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An Average-Case Sublinear Exact Li and Stephens Forward Algorithm

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Abstract

Hidden Markov models of haplotype inheritance such as the Li and Stephens model allow for computationally tractable probability calculations using the forward algorithms as long as the representative reference panel used in the model is sufficiently small. Specifically, the monoploid Li and Stephens model and its variants are linear in reference panel size unless heuristic approximations are used. However, sequencing projects numbering in the thousands to hundreds of thousands of individuals are underway, and others numbering in the millions are anticipated.

To make the Li and Stephens forward algorithm for these datasets computationally tractable, we have created a numerically exact version of the algorithm with observed average case $O(nk^{0.35})$ runtime in number of genetic sites $n$ and reference panel size $k$. This avoids any tradeoff between runtime and model complexity. We demonstrate that our approach also provides a succinct data structure for general purpose haplotype data storage. We discuss generalizations of our algorithmic techniques to other hidden Markov models.

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1 Introduction

Probabilistic models of haplotypes describe how variation is shared in a population. One application of these models is to calculate the probability \( P(o|H) \) of a haplotype \( o \) given the assumption that it is a member of a population represented by a reference panel of haplotypes \( H \). This computation has been used in estimating recombination rates [8], a problem of interest in genetics and in medicine. It may also be used to detect errors in genotype calls.

Early approaches to haplotype modeling used coalescent [7] models which were accurate but computationally complex, especially when including recombination. Li and Stephens wrote the foundational computationally tractable haplotype model [8] with recombination. Under their model, the probability \( P(o|H) \) can be calculated using the forward algorithm for hidden Markov models. Generalizations of their model have been used for haplotype phasing and genotype imputation. Most of these algorithms [10, 1, 14, 3, 12] use the forward probabilities calculated as intermediate values in the forward algorithm.

1.1 The Li and Stephens model

Consider a reference panel \( H \) of \( k \) haplotypes sampled from some population. Each haplotype \( h_j \in H \) is a sequence \( (h_{j,1}, \ldots, h_{j,n}) \) of alleles at a contiguous sequence \( 1, \ldots, n \) of genetic sites. Classically [8], the sites are biallelic, but the model extends to multiallelic sites. [11]

Consider an observed sequence of alleles \( o = (o_1, \ldots, o_n) \) representing another haplotype.

The monoploid Li and Stephens model (LS) [8] specifies a probability that \( o \) is descended from the population represented by \( H \). LS can be written as a hidden Markov model where the haplotype \( o \) is assembled by copying (with possible error) consecutive contiguous subsequences of haplotypes \( h_j \in H \).

▶ Definition 1 (Li and Stephens HMM). Define \( x_{j,i} \) as the event that the allele \( o_i \) at site \( i \) of the haplotype \( o \) was copied from the allele \( h_{j,i} \) of haplotype \( h_j \in H \). Take parameters

\[
\begin{align*}
\rho_{i-1\rightarrow i}^* & \quad \text{the probability of any recombination between sites } i-1 \text{ and } i \\
\mu_i & \quad \text{the probability of a mutation from one allele to another at site } i
\end{align*}
\]

and from them define the transition and recombination probabilities

\[
\begin{align*}
p(x_{j,i}|x_{j',i-1}) & = \begin{cases} 
1 - (k - 1)\rho_{i} & \text{if } j = j' \\
\rho_{i} & \text{if } j \neq j'
\end{cases} \quad \text{where } \rho_i = \frac{\rho_{i-1\rightarrow i}^*}{k-1} \quad (3) \\
p(o_i|x_{j,i}) & = \begin{cases} 
1 - (A - 1)\mu_{i} & \text{if } o_i = h_{j,i} \\
\mu_{i} & \text{if } o_i \neq h_{j,i}
\end{cases} \quad \text{where } A = \text{number of alleles} \quad (4)
\end{align*}
\]

We will write \( \mu_i(j) \) as shorthand for \( p(o_i|x_{j,i}) \). We will also define the values of the initial probabilities \( p(x_{j,1},o_1|H) = \frac{\mu_i(1)}{k} \).

