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Authors

Heeney, Dustin D
Gareau, Mélanie G
Marco, Maria L

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Intestinal *Lactobacillus* in health and disease, a driver or just along for the ride?

Dustin D Heeney¹, Mélanie G Gareau² and Maria L Marco¹

Metagenomics and related methods have led to significant advances in our understanding of the human microbiome. Members of the genus *Lactobacillus*, although best understood for essential roles in food fermentations and applications as probiotics, have also come to the fore in a number of untargeted gut microbiome studies in humans and animals. Even though *Lactobacillus* is only a minor member of the human colonic microbiota, the proportions of those bacteria are frequently either positively or negatively correlated with human disease and chronic conditions. Recent findings on *Lactobacillus* species in human and animal microbiome research, together with the increased knowledge on probiotic and other ingested lactobacilli, have resulted in new perspectives on the importance of this genus to human health.

Addresses

¹Department of Food Science & Technology, University of California, Davis, USA

²Department of Anatomy, Physiology & Cell Biology, School of Veterinary Medicine, University of California, Davis, USA

Corresponding author: Marco, Maria L (mmarco@ucdavis.edu)

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Introduction

Members of the genus *Lactobacillus* were long thought to be among the most abundant microorganisms in the human gastrointestinal (GI) tract and associated with good intestinal health. Following the development of culture-independent, DNA-sequence analysis methods, the numbers of autochthonous *Lactobacillus* were adjusted to $\leq 1\%$ of the total bacterial population in the distal human gut. One consequence of this change is that the relevance of this genus to human health has come under scrutiny. In contrast, there is increased acceptance of the application of allochthonous probiotic *Lactobacillus* in fermented foods and supplements as probiotics to maintain health and prevent and treat disease [1,2]. Although human studies frequently show a benefit with probiotic

administration [3], the importance of intestinal *Lactobacillus* remains under question.

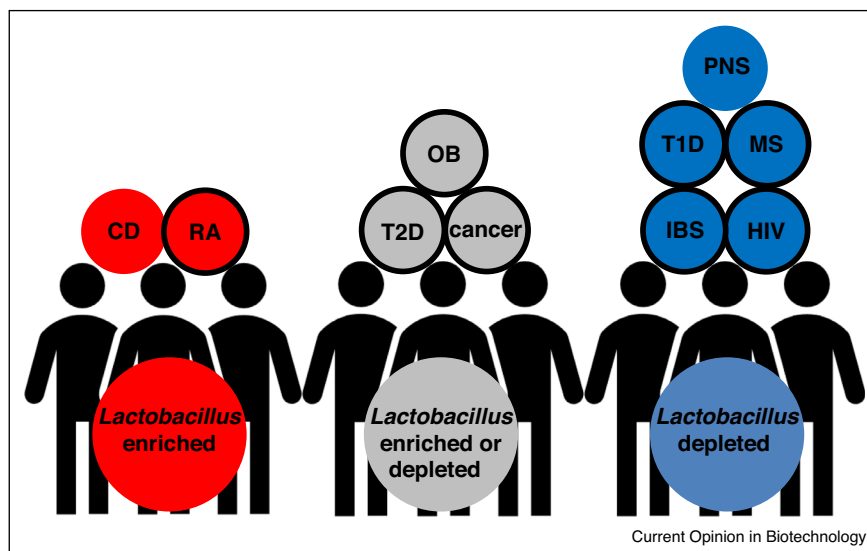
Human disease is increasingly correlated with fecal microbiota composition. Similarly, intestinal bacteria are frequently correlated with numerous other host (genetics, age) and environmental (diet, medication) factors. Such associations have been useful for identifying pathobionts associated with disease as well as taxa such as *Faecalibacterium prausnitzii* and *Akkermansia muciniphila* as beneficial members of the indigenous microbiota. Similarly, a number of recent publications in which culture-independent methods were employed (e.g. 16S rRNA gene amplicon sequencing) identified *Lactobacillus* as being significantly enriched in the distal gut during either health or disease (Figure 1 and Table 1). Because these approaches are largely untargeted, the outcomes provide an unbiased perspective on the relative importance of this genus weighed against other bacterial inhabitants of the GI tract. This review will address findings on the diversity and abundance of intestinal *Lactobacillus* resulting from gut microbiome studies and emerging mechanistic evidence of endogenous and ingested (probiotic) *Lactobacillus* species in the GI tract.

Abundance and diversity of intestinal *Lactobacillus*

Lactobacillus species have been isolated from the entirety of the human GI tract (oral cavity to feces) as well as the skin and vagina [4,5]. This genus is estimated to constitute 6% of the total bacterial cell numbers in the human duodenum [6] and approximately 0.3% of all bacteria in the colon [4] (Figure 2). These levels are similar to the numbers of lactobacilli found in pigs, ranging from 5% to 0.1% of total bacteria in the proximal [7] and distal [8] gut, respectively. *Lactobacillus* was found in higher quantities in rhesus macaques (up to 30% and 10% of all bacteria in the small and large intestine, respectively) [9]. Proportions of *Lactobacillus* in rodent models ranged between 30% and 60% of bacterial numbers in the ileum and approximately 25% in the colon [10,11] (Figure 2). *Lactobacillus* can also dominate the human vaginal microbiota (90 to 100% of total bacteria present) and is found on the skin, but in much lower relative abundance [5] (Figure 2).

