHE)
AVFLG=1
CALC'MEAN=(ID2;AVFLG)
IF(ID2.EQ.AV) GO TO 2
C**** COMPUTE AVERAGED SPECTRUM
59 CC 100 I=1,NPTS
CO(I)=SIGMA(I)/FLOAT(NOSPEC)
CONTINUE
IF(NOSPEC.LT.2) GO TO 103
C**** COMPUTE 95 PERCENT CONFIDENCE LIMITS
DO 101 J=1,NOSPEC
DO 100 I=1,NPTS
DIFFER(I)=(CD(I)-AVSPCT(J,1))**2*DIFFER(I)
CONTINUE
CC=102=I+1,NPTS
DIFFER(1)=SQRT(DIFFER(1)/FLOAT(NOSPEC)-1.01)
PRECOND(I)=1.L(NOSPEC)*DIFFER(I)/SQRT(FLOAT(NOSPEC))
CONTINUE
CC=104=I+1,NPTS
GC 104
GO TO 316
315 WRITE(7,T237) IDI,ISECID,1DOULD
PUNCH ZD4,101,ISECID,1DOOLD
316 WRITE(7,T06)
102=2HRS
103=2HRT
WRITE(7,T67) IDI,IDO1,IDO3,1CCCTRL(I),I=1,5,SCALE
SPECT(TCTNUM)=IDI
CC 217 I=1,NPTS
A(TCTNUM,I)=CD(I)
CONF,I(TCTNUM,I)=PRCONF(I)
317 CONTINUE
TCTNUM=TCTNUM+1
C**=AVFG-FLAGS-PESULTANT-DATA-FOR-PROPER-DISPOSAL (TO PRINTER)
AVFLG=0
CALL LETSEE (IDO1,AVFLG)
GO TO 1
C**=CC IT AGAIN OPTION
400 EECODE(2,726,IDO2) ID5,106
IF (CCN1FL(I),EQ,1CCCTRL(I)) GC TO 403
ED 402 I=I+6
CONTROL(I)=CCCTRL(I)
402 CONTINUE
NPTS=NPTSSV
403 IF (IDO5,EQ,18) ID2=SM
IF (IDO5,EQ,1H) ID2=MS
IF (IDO5,EQ,1E) ID2=SU
IF (IDO5,EQ,1F) ID2=US
ED 404 I=1,NPTS
AREAD(3C,104,KILLEPT)
404 CONTINUE
I=ID5-1
CC 405 NUM=1,TCTNUM
IF (SPECT(NUM),EQ,1DI) GO TO 406
405 CONTINUE
WRITE(7,T421) ID1
GO TO 1
406 CC 407 I=1,NPTS
CD(I)=A(NUM,I)
PRECNF(I)=CONF(I)
407 CONTINUE
GO TO 300
END
SUBROUTINE LETSEE (101, AVFLG)

CMAIN /PRINT/FLCT/SPUNCH/DATE

COMMON/CG/CD(30)

COMMON/CNSCH/CONS(16), SVCTRL(6), NPTS, NPTSSV

COMMON/PRCONF(300)

COMMON/AVE/WAVE(300)

COMMON/ICOM/IC, 113, IDX, ID05, IC6, ID7, ID8

COMMON/IPUNCH/IPUNPR(31)

REAL CC, CCNTRD, PRCONF, SVCTRL, WAVE

INTEGER AVFLG, IPUNPR

IF (AVFLG.EQ.2) IODEV=7

IF (AVFLG.EQ.1) IODEV=10

IF (AVFLG.EQ.0) IODEV=7

DO 70 I=1,NPTS

10 *WAVE(I)=CONTRL(I)-FLOAT(I-1)*CCNTRD(3)

CONTINUE

100 *NPTS2=I+1

IF (PRCONF(I).GE.0) GC TO 72

WRITE(IODEV,700)

CO 71 I=1,11

J=11

K=J+1

WRITE(IODEV,701) WAVE(1), CD(1), PRCONF(1), WAVE(2), CD(2), PRCONF(2),

12 WAVE(3), CD(3), PRCONF(3),

CONTINUE

72 *WRITE(IODEV,710) CD TO 74

CC=73 I=1,11

J=11

K=J+1

WRITE(IODEV,711) WAVE(1), CD(1), PRCONF(1), WAVE(2), CD(2), PRCONF(2),

13 WAVE(3), CD(3), PRCONF(3),

CONTINUE

74 *WRITE(IODEV,712) 101

XMAX=CONTRL(1)

XINC=CONTRL(2)-CONTRL(1)/TCo:0

TICE=XINC/6.5

*WRITE

IF (*) WRITE(10,NE,1) XINC=0.5*(M+1)

ISY=1

CALL PKNPLT (*WAVE(I), CD(I), XMAX, XINC, XMAY, YMAX, 0, ISY, NPTS, IDEV)

IF (IPUNCH.EQ.2) REAL, CO3, HNO, AND, AVFLG.EQ.1) RETURN

IF (IPUNCH.EQ.1) REAL, CO3, HNO, AND, AVFLG.EQ.2) RETURN

CO 75 I=1,NPTS

IF (CD(I).LT.-59.9, OR, CC(I).GT.999.01 IFORMT=1

CONTINUE

10 IF (IFORMT.NE.1) IFORMT=0

CONT=CCNTRD(1)=1000

SCATE=CONTRL(1)+1000.0

PUNCH 713,101,102,103, (CONTRL(I),I=1,2), CCNTRD, SCALE, (CONTRL(J),

15 J=4,5), IFORMT

IF (IFORMT.EQ.1) GC TO 76

PUNCH 714, (CD(I),I=1,NPTS)

GC TO 77

76 PUNCH 715, (CD(I),I=1,NPTS)

PUNCH 716, 101

RETURN

END
SUBROUTINE SHIFTS (CRCN)