Let \( P(o|H) \) be the probability that haplotype \( o \) was produced from population \( H \). The forward algorithm for hidden Markov models allows calculation of this probability in \( O(nk^2) \) time using an \( n \times k \) dynamic programming matrix of forward states

\[
p_{i}[j] = P(x_{j,i},o_1,\ldots,o_i|H) \quad (5)
\]

In practice, the Li and Stephens forward algorithm is \( O(nk) \). (See §3)
1.1.1 Li and Stephens like algorithms for large populations

The $O(nk)$ time complexity of the forward algorithm is intractable for reference panels with large size $k$. The UK Biobank has amassed $k = 500,000$ array samples. Whole genome sequencing projects, with a denser distribution of sites, are catching up. Major sequencing projects with $k = 100,000$ or more samples are nearing completion. Others numbering $k$ in the millions have been announced. These large population datasets have significant potential benefits: They are statistically likely to more accurately represent population frequencies and those employing genome sequencing can provide phasing information for rare variants.

In order to handle datasets with size $k$ even fractions of these sizes, modern haplotype inference algorithms depend on models which are simpler than the Li and Stephens model or which sample subsets of the data. For example, the common tools Eagle-2, Beagle, HAPI-UR and Shapeit-2 and -3 [10, 1, 14, 3, 12] either restrict where recombination can occur, fail to model mutation, model long-range phasing approximately or sample subsets of the reference panel.

Lunter’s “fastLS” algorithm [11] demonstrated that haplotypes models which include all $k$ reference panel haplotype could find the Viterbi maximum likelihood path in time sublinear in $k$, using preprocessing to reduce redundant information in the algorithm’s input. However, his techniques do not extend to the forward and forward-backward algorithms.

1.2 Our contributions

We have developed an arithmetically exact forward algorithm whose expected time complexity is a function of the expected allele distribution of the reference panel. This expected time complexity proves to be $O(k^{0.35})$ in reference panel size. We have also developed a technique for succinctly representing large panels of haplotypes whose size also scales as a sublinear function of the expected allele distribution.

Our forward algorithm contains three optimizations, all of which might be generalized to other bioinformatics algorithms. In (§2), we rewrite the reference panel as a sparse matrix containing the minimum information necessary to directly infer all allele values. In (§3), we define recurrence relations which are numerically equivalent to the forward algorithm but use minimal arithmetic operations. In (§4), we delay computation of forward states using a lazy evaluation algorithm which benefits from blocks of common sequence. Our methods apply to other models which share certain properties with the monoploid Li and Stephens model.

2 Sparse representation of haplotypes

The forward algorithm to calculate the probability $P(o|H)$ takes as input a length $n$ vector $o$ and a $k \times n$ matrix of haplotypes $H$. Therefore time complexity better than $O(nk)$ is impossible unless there is preprocessing of its input. However, such preprocessing can be amortized over many queries $o$.

2.1 Information content of a reference panel

Recall that $(o_i)_{i=1}^n$ is the allele sequence of the emitted haplotype $o$. (§3) will show that $\phi_i(o_i), 1 \leq i \leq n$ defined below are sufficient data to calculate $P(o|H)$.

Definition 2. The information content $\phi$ of $H$ for allele $a$ at site $i$ is defined as

$$\phi_i(a) = \begin{cases} Match_i(a) & \text{if } |Match_i(a)| \leq |NonMatch_i(a)| \\ NonMatch_i(a) & \text{if } |NonMatch_i(a)| < |Match_i(a)| \end{cases}$$

(6)
We will often abuse notation and refer to the \( h_j \in \phi_i(a) \) by their indices \( j \) alone.

### 2.2 Relation of information content to allele frequency spectrum

Our sparse representation of the haplotype reference panel benefits from the recent finding \(^2\) that the distribution over sites of minor allele frequencies is biased towards low frequencies\(^3\).