Only a few out of the >200 known *Lactobacillus* species have been consistently and repeatedly associated with the human GI tract. Recently, this number was increased to over 50 *Lactobacillus* species that were repeatedly detected in the stools of healthy volunteers [12*]. The

Figure 1



Alteration of intestinal *Lactobacillus* in health and disease. Blue circles indicate *Lactobacillus* is depleted in disease compared to healthy controls. Red circles indicate *Lactobacillus* is increased in disease. Gray circles indicate *Lactobacillus* levels were found to be either increased or decreased with disease, depending on the study. Circles with black edges indicate a benefit for consumption of probiotics for treating disease. CD = Crohn's disease, RA = rheumatoid arthritis, OB = obesity, T2D = type 2 diabetes, IBS = irritable bowel syndrome, T1D = type 1 diabetes, PNS = prenatal stress, HIV = human immunodeficiency virus, MS = multiple sclerosis.

most abundant lactobacilli included *L. casei*, *L. delbrueckii*, *L. murinus*, *L. plantarum*, *L. rhamnosus*, and *L. ruminus*. Some of these species (e.g. *L. rhamnosus* and *L. murinus*) are rarely isolated from environments outside the intestine and are considered gut-autochthonous microorganisms. Other mucosal sites are colonized by distinct species (e.g. *L. crispatus* in the vagina) [13^{*}]. There also appears to be host-specificity among some *Lactobacillus* species, as shown for lineages of *L. reuteri* [14].

Infectious disease

Both human immunodeficiency virus (HIV)-infected humans and simian immunodeficiency virus (SIV)-infected rhesus macaques harbor reduced numbers of intestinal *Lactobacillus* [15,16] (Table 1). *Lactobacillus* depletion in rhesus macaques was associated with the loss of gut barrier-promoting T-helper 17 (Th17) cells and increased microbial translocation [16]. The potential of *Lactobacillus* to prevent or reverse intestinal damage during infection was demonstrated with the reduced interleukin-1 β -mediated inflammation and improved barrier function upon inoculation of *L. plantarum* directly into ileal loops of SIV+ macaques shortly after SIV infection [17]. The intestinal epithelium in healthy animals responded similarly to *L. plantarum*, consistent with the finding that the ileal transcriptomes of *L. plantarum* were indistinguishable between SIV+ and SIV- animals [18]. In human populations, HIV+ patients on a multi-strain probiotic supplement containing strains of *Lactobacillus* and other genera contained higher numbers of memory Th17 cells in peripheral blood and in the intestine, and

histological examination of colonic biopsies indicated increased intestinal barrier function [19].

Several recent animal studies have indicated a broader role for *Lactobacillus* in prevention and resolution of infectious disease. Tryptophan metabolites (indole aldehydes) produced by indigenous *L. reuteri* strains activate host aryl hydrocarbon receptors (AHR) to promote gut and vaginal epithelial barrier and antimicrobial responses required for limiting the expansion of *Candida albicans*, an opportunistic pathogen [20^{**}]. Autochthonous *Lactobacillus* might also have a role in the resolution of infectious disease and recovery of immune homeostasis. Although *Yersinia enterocolitica* infection was cleared from toll-like receptor 1 (TLR1) knockout mice, the intestine was activated toward an inflammatory phenotype and the gut microbiota was enriched with *Desulfovibrionaceae* while containing lower numbers of *Lactobacillus* [21^{**}]. Oral gavage with *L. reuteri* reduced anti-commensal antibodies, innate cytokines, and Th17 responses; thereby ameliorating immune hyper-reactivity [21^{**}]. Conversely, post-*Yersinia pseudotuberculosis* infection lactobacilli were cultured from enlarged gut-associated lymphoid tissue and were associated with chronic lymphadenopathy, indicating that these bacteria might contribute to chronic, immune hyper-reactivity [22].

Irritable bowel syndrome (IBS) and inflammatory bowel disease (IBD)

A meta-analysis of reports investigating the fecal microbiomes from IBS patients and healthy subjects concluded

Table 1

Recent human studies analyzing microbiomes in health and disease

Disease/condition	Study design ^a	<i>Lactobacillus</i> proportions ^b	<i>Lactobacillus</i> species ^c	Method ^d	Reference
IBS	Meta-analysis 13 studies	↓		qRT-PCR	[23]
	Meta-analysis 10 Chinese studies	↓		Fecal bacterial counts	[24]
	Meta-analysis 7 global studies	-		Fecal bacterial counts	[24]
CD	28 CD, 26 HC	↑		Metagenomics	[26]
	15 CD, 21 HC	↑		qRT-PCR	[27]
HIV	8 infected, 8 HC	↓		16S rRNA	[15]
Rheumatoid arthritis	77 RA, 80 HC	↑	<i>L. salivarius</i> , <i>L. ruminus</i> , <i>L. iners</i>	Metagenomics	[30*]
Type 1 diabetes	21 T1D, 32 HC	↓		16S rRNA	[34*]
	28 T1D, 27 HC	↓		16S rRNA microarray	[35]
Multiple sclerosis	31 MS, 36 HC	↓		16S rRNA	[37]
Obesity	15 obese, 17 HC	↓	<i>L. plantarum</i>	qRT-PCR	[45]
	30 obese, 24 OW, 30 HC	↑		qRT-PCR	[42]
	42 obese, 36 HC	-		16S rRNA	[43]
	67 obese, 67 HC	-		16S rRNA	[44]
Type 2 diabetes	53 T2D, 43 HC	↑	<i>L. gasseri</i>	16S rRNA	[46]
	93 Met+T2D, 106 Met-T2D, 554 HC	↓		Metagenomics	[47]
Colon cancer	Systematic review 31 studies	↓		16S rRNA	[53]
Breast cancer	17 BC, 16 BD	↑		16S rRNA	[55]
Head and neck squamous cell cancer	17 HNSCC, 25 HC	↑		16S rRNA	[56]
Prenatal stress	56 mother-infant pairs	↓		16S rRNA	[57*]

^a CD = Crohn's disease, HC = healthy control, OW = overweight, Met+T2D = metformin treated type 2 diabetes patients, Met-T2D = patients not treated with metformin, BC = breast cancer, BD = benign disease, HNSCC = head and neck squamous cell cancer.