CALL TAILING-SHIFT CALCULATIONS AND CORRECTIONS
COMMON/CO/CO(300)
COMMON/CNSCH/CNTRL(6), SVCTRL(6), NPTS, NPTSSV
COMMON/R/R(300)
COMMON/IC/IC, IC+1, ID, ID+1, ID+2, ID+3
REAL CC,CNTRL,R,SVCTRL

757 FORMAT (I1)
751 FORMAT(A1,11)

C*** XI(HELENTH) HANDLER
ECODE(2,750,103) 104
ECCON(2,751,102) 105, 106
SHAVE=100*ID6*ID4
I=I+ICNTRL(1)-SHAVE/CONTFL(3)
DIFF=R(I-1)-R(I)+R(I-1)-R(I-2)
GO 200 J=1,NPTS
R(J)=R(J)+DIFF

200 CONTINUE
ID2=ZHAV
ID6=0
CRCN=DIFF
RETURN

C*** EM HANDLER
205 CRCA=0
NPNT=3
GO 206 I=3,30
DIFF=((I-1)-R(I)+R(I-1)-R(I-2)
IF (ABS(DIFF), LE, ABS(CRCA)) GO TO 206
CRCN=DIFF
NPNT=1

206 CONTINUE
IF(CRCA.EQ.0) RETURN
GO 207 J=NPNT+1,NPTS
R(J)=R(J)+CRCA

207 CONTINUE
IF (ID2.EQ.2HSH) ID2=ZHAW
RETURN
END

SUBROUTINE WRAPUP

C*** CLOSE UP SHOP
ENDFILE 10
RETURN 10
ENDFILE 7
RETURN 7
CALL EXIT
END
4. Program PROCESS

A) Operation

i) Input

The punched deck from SMOOTH (see note, below).

ii) Output

The file OUTPUT contains listings and plots of all spectra resulting from the manipulations governed by the variable OPTION.

The file TAPE17 contains resultant files for which NEWID was blank or zero. It may be recovered with DISPOSE, but is usually deleted.

The file TAPE7 contains resultant files for which NEWID was non-zero. It is usually saved, either by DISPOSE to the punch or by storage on GSS or PSS.

iii) Errors

The order of spectra is not important, but a spectrum required in a manipulation must either be read in as input or created prior to the request for all necessary spectra. If the spectrum is not present, an error message is left and the next control card is read.

NOTE: NON-FATAL ERROR

In all options but LOOK, MULT, DIFF, PSCD, and EMCD, the quantity CONTRL(5)/CONTRL(6) (i.e. OD/E from Super Spectrum) is first multiplied with the data points in each spectrum before the remainder of the algebra is performed. It is vital that all required spectra are put on the right basis (e.g. equal path lengths for the FDCD, CD, and absorbance spectra in the FDCD option) prior to starting a maneuver.
Example:

<table>
<thead>
<tr>
<th>Spectrum Type</th>
<th>Path length</th>
<th>OD or E or</th>
<th>CONTROL(5)</th>
<th>CONTROL(6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>FDCD</td>
<td>2 mm</td>
<td>1 → 1</td>
<td>500 → 100</td>
<td></td>
</tr>
<tr>
<td>CD</td>
<td>3 mm</td>
<td>3 → 1</td>
<td>1000 → 100</td>
<td></td>
</tr>
<tr>
<td>absorbance</td>
<td>1 cm</td>
<td>1 → 1</td>
<td>100 → 100</td>
<td></td>
</tr>
</tbody>
</table>

where the number to the left of the arrow represents the value of the constant up through the punching of the data deck by SMOOTH and the number to the right of the arrow represents the value of the constant as changed to normalize the spectra to one path length before running in PROCESS.

B) Listing

i) Program PROCESS

ii) Subroutine WRAPUP

iii) Subroutine SEARCH

iv) Subroutine DOMATH

v) Subroutine TITLES

vi) Subroutine SEEIT

vii) Subroutine PRNPLT and Subroutine PLSCAL (see PREPARE)
PROGRAM PROCESS (INPUT, OUTPUT, TAPE, TAPE1)

THIS PROGRAM PERFORMS AN ALGEBRAIC ADDITION OF UP TO FIVE SPECTRA. INPUT OF DATA AND COMMANDS IS THROUGH PUNCHED CARDS. OUTPUT OF RESULTS IS EITHER THROUGH PUNCHED CARDS OR TO PSS OR TAPE, BOTH OF WHICH ARE PERFORMED VIA A DISPOSE OF TAPE.

THE PUNCHED DECK FROM SMOOTH PROGRAM IS HAND IN FIRST, FOLLOWED BY A CARD WITH STOP IN FOUR LEFT COLUMNS.

ALGEBRAIC MANIPULATION CONTROL DECK

GENERAL DECK CARD IS LABEL CARD, CONTROL CARD, LABEL CARD,
CONTROL CARD, ETC.... HOWEVER, IF A DELTA EPSILON CALCULATION IS PERFORMED WITH FDOC, PSCD, OR EMCD, A CARD WITH THE PATH LENGTH (N) AND THE CONCENTRATION (MOL) MUST FOLLOW THE CONTROL CARD FORMAT IS FR.A, E14.4.
LABEL CARD HAS LABEL (SPACE) IN FIRST 4 COLUMNS, FOLLOWED BY ANY PARTICULAR ALPHANUMERIC STRING TO BE PRINTED AT THE TOP OF EACH LISTING AND PLOT.
CONTROL CARD FORMAT IS 61(FR.A, E14.4, A6, I6).
C(1) IS IN FIRST FB.4, C(2) IS IN SECOND FB.4.
SPECTRU IS IN NTH 14 (UP TO 61).
L2 IS OPTION AND L4 IS NEWL (IF DESIRED).
OPTIONS

1) IS SPECTRUM SPECTRUM.
2) IS SPECTRUM SPECTRUM.
LOOK - WRITES AND PLOTS SPECTRUM ONLY.
MULT - MULTIPLIES SPECTRUM BY C(1) OR DIVIDES SPECTRUM BY C(2)
OR MULTIPLIES SPECTRUM BY C(1)/C(2)!
HEIGHT (H) ARE NONZERO.