We will compute the expected time sum of the information content over all sites assuming first that all sites are biallelic\(^3\). In the biallelic case \( \phi_i(\cdot) \) is always the set of haplotypes displaying the minor allele at site \( i \) and the distribution of \( \phi_i(a) \) is the allele frequency spectrum.

\[
\text{Lemma 3. Let } E(\bar{T})(k) \text{ be the expected mean minor allele frequency for } k \text{ genotypes. Then }

E\left[ \frac{1}{n} \sum_{i=1}^{n} |\phi_i(a)| \right] = E(\bar{T})(k)
\]

\[
\text{(7)}
\]

\[
\text{Corollary 4. If } O(E(\bar{T})) < O(k), \text{ then } O(\sum_i |\phi_i(a)|) < O(nk) \text{ in expected value.}
\]

### 2.3 Implementation

For biallelic sites, we store our \( \phi_i \)'s using a length-\( n \) vector of length \(|\phi_i|\) vectors containing the indices \( j \) of the haplotypes \( h_j \in \phi_i \) and a length-\( n \) vector listing the major allele at each site. (See Figure 1 panel iii) Random access by key \( i \) to iterators to the first elements of sets \( \phi_i(a) \) is \( O(1) \) and iteration across these \( \phi_i(a) \) is linear in the size of \( \phi_i(a) \). For multiallelic sites, the data structure uses slightly more space but has the same speed guarantees.

Generating these data structures takes \( O(nk) \) time but is embarrassingly parallel in \( n \). Our “*.slls” data structure doubles as a succinct haplotype index which could be distributed instead of a large vcf record. A vcf → slls conversion tool is found in our github repository.

Adding or rewriting a haplotype is constant time per site per haplotype unless this edit changes which allele is the most frequent. This allows our algorithm to extend to uses of the Li and Stephens model where one might want to dynamically edit the reference panel.

### 3 Efficient dynamic programming

We begin with the recurrence relation of the \( O(nk) \) Li and Stephens forward algorithm \(^8\).

To establish our notation, recall that \( p_i[j] = P(x_{j,i}, \alpha_1, \ldots, \alpha_i|H) \), that we write \( \mu_i(j) \) as

\[(7)
\]

\[\text{We confirm these results in section 5.2}
\]

\[\text{The generalization is trivial}\]
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shorthand for \( p(o_i|x_{j,i}) \) and that we have initialized \( p_1[j] = p(x_{j,1}, o_1|H) = \frac{n(j)}{k} \). For \( i > 1 \), we may then write:

\[
p_i[j] = \mu_i(j)(1 - k \rho_i)p_{i-1}[j] + \rho_i S_{i-1}
\]

(8)

\[
S_i = \sum_{j=1}^{k} p_i[j]
\]

(9)

We will reduce the number of summands in (9) and reduce the number indices \( j \) for which (8) is evaluated. This will use the information content defined in (§2.1).

**Lemma 5.** The summation (9) is calculable using strictly fewer than \( k \) summands.

**Proof.** Suppose first that \( \mu_i(j) = \mu_i \) for all \( j \). Then

\[
S_i = \sum_{j=1}^{k} p_i[j] = \mu_i \sum_{j=1}^{k} ((1 - k \rho_i)p_{i-1}[j] + \rho_i S_{i-1})
\]

(10)

\[
= \mu_i((1 - k \rho_i)S_{i-1} + k \rho_i S_{i-1}) = \mu_i S_{i-1}
\]

(11)

Now suppose that \( \mu_i(j) = 1 - \mu_i \) for some set of \( j \). We must then correct for these \( j \). This gives us

\[
S_i = \mu_i S_{i-1} + \frac{1 - \mu_i - \mu_i}{1 - \mu_i} \sum_{j \in \phi_i(o_i)} p_i[j]
\]

(12)

The same argument holds when we reverse the roles of \( \mu_i \) and \( 1 - \mu_i \). Therefore we can choose which calculation to perform according to which involves a sum with fewer summands. This gives us the following formula:

\[
S_i = \alpha S_{i-1} + \beta \sum_{j \in \phi_i(o_i)} p_i[j]
\]

(13)

where

\[
\alpha = \mu_i \quad \beta = \frac{1 - 2 \mu_i}{1 - \mu_i} \quad \text{if} \ \phi_i(a) = \text{Match}_i(a)
\]

(14)

\[
\alpha = 1 - \mu_i \quad \beta = \frac{2 \mu_i - 1}{\mu_i} \quad \text{if} \ \phi_i(a) = \text{NonMatch}_i(a)
\]

(15)

**Lemma 6.** If \( j \notin \phi_i(o_i) \) and \( j \notin \phi_{i-1}(o_{i-1}) \), then \( S_i \) can be calculated without knowing \( p_{i-1}[j] \) and \( p_i[j] \), as can \( p_i[j'] \) for \( j' \neq j \).