^b Proportions or numbers of *Lactobacillus* were lower (↓), higher (↑), or unchanged (-) in individuals with disease or chronic conditions compared to healthy individuals.

^c Species identification was conducted by authors and not assessed by reviewers.

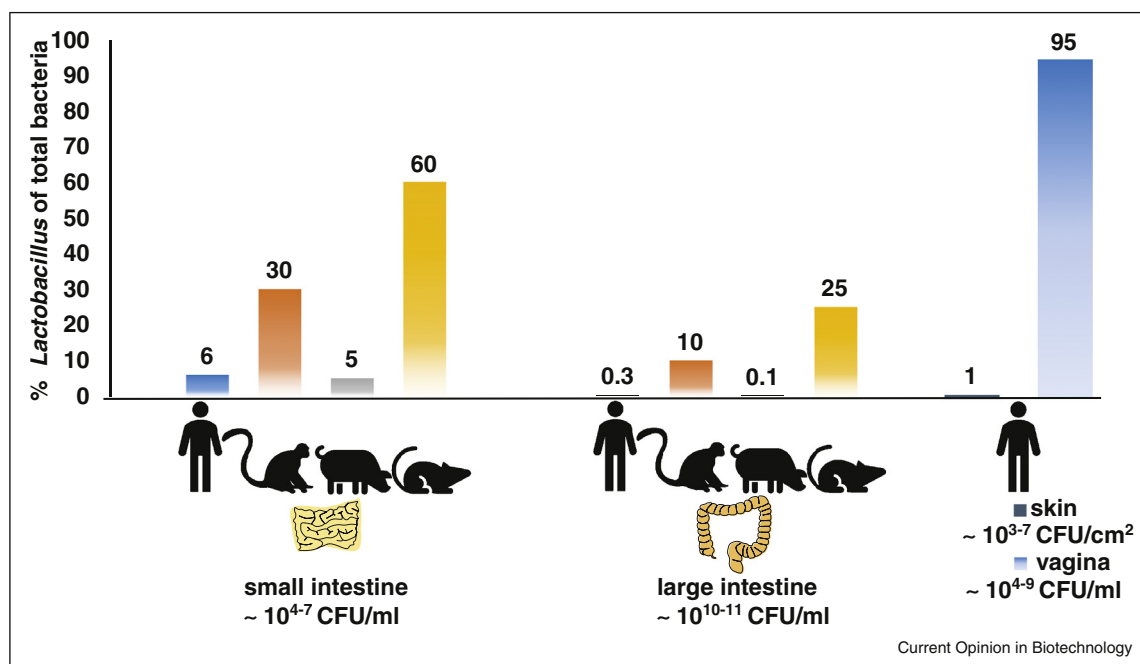
^d qRT-PCR = quantitative real time-PCR of 16S rRNA, metagenomics = shotgun whole genome sequencing, 16S rRNA = 16S ribosomal RNA sequencing.

Lactobacillus was depleted in diarrhea-dominant, IBS patients [23] (Table 1). Another meta-analysis of IBS cohort studies determined that intestinal *Lactobacillus* was depleted in all cases of IBS in Chinese patients, but this association was not found or was reversed in patients from other countries [24]. Consistent with these results, meta-analysis of probiotic intervention studies (43 randomized controlled trials (RCTs)) for treatment of IBS concluded that multi-species probiotics diminish symptoms (abdominal pain, bloating, and flatulence scores) [25].

Conversely, intestinal abundance of *Lactobacillus* and other genera including *Bifidobacterium* were recently positively correlated with Crohn's disease (CD) patients

[26,27] (Table 1). In both studies, *Lactobacillus* enrichment coincided with depletion of *F. prausnitzii*. Whether *Lactobacillus* is participating in disease or is simply adapted to survive the pro-inflammatory gut environment is not known. These findings contrast with ulcerative colitis (UC) in which probiotic *Lactobacillus* consumption has been with improved clinical symptoms [28]. *Lactobacillus* might be particularly supportive in CD and UC patients with caspase recruitment domain family member 9 (CARD9) risk alleles whose microbiome has a reduced production of AHR ligands [29**]. Consistent with this possibility, intestinal inflammation in CARD9 knockout mice was attenuated after inoculation of mice with *Lactobacillus* strains capable of metabolizing tryptophan [29**].

Figure 2



Relative abundance of *Lactobacillus* in humans and animals. Numbers underneath anatomical locations indicate estimates for total bacterial community cell numbers.

Rheumatoid arthritis (RA)

The intestinal microbiota of patients with severe and early onset RA were shown to have increased proportions of *L. salivarius*, *L. ruminus*, and *L. iners* when compared to healthy, age-matched individuals [30*] (Table 1). Enrichment of *Lactobacillus* spp. was also observed in collagen-induced, arthritic mice [31]. These results are in opposition to recent RCTs of probiotics in RA patients. In one study, patients consuming *L. casei* had reduced disease activity scores, higher quantities of serum IL-10, and decreased levels of serum TNF α , IL-6 and IL-12 [32]. The other RCT concluded that a mixed strain probiotic supplement significantly improved disease activity scores and lowered levels of serum C-reactive protein (CRP) [33]. Such findings might indicate species or strain-specific differences between autochthonous and allochthonous *Lactobacillus* on RA disease activity.