FLEX - CALCULATES SPECTRA + (C(1)/C(2)) * SPECTRA.
FLEX - CALCULATES AN EXCITATION PROFILE CORRECTED FOR PRE-SLIT
ABSORPTION FROM EXCITATION PROFILE(B), ABS(2), PATHLENGTH TO
SLIT IN CM(C(1)), AND PATHLENGTH OF ABS SPECTRUM IN CM(C(2)).
QIUN - CALCULATES A QUANTITY PROPORTIONAL TO THE QUANTUM YIELD
FROM THE QUANTITIES USED IN FLEX, PLUS THE SLIT WIDTH IN
CM(C(2)). SEE FREDERICKS AND MOUSSIER, BIOPOLYMERS 11, 2281-

BASE (1971) FOR EQUATIONS.
NORM - NORMALIZES SPECTRA TO SPECTRA AT A WAVELENGTH C(1) IN

NM.
RATIO - COMPUTES SPECTRA/SPECTRA RATIO.
FDOC - CALCULATES FLUOROPHORE ANISOTROPY FROM(1)FDOC, (2)CDO
AND (3)ABS. FLUOROPHORE DELTA EPSILON IS CALCULATED WITH (4)ABS.
FLUOROPHORE, IN ADDITION TO OTHERS, A COMPONENT ANALYSIS IS
PROVIDED FOR ANISOTROPY CALCULATIONS ONLY, AND PRODUCES A LIST
OF THAT PORTION OF THE ANISOTROPY DERIVED FROM THE FDOC SIGNAL
AND THAT PORTION DUE TO THE CD SIGNAL.
PSDC - CALCULATES AVERAGE FLUOROPHORE ANISOTROPY FROM FDOC
ANISOTROPIES IN 11WTH POLARIZATION (2) WITH POLARIZATION PLANE
VERTICAL, AND (3) WITH POLARIZATION PLANE HORIZONTAL. AVERAGE
FLUOROPHORE DELTA EPSILON IS CALCULATED WITH (4)ABS FLUOROPHORE
IN ADDITION TO OTHERS.
EMCD - CALCULATES FLUOROPHORE ANISOTROPY IN 33 DIRECTION FROM
FDOC ANISOTROPIES IN THE SAME ORDER AS IN PSCD. FLUOROPHORE
DELTA EPSILON IN 33 DIRECTION IS CALCULATED FROM ABSORPTION AND
C   PCDO MEASUREMENTS IN THE SAME ORDER AS IN PCDO ± ALUS.
C   PCDO - CALCULATES TOTAL ANISOTROPY FROM (1)PCDO AND (2)ABS.
C   SCDO - CALCULATES SCATTERER DELTA EPSILON FROM (1)PCDO AND
C   (2)ABS.
C   NEWID
C   IF NEWID IS BLANK OR LESS THAN -1000, THE RESULTING
C   SPECTRUM IS NOT STORED INTERNALLY. IF NEWID IS BETWEEN -1 AND
C   -999, THE ID IS #TEMPORARY# AND IS NOT LOGGED IN THE LAB
C   RECORD SO IT MAY BE USED AGAIN (BUT NOT IN THE SAME RUN THROUGH
C   PROCESS). IN ALL CASES NOT COVERED UNDER THE FIRST SENTENCE,
C   THE SPECTRUM IS STORED INTERNALLY, UP TO 100 SPECTRA MAY BE
C   STORED IN ONE RUN (300 POINTS EACH).
C   A LABEL CARD THAT IS BLANK OR ONE THAT HAS STOP IN THE FIRST
C   SIX COLUMNS WILL HALT THE PROGRAM.

COMMON/ALO(100,300)
COMMON/CD/CD(300),CPRT(300),CPRT(300)
COMMON/CTRL/CTRL(10),A
COMMON/SPRT/SPRT(5),SPECT(5),INDEX(5),LBL(12)
COMMON/LID/LID(12)
COMMON/ARRAY/ARRAY(10),NUM,NEWID,OPTION,10NUM,WVENMAX,WVENIN
REAL A,CD,CTRL,CDMAX,CVENMAX,CLEN
INTEGER II,INDEX,LBL,SPECT,10NUM
INTEGER (HEAD(12)

611 FORMAT(*$,8X,*), THE FOLLOWING SPECTRA WERE READ INTO MEMORY$,//,
   $6X$, *$SCALE$, 8X, *$CD$, 10X, *$EX$*$\times$)
601 FORMAT(*$,14,A1,2*,2X,3(F7.2,2X,3(F10.3,2X)
653 FORMAT(*$,14,*$, SPECTRA READ INTO MEMORY$)
614 FORMAT(*$,12,12$)
!
701 FORMAT(*$,12,2X$)
701 FORMAT(*$,14,2$, 42, 2X, 3(F8.3,2X),E13.6,2X,E13.6,1X,II$)
702 FORMAT(*$,12F8.3$
704 FORMAT(*$,12F8.2$
705 FORMAT(*$,12X,2X$
706 FORMAT(*$,12,4,14$,2X,4*,41)