**Proof.** By inspection of equation (13).

**Corollary 7.** The recurrences (9) and the minimum set of recurrences (8) needed to compute (9) can be evaluated in \( O(|\phi|) \) time, assuming that \( p_{i-1}[j] \) have been computed \( \forall j \in \phi_i(o_i) \).

We address the assumption on prior calculation of the necessary \( p_{i-1}[j] \)'s in section 4.

### 3.1 Time complexity

Recall that we defined \( E[J](k) \) as the expected mean minor allele frequency in a sample of size \( k \). By Corollary 7 the procedure in eq. (13) has expected time complexity \( O(nE[J](k)) \).
4 Lazy evaluation of dynamic programming rows

Figure 2 Using the example that at site \( i \), \( \phi_i(\alpha_i) = \{ b_3 \} \), we illustrate the number of arithmetic operations used in i) the conventional \( O(nk) \) Li and Stephens HMM recurrence relations ii) Our procedure specified in equation (13). Black lines correspond to arithmetic operations; operations which cannot be parallelized over \( j \) are colored yellow.

4.1 Eliminating redundant recurrence evaluations

The recurrence relations (8) are linear maps \( r_i[j] : \mathbb{R} \rightarrow \mathbb{R} \) of the form

\[
\begin{align*}
  r_i[j] : x_j &\mapsto \alpha_i x_j + \beta_i \\
\end{align*}
\]

Lemma 9. At each \( i \) there exist only two unique maps among the \( r_i[j] \).

Proof. Assume, without loss of generality, that \( \phi_i(a) = \text{Match}_i(a) \). Then define \( \mu_i^\circ = \mu_i \) and \( \mu_i^\bullet = 1 - \mu_i \). Then

\[
\begin{align*}
  r_i^\circ(x) &= \mu_i^\circ ((1 - k\rho)x + \rho S_i) \\
  r_i^\bullet(x) &= \mu_i^\bullet ((1 - k\rho)x + \rho S_i) = \frac{\mu_i^\circ \mu_i^\bullet}{\mu_i^\circ} S_i \\
\end{align*}
\]

If \( \phi_i(a) \neq \text{Match}_i(a) \) then the same is true with the definitions of \( \mu_i^\circ \) and \( \mu_i^\bullet \) switched. This lemma allows us to rewrite each \( r_{i_1 \rightarrow i_2}[j] \) as a binary vector of the form \( (\circ, \circ, \bullet, \ldots, \circ, \bullet, \circ) \).

4 This approach is known as lazy evaluation.
Algorithm 1: General approach to evaluating $p_{(\cdot)}[j]$ for a given haplotype $j$.

Calculate $p_1[j]$ and set $\ell \leftarrow 1$
for all sites $i > 1$
do
  if $r_{\ell \rightarrow i}[j] = (\circ, \circ, \ldots, \circ)$ then Do nothing
  if $r_{\ell \rightarrow i}[j] = (\circ, \circ, \ldots, \bullet)$ then Evaluate $p_i[j] = r_{\ell \rightarrow i}(p_\ell[j])$ and set $\ell \leftarrow i$

4.2 Equivalence classes of update map prefixes

Consider two different instances of the for loop in Algorithm 1, where the first is to evaluate $p_{(\cdot)}[j_1]$ and the second to evaluate $p_{(\cdot)}[j_2]$. Suppose that both are halted at the same step $i$, and suppose that at this step, the marker variable $\ell$ is the same for both of them. Then the sequences of $h_{j_1}$ and $h_{j_2}$ are identical between $\ell$ and $i$, and therefore

$$r_{\ell \rightarrow i}[j_1] = r_{\ell \rightarrow i}[j_2] = (\circ, \circ, \ldots, \circ)_{i-\ell}$$  \hspace{1cm} (19)

Therefore at each step $i$, we can divide the haplotype indices $j$ into equivalence classes $J[\ell]$ for which

1. The current marker variable $\ell$ is the same for all $j$ in this equivalence class
2. The map $r_{\ell \rightarrow i}[j]$ is the same for all $j$ in this equivalence class

And so we only need to calculate $r_{\ell \rightarrow i}[j]$ at most once per nonempty equivalence class $J[\ell]$, and for this map we write $r_{\ell \rightarrow i}$. The following lemma allows us to be even more efficient.