Type 1 diabetes (T1D)

The proportions of *Lactobacillus* were lower in adults with T1D than healthy, first-degree relatives and unrelated healthy individuals according to untargeted 16S rRNA gene analysis [34*] (Table 1). A similar reduction in *Lactobacillus* was observed in children with T1D [35] (Table 1). Interestingly, children exposed to probiotic *Lactobacillus* early in life were found to have a significantly reduced risk of developing islet autoimmunity [36]. It is not yet understood how *Lactobacillus* could be regulating islet beta-cell autoimmunity, although it has been

suggested that a lack of intestinal lactate-producing bacteria depletes butyrate-producing taxa leading to aberrant immune responses [35].

Multiple sclerosis (MS)

A cohort study found that the relative abundance of intestinal *Lactobacillus* was lower in MS patients compared to healthy adults [37] (Table 1). Similar depletions in intestinal *Lactobacillus* were observed in a preclinical, rodent model of MS [38]. Consistent with a benefit of *Lactobacillus* in this autoimmune disease were the findings from a recent RCT of MS patients, whereby consumption of a multi-species probiotic improved the expanded disability status score, self-reported depression, anxiety and stress, as well as decreased serum CRP [39]. Because circulating levels of AHR ligands are lower in MS patients compared to healthy adults [40], *Lactobacillus* might be useful for the maintenance or replenishment of these compounds. To this regard, *Lactobacillus*-produced indole aldehydes had a potent anti-inflammatory effect on brain glial cells (astrocytes) to limit central nervous system inflammation in a mouse model of human MS [41*].

Obesity and type 2 diabetes (T2D)

There are conflicting reports on the association of intestinal *Lactobacillus* with obesity in humans [42–45] (Table 1). Likewise, initial studies found increased levels of *Lactobacillus* in patients with T2D [46], although this trend was eliminated or reversed when controlling for

metformin treatment [47] (Table 1). Moreover, meta-analysis of RCT studies found that probiotic *Lactobacillus* improved weight management outcomes in obese adults [48]. Consumption of yogurt and other dairy products fermented by *Lactobacillus* is also correlated with protection from T2D and obesity (recently reviewed in [2]).

Because *Lactobacillus* species appear to be either associated with weight gain or weight loss [49], the disparate findings among obese individuals might be due to genetic differences among the lactobacilli. Strain and species distinctions could result in variations in carbohydrate metabolism and production of fermentation end-products, such as lactate [50]. The synthesis of bile salt hydrolases is another distinguishing feature of some *Lactobacillus* species, an activity that can alter the activation of farnesoid x receptor (FXR) signaling and hepatic lipid metabolism [51,52].

Cancer

In a systematic review of thirty-one studies, *Lactobacillus* along with a limited number of butyrogenic genera were consistently diminished in colorectal cancer patients [53] (Table 1). Preventative and therapeutic roles of *Lactobacillus* in cancer are supported in studies with preclinical, rodent models, including a recently study in which a multi-strain probiotic altered Th-cell polarization away from Th17 cells in a mouse model of hepatocellular carcinoma [54]. However, *Lactobacillus* might not always be beneficial in certain extra-intestinal sites as shown by the higher levels of *Lactobacillus* in malignant breast cancer compared to benign-disease tissues [55]. There was also a positive association between the levels of this genus in the oral microbiome and head and neck squamous cell carcinoma [56] (Table 1).

Cognitive development and behavior

Maternal prenatal stress might influence the infant microbiome, potentially damaging cognitive development. In humans, prenatal cortisol concentrations were inversely correlated with infant levels of intestinal *Lactobacillus* and *Lactococcus*, whereas *Proteobacteria* were enriched [57*] (Table 1). A comparable depletion of *Lactobacillus* was observed in rodent models of prenatal stress, with the microbiome of the offspring remaining disrupted into adulthood [58]. Prenatal low-dose penicillin [59] or high fat diet [60**] could similarly induce long-term dysbiosis and behavioral deficits in mice. These deficits could be prevented by concurrent administration of *Lactobacillus*-containing probiotics to the dam [59] or by indigenous *L. reuteri* to offspring [60**].

In adult mouse models of microbiota–gut–brain axis deficits, administration of *Lactobacillus* probiotics was found to beneficially impact both cognition and colonic function, while reversing intestinal dysbiosis [61,62]. Such results might also be relevant to emotional disorders

and this is supported in probiotics studies which have indicated that probiotic *Lactobacillus* might improve symptoms of human depression [63,64]. Therefore, beneficially modulating the microbiota using *Lactobacillus* can impact the microbiota–gut–brain axis and should be more thoroughly studied in human mother–infant cohorts.

Conclusions

Our increased understanding of intestinal *Lactobacillus* from untargeted microbiome studies supports the premise that general properties conferred by this genus have far-reaching consequences on human health. This possibility could be further investigated via studies designed to determine the proximity of *Lactobacillus* to the intestinal epithelium or which focus attention on other sites on the body wherein members of this genus can constitute the majority of bacteria present (e.g. vagina). Moreover, strain and/or species-specific differences (e.g. tryptophan and bile metabolism) might be useful to explain variations in the involvement of this genus, either in the prevention or mitigation of disease or, alternatively, as a contributing factor to disease outcomes. Furthermore, the notable variation in intestinal abundance of this genus between healthy and diseased, or health-compromised, individuals indicates that *Lactobacillus*, or at least certain species or genotypes of *Lactobacillus*, might be useful gut biomarkers. These considerations can also inform the improved development and use of probiotics in different human populations.