C   READ SPECTRA INTO MEMORY
C   10NUM IS THE TOTAL NUMBER OF SPECTRA READ IN
C
REWORK 7
PRINT 650
ON 5 M=1,100
READ 700,HEAD
IF(HEAD[1],EQ.6) STOP 1 GC TO 6
READ 702, ID(M), ID2, ID3, (CONTROL(M, ML), ML=1, 6), IERROR
NPTS=1(CONTROL(M,1)+CONTROL(M,2))/CONTROL(M,3)/10,
IF(1,1,2,3)
1 READ 702, (A(M,1), I=1, NPTS)
GO TO 4
2 READ 702, (A(M,1), I=1, NPTS)
GO TO 4
3 READ 704, (A(M,1), I=1, NPTS)
4 READ 705
5 CONTINUE
6 TOTHUM=4-1
PRINT 602, TOTHUM
7 GO TO ICLR=1,300
CLUD(ICALR)=0
FPTR(ICALR)=0
CPTR(ICALR)=0
8 CONTINUE

to MAX=2,0
MAX=MAX+1
READ 700, LBL
IF(NMLC(E(1), E2, SHAVE, 1, 1, NE, E, LABEL) AND CALL WRAPUP
PRINT 650, (LBL[N], NN=2,131)
READ 706, (C[K]), SPECT(K), XI=1,5, OPTION, NEWID
IOTA=0
CALL SEARCH (XOTIN)
IF(XOIN.EQ.2.1 GO TO 7
CALL DONAM
CALL SEEIT
GO TO 7
END
SUBROUTINE WRAPUP
   C   CLOSE FILE AND EXIT
   ENDFILE 7
   REWIND 7
   ENDFILE 17
   REWIND 17
   CALL EXIT
   END

SUBROUTINE SEARCH (ACTIN)
   COMMON/CPIF,TRL,CCTRL(1,52,6)
   COMMON/SPIF(5),SPECT(5),INDEX(5),LBL(13)
   COMMON/ID,INDEX(100)
   COMMON/PAR,NPTS,NUM,NEWID,OPTION,TOTNUM,WVMAX,WVMIN
   REAL CCTRL(2),WVMAX,WVMIN
   INTEGER ID,INDEX,LBL,SPECT,TOTNUM
   FORMAT(*,/,###/##),*SPECT(4),14,* IS NOT IN MEMORY*)
   DO 100 I=1,5
      INDEX(I)=1
   CONTINUE
   ILO=1
   IHI=2
   IF (OPTION.EQ.4,HSCCD) IHI=1
   IF (OPTION.EQ.4,HSCDD) ILO=2
   IF (OPTION.EQ.4,HSCCO) IHI=1
   IF (OPTION.EQ.4,HSCDO,OPTION.EQ.4,HSCCD) IHI=3
   IF (OPTION,EQ.4,HSCDO,OPTION.EQ.4,HSCCO,OPTION.EQ.4,HSCDD)
      IHI=4
   IF (IHI.LE.1) ILO=1
   IF (INDEX(I).EQ.ID(N)) GO TO 105
   IF (SPECT(I).LE.0.0000) GO TO 105
   IF (SPECT(I).LT.-999) GO TO 102
   IF (TOTNUM.NE.1) GO TO 103
   CONTINUE
   IF (INDEX(I).EQ.ID(N)) GO TO 103
   CONTINUE
   IF (INDEX(I).EQ.ID(N)) GO TO 103
   LIFE=INDEX(I) + N
   INDEX(I)=N
   CONTINUE
   INDEX(I)=N
   NNUM=INDEX(ILO)
   NPTS=(CCTRL(NUM,1)-CCTRL(NUM,2))/CCTRL(NUM,3)/10.0
   IF (WVMAX.GT.CCTRL(NUM,1)) WVMAX=CCTRL(NUM,1)
   IF (WVMAX.LT.CCTRL(NUM,2)) WVMIN=CCTRL(NUM,2)
   RETURN
   END
SUBROUTINE DOMAIN
C EXCL/ A (r, 20, 30)
C EXCL/ECD/CONT(30), CPRT(30)
C EXCL/CONT/CONT1(30)
C EXCL/CONT/CONT2(30)
C EXCL/SPECL/SPEC(5), INDEX(5), LBL(13)
C EXCL/CONT/CONT(10), 6
C EXCL/CONT/A, NEW, OPTION, TOTIM, T, TUMAX, TUMIN
INTEGER ID, INDEX, LBL, SPECT, TCTUH
REAL T, CONT, TUMAX, TUMIN

707
C ASSIGN INDICES AND SELECT PROPER OPTION
C
J = INDEX(1)
K = INDEX(2)
L = INDEX(3)
M = INDEX(4)

IF (OPTION.EQ.4HLOCA) GC TO 15
IF (OPTION.EQ.4HMULTI) GC TO 20
IF (OPTION.EQ.4HDOITF) GC TO 25
IF (OPTION.EQ.4HFLEXI) GC TO 30
IF (OPTION.EQ.4HOUAK) GC TO 30
IF (OPTION.EQ.4HNGEM) GC TO 33
IF (OPTION.EQ.4HRATE) GC TO 35
IF (OPTION.EQ.4HROCD) GC TO 40
IF (OPTION.EQ.4HPEC) GC TO 40
IF (OPTION.EQ.4HMDCD) GC TO 40
IF (OPTION.EQ.4HMCDO) GC TO 50
IF (OPTION.EQ.4HSOCDO) GC TO 60
RETURN
C WRITE OUT AND PLOT SPECTRUM
C
15 GC 17 1=1,NPTS
CONT 4(AJ,1)
11 CONTINUE
C
MULIPLY/ DIVIDCE SPECTRUM BY CONSTANT(S)
C
20 IF(C(1).EQ.0) C(1)=1.0
IF(C(2).EQ.0) C(2)=1.0
CO 22 1=1,NPTS
CONT 1(K(1)/C(1))*AJ,1)
22 CONTINUE
GC TO 64
C
ADD/SUBTRACT TWO SPECTRA
C
25 IF(C(1).EQ.0) C(1)=1.0
IF(C(2).EQ.0) C(2)=1.0
CO 26 1=1,NPTS
CO 11 = AS(J,1)+C(C(1)/C(2))*AI(K,1)
26 CONTINUE
CO TO 64
C
00 COMPUTE FLUORESCENCE EXCITATION PROFILES OR QUANTUM YIELDS