Lemma 10. If $i_1 < i_2 < i$, then $r_{i_1 \rightarrow i} = r_{i_2 \rightarrow i} \circ r_{i_1 \rightarrow i_2}$

Lemma 10 allows us to calculate intermediate prefixes of the maps $r_{\ell \rightarrow i}$ and extend them at a later time. To make this concrete, suppose that we have an index $\pi_\ell$ (“prefix”) where $\ell < \pi_\ell < i$. Then we can evaluate the prefix $r_{\ell \rightarrow \pi_\ell}$ of $r_{\ell \rightarrow i}$ knowing that $r_{\ell \rightarrow i} = r_{\pi_\ell \rightarrow \pi_\ell} \circ r_{\ell \rightarrow \pi_\ell}$ can be evaluated at a later time.

4.3 The lazy evaluation algorithm

Our full lazy evaluation algorithm stores the following state data at each step. The algorithm initialization is described in Algorithm 2 and the recurrence in Algorithm 3.

- The maps $j \mapsto J[\ell]$ from haplotype to its equivalence class; $\ell$ defined as the index at which $p_{(\cdot)}[j]$ was most recently calculated
- The maps $\ell \mapsto \pi_\ell$ mapping $\ell$ to the index $\pi_\ell$ for which a prefix $r_{\ell \rightarrow \pi_\ell}$ of $r_{\ell \rightarrow i}$ which was most recently calculated
- The maps $\ell \mapsto r_{\ell \rightarrow \pi_\ell}$ mapping $\ell$ to the prefix $r_{\ell \rightarrow \pi_\ell}$ of $r_{\ell \rightarrow i}$ which was most recently calculated
- The maps $r^1_\ell, r^2_\ell, \ldots, r^n_\ell$ from which all maps $r_{(\cdot) \mapsto (\cdot)}$ are formed

Calculating a closed form expression for the time complexity of the lazy evaluation algorithm 3 is not straightforward. It is easy to show that it bounded by $\mathcal{O}(nk)$, since the first loop is worst-case $\mathcal{O}(k)$. However, we find experimentally, this lazy evaluation component does not contribute to overall computational complexity. (See Fig. 6)
Algorithm 2: Lazy evaluation initialization.

**Input**: reference panel size \( k \) and active rows \( \phi_1(o_1) \)

All \( j \not\in \phi_1(o_1) \) are assigned to equivalence class \( J[0] \)

\[
\tau_{0 \rightarrow \pi_0} \leftarrow \tau_1^0 \quad \text{and} \quad \pi_0 \leftarrow 1
\]

All \( j \in \phi_1(o_1) \) are assigned to equivalence class \( J[1] \)

\[
\tau_{1 \rightarrow \pi_1} \leftarrow \text{identity} \quad \text{and} \quad \pi_1 \leftarrow 1
\]

Algorithm 3: Lazy evaluation recurrence.

**Input**: active rows \( \phi_i(o_i) \) and previous lazy evaluation state

\( J_{\text{active}} \leftarrow \) subset of \( \{ J[\ell] \}_{\ell \leq i} \) consisting of equivalence classes containing at least one index from the set of active indices \( \phi_i(o_i) \)

\( \ell_{\text{min}} \leftarrow \) smallest \( \ell \) such that \( J[\ell] \in J_{\text{active}} \)

\[
\sigma_{\ell - 1 \rightarrow i} \leftarrow \tau_1^\ell
\]

for \( \lambda = i - 1 \) to \( \ell_{\text{min}} \) do

\[
\sigma_{\lambda - 1 \rightarrow i} \leftarrow \sigma_{\lambda + 1 - 1 \rightarrow i} \circ r_{\lambda}
\]

for one \( \ell \) with \( \pi_{\ell} = \lambda \), \( J_{\text{active}} \leftarrow J_{\text{active}} \cup J[\ell] \)

for \( J[\ell] \in J_{\text{active}} \) with \( \pi_{\ell} \neq i \) do

for \( \ell_{\rightarrow \pi_{\ell}} \leftarrow \sigma_{\pi_{\ell} + 1 - 1 \rightarrow \ell} \circ r_{\ell \rightarrow \pi_{\ell}} \) and \( \pi_{\ell} \leftarrow i \)

for \( j \in \phi_i \) do

\[
\ell \leftarrow \text{index for which } j \in J[\ell]
\]