Conflicts of interest

None.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, Morelli L, Canani RB, Flint HJ, Salminen S *et al.*: **Expert consensus document: the International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic.** *Nat Rev Gastroenterol Hepatol* 2014, **11**:506–514.
 2. Marco ML, Heeney D, Binda S, Cifelli CJ, Cotter PD, Foligné B, Gänzle M, Kort R, Pasin G, Pihlanto A *et al.*: **Health benefits of fermented foods: microbiota and beyond.** *Curr Opin Biotechnol* 2017, **44**:94–102.
 3. Floch MH, Walker WA, Sanders E, Nieuwdorp M, Kim AS, Brenner DA, Qamar AA, Miloh TA, Guarino A, Guslandi M *et al.*: **Recommendations for probiotic use — 2015 update**

- proceedings and consensus opinion.** *J Clin Gastroenterol* 2015, **49**:S69-S73.
4. Almonacid DE, Kraal L, Ossandon FJ, Budovskaya YV, Cardenas JP, Bik EM, Goddard AD, Richman J, Apte ZS: **16S rRNA gene sequencing and healthy reference ranges for 28 clinically relevant microbial taxa from the human gut microbiome.** *PLOS ONE* 2017, **12**:1-15.
 5. Chu DM, Ma J, Prince AL, Antony KM, Seferovic MD, Aagaard KM: **Maturation of the infant microbiome community structure and function across multiple body sites and in relation to mode of delivery.** *Nat Med* 2017, **23**:314-323.
 6. Nistal E, Caminero A, Herran AR, Perez-Andres J, Vivas S, Ruiz De Morales JM, Saenz de Miera LE, Casqueiro J: **Study of duodenal bacterial communities by 16S rRNA gene analysis in adults with active celiac disease vs non-celiac disease controls.** *J Appl Microbiol* 2015, **120**:1691-1700.
 7. Fan P, Liu P, Song P, Chen X, Ma X: **Moderate dietary protein restriction alters the composition of gut microbiota and improves ileal barrier function in adult pig model.** *Sci Rep* 2017, **7**:1-12.
 8. Slifkier MJ, Friendship RM, Weese JS: **Longitudinal study of the early-life fecal and nasal microbiotas of the domestic pig.** *BMC Microbiol* 2015, **15**:1-12.
 9. Mohan M, Chow CET, Ryan CN, Chan LS, Dufour J, Aye PP, Blanchard J, Moehs CP, Sestak K: **Dietary gluten-induced gut dysbiosis is accompanied by selective upregulation of microRNAs with intestinal tight junction and bacteria-binding motifs in rhesus macaque model of celiac disease.** *Nutrients* 2016, **8**:1-18.
 10. Li D, Chen H, Mao B, Yang Q, Zhao J, Gu Z, Zhang H, Chen YQ, Chen W: **Microbial biogeography and core microbiota of the rat digestive tract.** *Sci Rep* 2017, **7**:1-16.
 11. Morikawa M, Tsujibe S, Kiyoshima-Shibata J, Watanabe Y, Kato-Nagaoka N, Shida K, Matsumoto S: **Microbiota of the small intestine is selectively engulfed by phagocytes of the lamina propria and Peyer's patches.** *PLOS ONE* 2016, **11**:1-16.
 12. Rossi M, Martinez-Martinez D, Amaretti A, Ulrici A, Raimondi S, Moya A: **Mining metagenomic whole genome sequences revealed subdominant but constant *Lactobacillus* population in the human gut microbiota.** *Environ Microbiol Rep* 2016, **8**:399-406.
- Over 50 *Lactobacillus* species were repeatedly detected in quantities up to 10⁹ cells/g in the stool of a human subject. Metagenomics enabled the identification of strain-level distinctions among the lactobacilli and individual *Lactobacillus* genotypes were consistently detected in the stool over a time span of nearly two years.
13. Gosmann C, Anahtar MN, Handley SA, Farcasanu M, Abu-Ali G, Bowman BA, Padavattan N, Desai C, Droit L, Moodley A *et al.*: ***Lactobacillus*-deficient cervicovaginal bacterial communities are associated with increased HIV acquisition in young South African women.** *Immunity* 2017, **46**:29-37.
- This prospective study found that women with low-diversity vaginal bacterial communities dominated by *Lactobacillus crispatus* had a lower risk of acquiring HIV and had decreased numbers of activated mucosal CD4⁺ T cells compared to women with diverse genital bacterial communities. The results were supported by the lower numbers of activated CD4⁺ T cells in the genital mucosa of germ-free mice intravaginally-inoculated with *L. crispatus* compared with HIV-associated bacterial taxa.
14. Duar RM, Frese SA, Fernando SC, Burkey TE, Tasseva G, Peterson DA, Blom J, Wenzel CQ, Szymanski CM, Walter J: **Experimental determination of host adaptation of *Lactobacillus reuteri* to different vertebrate species.** *Appl Environ Microbiol* 2017, **83**:1-44.
 15. Yang L, Poles MA, Fisch GS, Ma Y, Nossa C, Phelan JA, Pei Z: **HIV-induced immunosuppression is associated with colonization of the proximal gut by environmental bacteria.** *AIDS* 2016, **30**:19-29.