30 IF (C1, EQ, 01) C2 = 1.0
     GO TO 41, NPT$=
     J J = CONT(R(J, 1) - AVE)*1
     K K = CONT(R(K, 1) - AVE)*1
     A = CONT(R(J, 1) + CONT(R(K, 5)) / CONT(R(K, 6))
     EXP = EXP(R(J, 1))
     SLIT = (1.3 - C7, EXP)
     IF (OPTION, E2, PLEX) SLIT = 1.0
     CONT(R(J, 1) + CONT(R(K, 5)) / CONT(R(K, 6)) = (FCORR/SLIT)

31 CONTINUE
     GO TO 64

C NORMALIZE SPECT11 TO SPECT12 AT WAVELENGTH C1

32 WAVE = ((CONT(R(J, 1)) / CONT(R(J, 3))/10)+1
     IF (IN, EQ, 01) RETURN
     C = CONT(R(J, 1) - AVE)*1
     C = CONT(R(J, 5)) / CONT(R(K, 5)) / CONT(R(K, 6))
     CONT(R(J, 1) = C1
     CONT(R(J, 5) = C1

33 CONTINUE
     GO TO 64

C COMPUTE RATIO OF TWO SPECTRA

35 CONT(R(J, 1) = C1
     C = CONT(R(J, 1) - AVE)*1
     K K = CONT(R(K, 1) - AVE)*1
     A = CONT(R(J, 5)) / CONT(R(K, 5)) / CONT(R(K, 6)
     IF (IN, EQ, 01) A = CONT(R(K, 5))
     CONT(R(J, 1) = A

36 CONTINUE
     GO TO 64

C GET DATA FOR FORC, PSCC, AND EMCD

38 CONT(R(J, 1) = C1
     C = CONT(R(J, 1) - AVE)*1
     K K = CONT(R(K, 1) - AVE)*1
     L L = CONT(R(L, 1) - AVE)*1
     S1 = CONT(R(J, 1)
     S2 = CONT(R(K, 1)
     S3 = CONT(R(L, 1)
     IF (OPTION, EQ, 02) GO TO 46
     IF (OPTION, EQ, 04) GO TO 48
     RETURN