Evaluate \( p_{\ell}[j] = r_{\ell \rightarrow \pi_{\ell}}(p_{\ell}[j]) \) and reassign \( j \) to the new class \( J[i] \)

\[
\tau_{i \rightarrow \pi_i} \leftarrow \text{identity} \quad \text{and} \quad \pi_i \leftarrow i
\]

5 Results

5.1 Implementation

Our algorithm was implemented as a C++ library located at https://github.com/yoeirosen/sublinear-Li-Stephens Details of Algorithm 3 will be found there.

We also implemented the linear time monoploid Li and Stephens forward algorithm in C++ as to evaluate it on identical footing. Profiling was performed using a single Intel Xeon X7560 core running at 2.3 GHz on a shared memory machine. Our reference panels \( H \) were the phased haplotypes from the 1000 Genomes [2] phase 3 vcf records for chromosome 22 and subsamples thereof. Haplotypes \( o \) were randomly generated simulated descendants.

5.2 Minor allele frequency distribution for the 1000 Genomes dataset

We simulated haplotypes \( o \) of 1,000,000 bp length on chromosome 22 and recorded the sizes of the sets \( \phi_i(o_i) \) for \( k = 5008 \). These data produced a mean \( |\phi_i(o_i)| \) of 59.9, which is 1.2\% of the size of \( k \). We have plotted the distribution of \( |\phi_i(o_i)| \) which we observed from this experiment in (Fig. 4). It is skewed toward low frequencies; the minor allele is unique at 71\% of sites, and it is below 1\% frequency at 92\% of sites.

5.3 Comparison of our algorithm with the linear time forward algorithm

In order to compare the dependence of our algorithm’s runtime on haplotype panel size \( k \) against that of the standard linear LS forward algorithm, we measured the CPU time per
Figure 3 An illustration of the lazy evaluation states defined above as well as the steps of Algorithm 3. In this case, we have a lazy evaluation state at \( i = 5 \), which is updated with \( \phi_i(o_i) = \{ 2, 4 \} \). \( j = 3 \) is updated as well as specified by the first loop in the algorithm.

We also measured the time which our algorithm spent within its lazy evaluation subalgorithm. In the average case, the time complexity of our lazy evaluation subalgorithm does not contribute to the overall algebraic time complexity of the algorithm. (Fig. 6, right) The lazy evaluation runtime also contributes minimally to the total actual runtime of our algorithm. (Fig. 6, left)
5.4 Sparse haplotype encoding

5.4.1 Generating our sparse vectors

We generated the haplotype panel data structures from (§2) using the vcf-encoding tool vcf2slls which we provide. We built indices with multiallelic sites, which increases their time and memory profile relative to the results in (§5.2) but allows direct comparison to vcf records. Encoding of chromosome 22 was completed in 38 minutes on a single CPU core. Use of $M$ CPU cores will reduce runtime proportional to $M$.

5.4.2 Size of sparse haplotype index

In uncompressed form, our whole genome *.slls index for chromosome 22 of the 1000 genomes dataset was 285 MB in size versus 11 GB for the vcf record using uint16_t’s to encode haplotype ranks. When compressed with gzip, the same index was 67 MB in size versus 205 MB for the vcf record.

In the interest of speed (both for our algorithm and the $O(nk)$ algorithm) our experiments loaded entire chromosome sparse matrices into memory and stored haplotype indices as uint64_t’s. This requires on the order of 1 GB memory for chromosome 22. For long chromosomes or larger reference panels on low memory machines, algorithm can operate on sequential chunks of the reference panel.
6 Discussion and significance

To the best of our knowledge, ours is the first forward algorithm for any haplotype model to attain sublinear time complexity with respect to reference panel size. Our algorithms could be incorporated into haplotype inference strategies by interfacing with our C++ library. This opens the potential for tools which are tractable on haplotype reference panels at the scale of current 100,000 to 1,000,000+ sample sequencing projects.