 16. Vujkovic-Cvijin I, Swainson LA, Chu SN, Ortiz AM, Santee CA, Petriello A, Dunham RM, Fadrosch DW, Lin DL, Faruqi AA *et al.*: **Gut-resident *Lactobacillus* abundance associates with IDO1 inhibition and Th17 dynamics in SIV-infected macaques.** *Cell Rep* 2015, **13**:1589-1597.
 17. Hirao LA, Grishina I, Bourry O, Hu WK, Somrit M, Sankaran-Walters S, Gaulke CA, Fenton AN, Li JA, Crawford RW *et al.*: **Early mucosal sensing of SIV infection by paneth cells induces IL-1 β production and initiates gut epithelial disruption.** *PLoS Pathog* 2014, **10**:1-15.
 18. Golomb BL, Hirao LA, Dandekar S, Marco ML: **Gene expression of *Lactobacillus plantarum* and the commensal microbiota in the ileum of healthy and early SIV-infected rhesus macaques.** *Sci Rep* 2016, **6**:1-10.
 19. d'Ettoire G, Rossi G, Scagnolari C, Andreotti M, Giustini N, Serafino S, Schietroma I, Scheri GC, Fard SN, Trinchieri V *et al.*: **Probiotic supplementation promotes a reduction in T-cell activation, an increase in Th17 frequencies, and a recovery of intestinal epithelium integrity and mitochondrial morphology in ART-treated HIV-1-positive patients.** *Immunity Inflamm Dis* 2017 <http://dx.doi.org/10.1002/iid3.160>.
 20. Zelante T, Iannitti RG, Cunha C, DeLuca A, Giovannini G, Pieraccini G, Zecchi R, D'Angelo C, Massi-Benedetti C, Fallarino F *et al.*: **Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22.** *Immunity* 2013, **39**:372-385.
- This early report singled out intestinal and vaginal lactobacilli for a potent ability to produce tryptophan metabolites that activate host-AHRs in epithelium tissue. The activation of epithelial AHRs induces the production of a myriad of antimicrobial compounds and was found to be essential for control of opportunistic pathogens. Importantly, AHR knock-out mice infected with *Candida albicans* could not be rescued by introduction of tryptophan-metabolizing lactobacilli.
21. Kamdar K, Khakpour S, Chen J, Leone V, Brulc J, Mangatu T, Antonopoulos DA, Chang EB, Kahn SA, Kirschner BS *et al.*: **Genetic and metabolic signals during acute enteric bacterial infection alter the microbiota and drive progression to chronic inflammatory disease.** *Cell Host Microbe* 2016, **19**:21-31.
- In TLR1 knockout mice the acute intestinal infection of *Yersinia enterocolitica* destabilizes mucosal immunity allowing blooms of δ -*Proteobacteria* and diminished levels of other taxa, including lactobacilli. Chronic immune responses against the commensal microbiota were observed two months postinfection. Supplementation with *L. reuteri* was sufficient to revert the immune system towards homeostasis.
22. da Fonseca DM, Hand TW, Han S-J, Gerner MY, Zaretsky AG, Byrd AL, Harrison OJ, Ortiz AM, Quinones M, Trinchieri G *et al.*: **Microbiota-dependent sequelae of acute infection compromise tissue-specific immunity.** *Cell* 2017, **163**:354-366.
 23. Liu H-N, Wu H, Chen Y-Z, Chen Y-J, Shen X-Z, Liu T-T: **Altered molecular signature of intestinal microbiota in irritable bowel syndrome patients compared with healthy controls: a systematic review and meta-analysis.** *Dig Liver Dis* 2017, **49**:331-337.
 24. Zhuang X, Xiong L, Li L, Li M, Chen M: **Alterations of gut microbiota in patients with irritable bowel syndrome: a systematic review and meta-analysis.** *J Gastroenterol Hepatol* 2017, **32**:28-38.
 25. Ford AC, Quigley EMM, Lacy BE, Lembo AJ, Saito YA, Schiller LR, Soffer EE, Spiegel BMR, Moayyedi P: **Efficacy of prebiotics, probiotics, and synbiotics in irritable bowel syndrome and chronic idiopathic constipation: systematic review and meta-analysis.** *Am J Gastroenterol* 2014, **109**:1547-1561.
 26. Lewis JD, Chen EZ, Baldassano RN, Otley AR, Griffiths AM, Lee D, Bittinger K, Bailey A, Friedman ES, Hoffmann C *et al.*: **Inflammation, antibiotics, and diet as environmental stressors of the gut microbiome in pediatric Crohn's disease.** *Cell Host Microbe* 2015, **18**:489-500.
 27. Wang W, Chen L, Zhou R, Wang X, Song L, Huang S, Wang G, Xia B: **Increased proportions of *Bifidobacterium* and the *Lactobacillus* group and loss of butyrate-producing bacteria in inflammatory bowel disease.** *J Clin Microbiol* 2014, **52**:398-406.
 28. Ganji-Arjenaki M, Rafieian-Kopaei M: **Probiotics are a good choice in remission of inflammatory bowel diseases: a meta-analysis and systematic review.** *J Cell Physiol* 2017 <http://dx.doi.org/10.1002/jcp.25911>.
 29. Lamas B, Richard ML, Leducq V, Pham H-P, Michel M-L, Da Costa G, Bridonneau C, Jegou S, Hoffmann TW, Natividad JM *et al.*: **CARD9 impacts colitis by altering gut microbiota**