C COMPUTE FLUOROPHORE ANISTROPHY

C
SUBROUTINE Titles (J,K,L,M)
COMMON/VC/COI(20),FPRT(30),CPRT(30)
COMMON/CNTL/CONT(10),E
COMMON/SPNL/C(15),SPECT(15),INDEX(15),LBL(13)
COMMON/FRAM/FRM(10),FNM(10)
COMMON/NUM/NUM.NEW,OPTION,TOTNUM,VEVAX,VWEVIN
PEEL = CNTL.CENTRIC,VEVAX,VWEVIN
INTEGER ID,INDEX,LBL,SPECT,TOTNUM
625 FORMAT(*,14,A) IS *14,* X (*,E10,4,*),E10,4,*)
633 FORMAT(*,14,A) IS *14,* X (*,E10,4,* / *,E10,4,* ) X *,14,
650 FORMAT(*,14,A) IS FLUOROPHORE ANISOTROPY X1000 FROM FDCO = *14,
$1 FROM ANISOTROPY = *14,* AND PHI=90 ANISOTROPY = *
651 FORMAT(*,14,A) IS AVERAGE FLUOROPHORE ANISOTROPY X1000 FROM ANISOTROPY = *14,* AND PHI=90 ANISOTROPY = *
652 FORMAT(*,14,A) IS FLUOROPHORE ANISOTROPY ALONG 33 DIRECTION X1000
$1 FROM ANISOTROPY = *14,* AND PHI=90 ANISOTROPY = *
656 FORMAT(*,14,A) IS TOTAL ANISOTROPY X1000 FROM CO = *14,
$ AND ABS = *14,
657 FORMAT(*,14,A) IS SCATTER DELTA EPSILON FROM FDCO = *14,
$ AND ABS = *14,
660 FORMAT(*,14,A) IS CORRECTED FLUORESCENCE EXCITATION PROFILE OF
$14,* USING *14,* PRE-SLIT PATH = *8,4,* CM^6
661 FORMAT(*,14,A) IS PROPORTIONAL TO QUANTUM YIELD OF *14,* USING *
$14,* PRE-SLIT PATH = *8,4,* CM, SLIT WIDTH = *8,4,* CM^4
662 FORMAT(*,14,A) IS *14,* NORMALIZED TO *14,* AT = *8,4,* NM^4
664 FORMAT(*,14,A) LISTING
665 FORMAT(*,4,A) THIS
666 FORMAT(*,14,A) IS A TEMPORARY, PEUSABLE FILE NUMBER
676 FORMAT(*,14,A) IS AVG FL ANISO X1000 FROM FA = *14,* PHIO = *14,
$ AND PHIS = *14,
682 FORMAT(*,14,A) IS AVG FL ANISO 33 X1000 FROM FA = *14,* PHIO = *14,
$ AND PHIS = *14,
683 FORMAT(*,14,A) IS FL DEL EPS FROM FDC = *14,* CD = *14,
$ AND ABS = *14,
684 FORMAT(*,14,A) IS FL DEL EPS FROM FA = *14,* PHIO = *14,
$ AND ABS = *14,
685 FORMAT(*,14,A) IS FL DEL EPS 33 FROM FA = *14,* PHIO = *14,
$ AND ABS = *14,
686 FORMAT(*,14,A) IS TOTAL ANISOTROPY X1000 FROM CD = *14,* AND ABS = *,
862  SELF= *, SPLIT CM= *, 9, 4, *)
862  FORMA1(14, * IS *, 14, * NORM ALIZED TO *, 14, * AT *
862  FORMA1(14, * IS *, 14, * / *, 14, *)
864  FORMA1(14, * LISTING*)
C  T C E F E R E S U L T S IF DESIRED
C
[DEV= 7
IF (NEWID, GT, 0000) GC TC 300
IF (NEWID, LT, -9990, OR, NEA1D, EQ, 0000) GO TO 299
PRINT 681, NEW1D
GO TO 300
299  NEA1D= 0
IF (OPTION, EQ, 'HL1C0K') GC TC 303
PRINT 680
GC TC 303
300  TCTNUM= TCTNUM+1
IC(TCTNUM)= NEW1D
CONT(I(TCTNUM)=1)= VEWMAX
CONT(I(TCTNUM)=2)= VEWMIN
GC 321  I= 3, 6
CONT(I(TCTNUM)=11)= CONTR(I(NUM)=1)
CONTINUE
NPTS= (CONTR(I(TCTNUM)=11)-CONTR(I(TCTNUM)=1))/ (CONTR(I(TCTNUM)=31)/10, 1
GC 302  I= 1, NPTS
302  CONTINUE
NUM= TCTNUM
C
C  PRINT, PUNCH TITLES
C  TAP SLY IS THE "DEG LETTER FILE" FOR UNIDENTIFIED SPECTRA
C
303  IF (NEA1D, EQ, 0,1) DEV= 17
IF (OPTION, EQ, 'MULTI') GC TO 400
IF (OPTION, EQ, 'HDF1F') GC TO 401
IF (OPTION, EQ, 'HDF1D') GC TO 402
IF (OPTION, EQ, 'HDF2D') GC TO 404
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 405
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 406
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 407
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 408
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 409
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 410
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 411
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 412
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 413
IF (OPTION, EQ, 'HDF2D', AND, SPECT(4), 'HDF2D') GO TO 414
RETURN
400 WRITE(IDEV,9275) NEWID, SPECT(1), C(1), C(2)
PRINT 625, NEWID, SPECT(1), C(1), C(2)
RETURN
401 WRITE(IDEV,9275) NEWID, SPECT(1), C(1), C(2), SPECT(2)
PRINT 636, NEWID, SPECT(1), C(1), C(2), SPECT(2)
RETURN
402 WRITE(IDEV,9275) NEWID, ID(I), ID(L)
PRINT 656, NEWID, ID(I), ID(L)
RETURN
403 WRITE(IDEV,9275) NEWID, ID(I), ID(L)
PRINT 657, NEWID, ID(I), ID(L)
RETURN
404 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L)
PRINT 657, NEWID, ID(I), ID(D), ID(L)
RETURN
405 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L), ID(M)
PRINT 658, NEWID, ID(I), ID(D), ID(L), ID(M)
RETURN
406 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L), ID(M)
PRINT 659, NEWID, ID(I), ID(D), ID(L), ID(M)
RETURN
407 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L), ID(M)
PRINT 555, NEWID, ID(I), ID(D), ID(L), ID(M)
RETURN
408 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L)