6.1 Applications which use individual forward probabilities

Our algorithm attains its runtime specifically for the problem of calculating the single overall probability \( P(o|H, \rho, \mu) \) and does not compute all \( nk \) forward probabilities. We can prove that if \( m \) many specific forward probabilities are also required as output, and if the time complexity of our algorithm is \( O(\sum |\phi_i|) \), then the time complexity of the algorithm which also returns the \( m \) forward probabilities is \( O(\sum |\phi_i| + m) \).

In general, haplotype phasing or genotype imputation tools use stochastic traceback or other similar sampling algorithms. The standard algorithm for stochastic traceback samples states from the full posterior distribution and therefore requires all forward probabilities. The algorithm output and lower bound of its speed is therefore \( O(nk) \). The same is true for many applications of the forward-backward algorithm.

There are two possible approaches which might allow runtime sublinear in \( k \) for these applications. Using stochastic traceback as an example, first is to devise an \( O(f(m)) \) sampling algorithm which uses \( m = g(k) \) forward probabilities such that \( O(f \circ g(k)) < O(k) \). The second is to succinctly represent forward probabilities such that nested sums of the \( nk \) forward probabilities can be queried from \( O(\phi) < O(nk) \) data. This should be possible, perhaps using the positional Burrows-Wheeler transform [5] as in [11], since we have already devised a forward algorithm with this property for a different model in [13].

6.2 Generalizability of algorithm

The optimizations which we have made are not strictly specific to the monoploid Li and Stephens algorithm. Necessary conditions for our reduction in the time complexity of the recurrence relations are

▶ **Condition 1.** The number of distinct transition probabilities is bounded.

▶ **Condition 2.** The number of distinct emission probabilities is bounded.
Favourable conditions for efficient time complexity of the lazy evaluation algorithm are

1. **Condition 1.** The number of unique update maps added per step is bounded.
2. **Condition 2.** The update map extension operation is composition of matrices of bounded size. This can be generalized to a broad algebraic class\(^5\) of update operations provided that they have bounded runtime.

The reduction in time complexity of the recurrence relations depends on the Markov property, however we hypothesize that the delayed evaluation needs only the semi-Markov property.

### 6.2.1 Other haplotype forward algorithms

Our optimizations are of immediate interest for other haplotype copying models. The following related algorithms have been explored without implementation.

- **Example (Diploid Li and Stephens).** We have yet to implement this model but expect average runtime at least subquadratic in reference panel size \(k\). We build on the statement of the model and its optimizations in [9]. We have found the following recurrences which we believe will work when combined with a system of lazy evaluation algorithms:

\[
p_i[j_1,j_2] = \alpha p_{i-1}[j_1,j_2] + \beta (S_{i-1}(j_1) + S_{i-1}(j_2)) + \gamma_p S
\]

which use on the intermediate sums

\[
S_i := \alpha_c S_{i-1} + \beta_c \sum_{j \in \phi_i} S_{i-1}(j) + \gamma_c \sum_{(j_1,j_2) \in \phi_i^2} p_{i-1}[j_1,j_2]
\]

\[
O(|\phi_i|^2)
\]

where \(\alpha(\cdot), \beta(\cdot), \gamma(\cdot)\) depend only on the diploid genotype \(o_i\).

Implementing and verifying this extension of our algorithm will be among our next steps.

- **Example (Multipopulation Li and Stephens).** [4] We maintain separate sparse haplotype panel representations \(\phi_A^A(o_i)\) and \(\phi_B^B(o_i)\) and separate lazy evaluation mechanisms for the two populations \(A\) and \(B\). Expected runtime guarantees are similar.

This model, and versions for > 2 populations, will be important in large sequencing cohorts (such as NHLBI TOPMed) where assuming a single related population is unrealistic.

- **Example (More detailed mutation model).** It may also be desirable to model distinct mutation probabilities for different pairs of alleles at multiallelic sites. Runtime is worse than the biallelic model but remains average case sublinear.

- **Example (Sequence graph Li and Stephens analogue).** In [13] we described a hidden Markov model for a haplotype-copying with recombination but not mutation in the context of sequence graphs. Assuming we can decompose our graph into nested sites then we can achieve a fast forward algorithm with mutation.

- **Example (Semi-Markovian recombination model).** The lazy evaluation algorithm 3 may efficiently allow time-since-recombination dependent transition probabilities.

\(^5\) Specifically, any collection of operations forming a category in the sense of category theory
References