metabolism of tryptophan into aryl hydrocarbon receptor ligands. *Nat Med* 2016, **22**:598-605.

Human single nucleotide polymorphism (SNP) studies indicate caspase recruitment domain family member 9 (CARD9) alleles are associated with inflammatory bowel disease and that this risk allele may influence the development of a pro-inflammatory microbiome. By using CARD9 knockout mice, this report links CARD9 deficiencies to an altered intestinal microbiome with reduced production of AHR ligands. *L. reuteri* was identified as a significantly depleted taxon and replenishment of this species improved symptoms of colitis in CARD9 knockout mice.

30. Zhang X, Zhang D, Jia H, Feng Q, Wang D, Liang D, Wu X, Li J, Tang L, Li Y *et al.*: **The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment.** *Nat Med* 2015, **21**:895-905.

This human study utilizing metagenomic sequencing on the fecal microbiota identified several species of intestinal *Lactobacillus* species that were elevated in the oral and fecal microbiota of RA patients compared with healthy subjects.

31. Liu X, Zeng B, Zhang J, Li W, Mou F, Wang H, Zou Q, Zhong B, Wu L, Wei H *et al.*: **Role of the gut microbiome in modulating arthritis progression in mice.** *Sci Rep* 2016, **6**:1-11.
32. Vaghef-Mehrabany E, Alipour B, Homayouni-Rad A, Sharif S-K, Asghari-Jafarabadi M, Zavvari S: **Probiotic supplementation improves inflammatory status in patients with rheumatoid arthritis.** *Nutrition* 2014, **30**:430-435.
33. Zamani B, Golkar HR, Farshbar S, Emadi-Baygi M, Tajabadi-Ebrahimi M, Jafari P, Akhavan R, Taghizadeh M, Memarzadeh MR, Asemi Z: **Clinical and metabolic response to probiotic supplementation in patients with rheumatoid arthritis: a randomized, double-blind, placebo-controlled trial.** *Int J Rheum Dis* 2016, **19**:869-879.
34. Alkanani AK, Hara N, Gottlieb PA, Ir D, Robertson CE, Wagner BD, Frank DN, Zipris D: **Alterations in intestinal microbiota correlate with susceptibility to type 1 diabetes.** *Diabetes* 2015, **64**:3510-3520.

In this cohort study, the genetic component of T1D was partially controlled for by recruiting T1D patients and their healthy, first-degree relatives. The fecal microbiota was assessed by 16S rRNA sequencing as an environmental factor associated with disease. *Lactobacillus* proportions were significantly depleted in T1D patients.

35. de Goffau MC, Fuentes S, Van Den Bogert B, Honkanen H, De Vos WM, Welling GW, Hyöty H, Harmsen HJM: **Aberrant gut microbiota composition at the onset of type 1 diabetes in young children.** *Diabetologia* 2014, **57**:1569-1577.
36. Uusitalo U, Liu X, Yang J, Hummel S, Butterworth M, Rewers M, Hagopian W, She J, Simell O, Toppari J *et al.*: **Association of early exposure of probiotics and islet autoimmunity in the TEDDY study.** *JAMA Pediatr* 2016, **170**:20-28.
37. Chen J, Chia N, Kalari KR, Yao JZ, Novotna M, Soldan MMP, Luckey DH, Marietta EV, Jeraldo PR, Chen X *et al.*: **Multiple sclerosis patients have a distinct gut microbiota compared to healthy controls.** *Sci Rep* 2016, **6**:1-10.
38. Stanisavljević S, Lukić J, Soković S, Mihajlović S, Stojković MM, Mijlković D, Golić N: **Correlation of gut microbiota composition with resistance to experimental autoimmune encephalomyelitis in rats.** *Front Microbiol* 2016, **7**:1-12.
39. Kouchaki E, Tamtaji OR, Salami M, Bahmani F, Daneshvar Kakhaki R, Akbari E, Tajabadi-Ebrahimi M, Jafari P, Asemi Z: **Clinical and metabolic response to probiotic supplementation in patients with multiple sclerosis: a randomized, double-blind, placebo-controlled trial.** *Clin Nutr* 2016 <http://dx.doi.org/10.1016/j.clnu.2016.08.015>.
40. Lim CK, Bilgin A, Lovejoy DB, Tan V, Bustamante S, Taylor BV, Bessede A, Brew BJ, Guillemin GJ: **Kynurenine pathway metabolomics predicts and provides mechanistic insight into multiple sclerosis progression.** *Sci Rep* 2017, **7**:1-9.
41. Rothhammer V, Mascanfroni I, Bunse L, Takenaka M, Kenison J, Mayo L, Chao C-C, Patel B, Yan R, Blain M *et al.*: **Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor.** *Nat Med* 2016, **22**:586-597.

This mouse study causally links lactobacilli to the production of AHR ligands and mitigation of experimentally induced central nervous system

inflammation. Reconstitution of an antibiotic-disrupted microbiome with *L. reuteri* capable of metabolizing tryptophan was sufficient to improve markers of disease.