PRINT 651, NEWID, ID(I), ID(D), ID(L)
RETURN
409 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L)
PRINT 652, NEWID, ID(I), ID(D), ID(L)
RETURN
410 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L)
PRINT 652, NEWID, ID(I), ID(D), ID(L)
RETURN
411 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L), C(1)
PRINT 662, NEWID, ID(I), ID(D), ID(L), C(1)
RETURN
412 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L), C(1)
PRINT 662, NEWID, ID(I), ID(D), ID(L), C(1)
RETURN
413 WRITE(IDEV,9275) NEWID, ID(I), ID(D), ID(L)
PRINT 662, NEWID, ID(I), ID(D), ID(L)
RETURN
414 WRITE(IDEV,9275) NEWID, ID(I), ID(D)
PRINT 653, NEWID, ID(I), ID(D)
RETURN
415 WRITE(IDEV,9275) ID(I)
PRINT 664, ID(I)
RETURN
END
REAL WAVE(300)
672  FORMAT(=,=ANISOTROPY COMPONENT ANALYSIS OF *14)
673  FORMAT(=,=ANISOTROPY COMPONENT ANALYSIS OF *14)
674  FORMAT(=,=SPECTRUM *14)
675  FORMAT(=,=SPECTRUM *14)
676  FORMAT(=,=SPECTRUM *14)
677  FORMAT(=,=SPECTRUM *14)
678  FORMAT(=,=SPECTRUM *14)
679  FORMAT(=,=SPECTRUM *14)
680  FORMAT(=,=SPECTRUM *14)
681  FORMAT(=,=SPECTRUM *14)
682  FORMAT(=,=SPECTRUM *14)
683  FORMAT(=,=SPECTRUM *14)
684  FORMAT(=,=SPECTRUM *14)
685  FORMAT(=,=SPECTRUM *14)
686  FORMAT(=,=SPECTRUM *14)
687  FORMAT(=,=SPECTRUM *14)
688  FORMAT(=,=SPECTRUM *14)
689  FORMAT(=,=SPECTRUM *14)
690  FORMAT(=,=SPECTRUM *14)
691  FORMAT(=,=SPECTRUM *14)
692  FORMAT(=,=SPECTRUM *14)
693  FORMAT(=,=SPECTRUM *14)
694  FORMAT(=,=SPECTRUM *14)
695  FORMAT(=,=SPECTRUM *14)
696  FORMAT(=,=SPECTRUM *14)
697  FORMAT(=,=SPECTRUM *14)
698  FORMAT(=,=SPECTRUM *14)
699  FORMAT(=,=SPECTRUM *14)
700  FORMAT(=,=SPECTRUM *14)
701  FORMAT(=,=SPECTRUM *14)
702  FORMAT(=,=SPECTRUM *14)
703  FORMAT(=,=SPECTRUM *14)
704  FORMAT(=,=SPECTRUM *14)
705  FORMAT(=,=SPECTRUM *14)
706  FORMAT(=,=SPECTRUM *14)
707  FORMAT(=,=SPECTRUM *14)
708  FORMAT(=,=SPECTRUM *14)
709  FORMAT(=,=SPECTRUM *14)
710  FORMAT(=,=SPECTRUM *14)
711  FORMAT(=,=SPECTRUM *14)
712  FORMAT(=,=SPECTRUM *14)
713  FORMAT(=,=SPECTRUM *14)
714  FORMAT(=,=SPECTRUM *14)
715  FORMAT(=,=SPECTRUM *14)
716  FORMAT(=,=SPECTRUM *14)
717  FORMAT(=,=SPECTRUM *14)
718  FORMAT(=,=SPECTRUM *14)
719  FORMAT(=,=SPECTRUM *14)
720  FORMAT(=,=SPECTRUM *14)
721  FORMAT(=,=SPECTRUM *14)
722  FORMAT(=,=SPECTRUM *14)
723  FORMAT(=,=SPECTRUM *14)
724  FORMAT(=,=SPECTRUM *14)
725  FORMAT(=,=SPECTRUM *14)
726  FORMAT(=,=SPECTRUM *14)
727  FORMAT(=,=SPECTRUM *14)
728  FORMAT(=,=SPECTRUM *14)
729  FORMAT(=,=SPECTRUM *14)
730  FORMAT(=,=SPECTRUM *14)
731  FORMAT(=,=SPECTRUM *14)
732  FORMAT(=,=SPECTRUM *14)
733  FORMAT(=,=SPECTRUM *14)
734  FORMAT(=,=SPECTRUM *14)
735  FORMAT(=,=SPECTRUM *14)
736  FORMAT(=,=SPECTRUM *14)
737  FORMAT(=,=SPECTRUM *14)
738  FORMAT(=,=SPECTRUM *14)
739  FORMAT(=,=SPECTRUM *14)
740  FORMAT(=,=SPECTRUM *14)
741  FORMAT(=,=SPECTRUM *14)
742  FORMAT(=,=SPECTRUM *14)
743  FORMAT(=,=SPECTRUM *14)
744  FORMAT(=,=SPECTRUM *14)
745  FORMAT(=,=SPECTRUM *14)
746  FORMAT(=,=SPECTRUM *14)
747  FORMAT(=,=SPECTRUM *14)
748  FORMAT(=,=SPECTRUM *14)
749  FORMAT(=,=SPECTRUM *14)
750  FORMAT(=,=SPECTRUM *14)
751  FORMAT(=,=SPECTRUM *14)
752  FORMAT(=,=SPECTRUM *14)
753  FORMAT(=,=SPECTRUM *14)
754  FORMAT(=,=SPECTRUM *14)
755  FORMAT(=,=SPECTRUM *14)
756  FORMAT(=,=SPECTRUM *14)
757  FORMAT(=,=SPECTRUM *14)
758  FORMAT(=,=SPECTRUM *14)
759  FORMAT(=,=SPECTRUM *14)
760  FORMAT(=,=SPECTRUM *14)
761  FORMAT(=,=SPECTRUM *14)
762  FORMAT(=,=SPECTRUM *14)
763  FORMAT(=,=SPECTRUM *14)
764  FORMAT(=,=SPECTRUM *14)
765  FORMAT(=,=SPECTRUM *14)
```
DC PA N=1, NPTS

WAVE = CONTROLS, 1 = FLOAT (N-1) * (CONTROLS(NUM,37)/10).

CONTINUE
II = NPTS / 3 + 1
CO 87 I = II
J = II
K = J + II
PRINT 676, WAVE(I), CO(I), WAVE(I), CO(I), WAVE(I), CO(I)
CONTINUE
IF (OPTION, NE, 'MEMBER', PI, SPECT(47, NE, 0000) GO TO 89
PRINT 678, (L1L(111), NN, 3, 13)
PRINT 672, NEWID
PRINT 673
II = NPTS / 3 + 1
CO 88 I = II
J = II
K = J + II
PRINT 674, WAVE(I), FPRT(I), CPRT(I), WAVE(I), FPRT(I), CPRT(I)
CONTINUE
PRINT 678, (L1L(111), AN, 2, 13)
PRINT 677, NEWID
XINC = (CONTROLS(NUM, 2)) / 100.0
THICE = XINC / 0.5
M = THICE
IF (M = #, #, THICE) XINC = 0.5 * (#+1)
ISY = 1
CALL PRINTL (WAVE(1), CC(1), XMAD, XINC, YMAD, YINC, O, ISY, NPTS)
RETURN
END```

5. Control Deck for Running PREPARE and SMOOTHs in Tandem