42. Ignacio A, Fernandes MR, Rodrigues VAA, Groppo FC, Cardoso AL, Avila-Campos MJ, Nakano V: **Correlation between body mass index and faecal microbiota from children.** *Clin Microbiol Infect* 2016, **22**:1-8.
43. Riva A, Borgo F, Lassandro C, Verduci E, Morace G, Borghi E, Berry D: **Pediatric obesity is associated with an altered gut microbiota and discordant shifts in Firmicutes populations.** *Environ Microbiol* 2017, **19**:95-105.
44. Hu HJ, Park SG, Jang HB, Choi MG, Park KH, Kang JH, Park SI, Lee HJ, Cho SH: **Obesity alters the microbial community profile in Korean Adolescents.** *PLOS ONE* 2015, **10**:1-14.
45. Teixeira TFS, Grześkowiak M, Salminen S, Laitinen K, Bressan J, Gouveia Peluzio M, do C: **Faecal levels of *Bifidobacterium* and *Clostridium coccooides* but not plasma lipopolysaccharide are inversely related to insulin and HOMA index in women.** *Clin Nutr* 2013, **32**:1017-1022.
46. Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, Nielsen J, Bäckhed F: **Gut metagenome in European women with normal, impaired and diabetic glucose control.** *Nature* 2013, **498**:99-103.
47. Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, Prifti E, Vieira-Silva S, Gudmundsdottir V, Krogh Pedersen H *et al.*: **Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota.** *Nature* 2015, **528**:262-266.
48. Sáez-Lara MJ, Robles-Sanchez C, Ruiz-Ojeda FJ, Plaza-Diaz J, Gil A: **Effects of probiotics and synbiotics on obesity, insulin resistance syndrome, type 2 diabetes and non-alcoholic fatty liver disease: a review of human clinical trials.** *Int J Mol Sci* 2016, **17**:1-15.
49. Drissi F, Raoult D, Merhej V: **Metabolic role of lactobacilli in weight modification in humans and animals.** *Microb Pathog* 2016 <http://dx.doi.org/10.1016/j.micpath.2016.03.006>.
50. Le Roy CI, tepetova J, Sepp E, Songisepp E, Sandrine P, Mikelsaar M: **New insights into the impact of *Lactobacillus* population on host-bacteria metabolic interplay.** *Oncotarget* 2015, **6**:30545-30556.
51. Gonzalez FJ, Jiang C, Patterson AD: **An intestinal microbiota-farnesoid X receptor axis modulates metabolic disease.** *Gastroenterology* 2016, **151**:845-859.
52. Zhang L, Xie C, Nichols RG, Chan SHJ, Jiang C, Hao R, Smith PB, Cai J, Simons MN, Hatzakis E *et al.*: **Farnesoid X receptor signaling shapes the gut microbiota and controls hepatic lipid metabolism.** *mSystems* 2016, **1**:1-17.
53. Borges-Canha M, Portela-Cidade JP, Dinis-Ribeiro M, Leite-Moreira AF, Pimentel-Nunes P: **Role of colonic microbiota in colorectal carcinogenesis: a systematic review.** *Rev Esp Enferm Dig* 2015, **107**:659-671.
54. Li J, Sung CYJ, Lee N, Ni Y, Pihlajamäki J, Panagiotou G, El-Nezami H: **Probiotics modulated gut microbiota suppresses hepatocellular carcinoma growth in mice.** *Proc Natl Acad Sci U S A* 2016, **113**:E1306-E1315.
55. Hieken TJ, Chen J, Hoskin TL, Walther-Antonio M, Johnson S, Ramaker S, Xiao J, Radisky DC, Knutson KL, Kalari KR *et al.*: **The microbiome of aseptically collected human breast tissue in benign and malignant disease.** *Sci Rep* 2016, **6**:1-10.
56. Guerrero-Preston R, Godoy-Vitorino F, Jedlicka A, Rodríguez-Hilario A, González H, Bondy J, Lawson F, Folawiyo O, Michailidi C, Dziedzic A *et al.*: **16S rRNA amplicon sequencing identifies microbiota associated with oral cancer, human papillomavirus infection and surgical treatment.** *Oncotarget* 2016, **7**:51320-51334.
57. Zijlmans MAC, Korpela K, Riksen-Walraven JM, de Vos WM, de Weerth C: **Maternal prenatal stress is associated with the infant intestinal microbiota.** *Psychoneuroendocrinology* 2015, **53**:233-245.

16S rRNA sequencing of fecal microbiomes from a mother–infant cohort before and after birth allowed Zijlmans and colleagues to link maternal prenatal stress to alterations in the infant microbiome. Pathobionts were enriched in infants whose mothers were more stressed. *Lactobacillus* cell numbers were depleted.

58. Golubeva AV, Crampton S, Desbonnet L, Edge D, O'Sullivan O, Lomasney KW, Zhdanov AV, Crispie F, Moloney RD, Borre YE *et al.*: **Prenatal stress-induced alterations in major physiological systems correlate with gut microbiota composition in adulthood.** *Psychoneuroendocrinology* 2015, **60**:58-74.
59. Leclercq S, Mian FM, Stanisz AM, Bindels LB, Cambier E, Ben-Amram H, Koren O, Forsythe P, Bienenstock J: **Low-dose penicillin in early life induces long-term changes in murine gut microbiota, brain cytokines and behavior.** *Nat Commun* 2017, **8**:1-12.
60. Buffington SA, Di Prisco GV, Auchtung TA, Ajami NJ, Petrosino JF, ●● Costa-Mattioli M: **Microbial reconstitution reverses maternal diet-induced social and synaptic deficits in offspring.** *Cell* 2016, **165**:1762-1775.

Maternal high-fat diets have been linked to increased risk of disease in offspring; however, this is the first report of such diets conferring deficits

to mental health. Interestingly, offspring of HF-fed dams exhibited significant reductions in cortical oxytocin receptor levels and colonization of these animals with *L. reuteri* was sufficient to correct this deficit.

61. Smith CJ, Emge JR, Berzins K, Lung L, Khamishon R, Shah P, Rodrigues DM, Sousa AJ, Reardon C, Sherman PM *et al.*: **Probiotics normalize the gut–brain–microbiota axis in immunodeficient mice.** *Am J Physiol: Gastrointest Liver Physiol* 2014, **307**:G793-G802.
62. Emge JR, Huynh K, Miller EN, Kaur M, Reardon C, Barrett KE, Gareau MG: **Modulation of the microbiota–gut–brain axis by probiotics in a murine model of inflammatory bowel disease.** *Am J Physiol: Gastrointest Liver Physiol* 2016, **310**:G989-G998.
63. Huang R, Wang K, Hu J: **Effect of probiotics on depression: a systematic review and meta-analysis of randomized controlled trials.** *Nutrients* 2016, **8**:1-12.
64. Steenbergen L, Sellaro R, van Hemert S, Bosch JA, Colzato LS: **A randomized controlled trial to test the effect of multispecies probiotics on cognitive reactivity to sad mood.** *Brain Behav Immun* 2015, **48**:258-264.