```
RUNC'D.7,200,100000.448402  
*NOSTAGE
FETCHPS,CDLIB,LGO,PREPARE
LIBCOPY,CDL13,TAPE5/BR,DATA.
LINK,X,PP=[TAPE5,OUTPUT,CONTROL].
DELETE,TAPE5,LGO.
COPY,OUTPUT/RB,ORXR,TAPE6/BR.
DISPOSE,TAPE6=PR,H0,DT=I.
STOTAPE,TAPE30=/NELSONJW/KSD/DATA/prepare/Dataset,10515.
FETCHPS,CDLIB,LGO,SMOOTHs.
LINK,X,PP=[TAPE30,OUTPUT,PUNCH].
DELETE,TAPE10,TAPE30,LGO.
DISPOSE,OUTPUT=PR,DT=I.
END.
```

6. Conversion of a Spectrum

A) The spectrum as printed by the -G- option of Super Spectrum.

```
RUN ID 0000STKD
OD = 0.100000E+01
E = 0.100000E+01

LAMBDA (A) RAW DATA
4000 .0000
3900 .1999
3800 .3999
3700 .5999
3600 .7999
3500 .9999
3400 1.1999
3300 1.3999
3200 1.5999
3100 .9999
3000 .0999
2900 1.5999
2800 1.3999
2700 1.1999
2600 .9999
2500 .7999
2400 .5999
2300 .3999
2200 .1999
2100 .0000
```
B) The spectrum as represented internally in the PDP/8E by Super Spectrum during transmission.

All numbers are octal.

Locations give data field in first digit, address in remaining digits, e.g. 12002 is location 2002 of data field 1.

<table>
<thead>
<tr>
<th>Location</th>
<th>Contents.... Data Points</th>
</tr>
</thead>
<tbody>
<tr>
<td>14000</td>
<td>0000 0310 0620 1130 1440 1750 2260 2570</td>
</tr>
<tr>
<td>14010</td>
<td>3100 0144 0144 3100 2570 2260 1750 1440</td>
</tr>
<tr>
<td>14020</td>
<td>1130 0620 0310 0000 0000 0000 0000 0000</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Location</th>
<th>Contents.... Spectrum Parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td>15740</td>
<td>0000 0000 0260 0260 0260 0260 0323 0324</td>
</tr>
<tr>
<td>15750</td>
<td>0313 0304 0000 7640 0310 7634 7754 0001</td>
</tr>
<tr>
<td>15760</td>
<td>2000 0000 0001 2000 0000 7767 2030 4467</td>
</tr>
<tr>
<td>15770</td>
<td>0000 0000 0000 0000 0000 0000 0000 0000</td>
</tr>
</tbody>
</table>

The spectrum ID (ASCII 8-bit characters) is at 15742+
The starting wavelength (in Å, double precision) is at 15752+
The ending wavelength (in nm, single precision) is at 15754
The increment between points (in Å, single precision) is at 15755
The number of points (negated, single precision) is at 15756
The constant OD (floating point) is at 15757+
The constant E (floating point) is at 15762+
The scale (x 0.001) (floating point) is at 15765+
The data points (each x 1000, single precision) are at 14000+
C) The spectrum as typed at the keyboard during transmission and stored in PSS.

This is the input to PREPARE.

The parameter line is first, followed by data line(s).

```
0B00B00B00800030D40CB0C4000FA00C8F9CFEC001400000001400000
FF74189370000000013D
```

There are 25 12-bit numbers represented per line with a checksum at the end. A line is filled in with zeroes if there are fewer than 25 numbers to send.

**Hexadecimal to Binary Conversion**

<table>
<thead>
<tr>
<th>Hex</th>
<th>Binary</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0000</td>
</tr>
<tr>
<td>1</td>
<td>0001</td>
</tr>
<tr>
<td>2</td>
<td>0010</td>
</tr>
<tr>
<td>3</td>
<td>0011</td>
</tr>
<tr>
<td>4</td>
<td>0100</td>
</tr>
<tr>
<td>5</td>
<td>0101</td>
</tr>
<tr>
<td>6</td>
<td>0110</td>
</tr>
<tr>
<td>7</td>
<td>0111</td>
</tr>
<tr>
<td>8</td>
<td>1000</td>
</tr>
<tr>
<td>9</td>
<td>1001</td>
</tr>
<tr>
<td>A</td>
<td>1010</td>
</tr>
<tr>
<td>B</td>
<td>1011</td>
</tr>
<tr>
<td>C</td>
<td>1100</td>
</tr>
<tr>
<td>D</td>
<td>1101</td>
</tr>
<tr>
<td>E</td>
<td>1110</td>
</tr>
<tr>
<td>F</td>
<td>1111</td>
</tr>
</tbody>
</table>

**Sample Conversion for ID Characters K and D**

K = 0 3 1 3 in 8-bit ASCII

```
000 011 001 011
```

in binary

```
0000 1100 1011
```

in hexadecmial

D = 0 3 0 4 in 8-bit ASCII

```
000 011 000 100
```

in binary

```
0000 1100 0100
```

in hexadecimal

```
0 C 4
```

These are found in the first line above, starting with the 19th character.
D) The spectrum as written on TAPE30 by PREPARE after conversion.

This is the input to SMOOTHS.

```
0000STKD 400.00 200.00 10.00   1.000 1.000 0.001000 20
0 200 400 600 8001000120014001600 100 1001600140012001000 800 600
400 200 0
```

E) The spectrum as punched by SMOOTHS.

Punch raw data option is on, smoothing is off.

```
0000STKD  400.000  200.000  100.000  1.000 0.100000E+01
0.100000E+01 0
.0000  .2000  .4000  .6000  .8000  1.0000  1.2000  1.4000
1.6000  .1000
.1000  1.6000  1.4000  1.2000  1.0000  .8000  .6000  .4000
.2000  .0000
```

END OF 0000 SMOOTHS DECK
Appendix D

DNA/ETHIDIUM ION COMPLEX CHARGE DENSITY

1/ Ethidium ion is 3.4 Å thick and carries a +1 charge. B form DNA in the absence of dye binding is 3.4 Å long per base pair. Each base pair contains two phosphate groups, each with a -1 charge.

2/ Assume nearest neighbor exclusion model. Then, for a DNA helix 2n base pairs long there are 4n phosphate groups but only n binding sites. Each binding site is composed of 2 nucleotide units per strand.

3/ Binding site states are:

   Empty       2 base pairs = 6.8 Å long
               4 phosphates = -4 net charge

   Occupied    2 base pairs + dye = 10.2 Å long
               4 phosphates + dye = -3 net charge

4/ Use r, the extent of binding, to follow titration.

   \[ r = \frac{\text{ethidium ion bound/DNA (as phosphate)}}{k/4n}, \text{ where } k = \text{number of bound dyes} \]  
   \[ r_{\text{max}} = 0.25 \text{ in neighbor exclusion limit} \]  

5/ \( b = \text{distance/charge} = \)

   \[ \frac{[6.8(n - k) + 10.2(k)]}{[4(n - k) + 3(k)]} \]  

Combine (1) and (2) to eliminate k, then

\[ b = 1.7[1 + 3r/(1 - r)] \]

The \( 1/(1 - r) \) dependence was previously found for DNA titration by acids by Record et al. (1976).