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## Title

Intraspecific trait variation across scales: implications for understanding global change responses

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18

20 Abstract

21 Recognition of the importance of intraspecific variation in ecological processes 22 has been growing, but empirical studies and models of global change have only 23 begun to address this issue in detail. This review discusses sources and patterns 24 of intraspecific trait variation and their consequences for understanding how 25 ecological processes and patterns will respond to global change. We examine 26 how current ecological models and theories incorporate intraspecific variation, 27 review existing data sources that could help parameterize models that account 28 for intraspecific variation in global change predictions, and discuss new data that 29 may be needed. We provide guidelines on when it is most important to consider 30 intraspecific variation, such as when trait variation is heritable or when non-31 linear relationships are involved. We also highlight benefits and limitations of 32 different model types, and argue that many common modeling approaches such 33 as matrix population models or global dynamic vegetation models would allow a 34 stronger consideration of intraspecific trait variation if the necessary data were 35 available. We recommend that existing data needs to be made more accessible, 36 and in some case new experiments are needed to disentangle causes of variation. 37

# 39 Introduction

40	Recently, many authors have stressed the importance of moving towards
41	more mechanistic models to be able to describe the response of species and
42	ecosystems to global change (Kearney & Porter, 2009; Chevin et al., 2010;
43	Pereira et al., 2010; Schurr et al., 2012). Current mechanistic and process-based
44	models are typically based on species-level traits and properties or even on
45	functional types, but some models are beginning to account for the fact that
46	species' traits are neither static nor homogenous in space or time (Bolnick et al.,
47	2011; Violle et al., 2014). For instance, many species exhibit trait differences
48	between populations, frequently taking the form of geographic clines that
49	correspond to environmental gradients (Aitken & Whitlock, 2013), and
50	important phenotypic differences also exist between individuals within
51	populations, and even within sex, age, or size classes (Bolnick et al., 2011;
52	Richardson et al., 2014). Such intraspecific variation can strongly influence
53	ecological processes and the conclusions drawn from models thereof (Boyce,
54	1977; Kendall & Fox, 2002; Grear & Elderd, 2008; Kearney <i>et al.</i> , 2009; Kramer <i>et</i>
55	al., 2010; Schindler et al., 2010; Oney et al., 2013). Understanding and
56	incorporating variation in traits is therefore important for basic science, for
57	making predictions about global change impacts, and for managing species
58	affected by global change.
59	In this review, we address the following questions:
60	1. What are the sources and patterns of intraspecific trait variation?
61	2. When will intraspecific variation be crucial for ecological responses to
62	environmental change?

63	3. How do existing models attempt to address trait variation, and have their
64	capabilities to do so been fully utilized?
65	4. Are there useful data sources that have been overlooked, or new data that
66	must be collected?
67	
68	We will define "traits" broadly, as including phenotypic characters (e.g.
69	coat color), behavioral or growth characteristics (e.g. fast vs. slow growth rate),
70	or individual-level demographic rates (e.g. expected number of offspring).
71	
72	Sources and patterns of trait variation
73	<u>Sources</u>
74	Intraspecific trait variation may be due to heritable differences
75	between individuals, or it may be due to phenotypic plasticity in trait values
76	across varying environmental conditions. Understanding which of these
	courses is recoonsible for trait variability is crucial for predicting global change
//	sources is responsible for trait variability is crucial for predicting global change
77	responses. If differences in trait values within a population are due to genetic
77 78 79	responses. If differences in trait values within a population are due to genetic differences they will be heritable, and therefore population-level means will be
77 78 79 80	responses. If differences in trait values within a population are due to genetic differences they will be heritable, and therefore population-level means will be subject to change through natural selection over generational timescales.
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<ol> <li>77</li> <li>78</li> <li>79</li> <li>80</li> <li>81</li> <li>82</li> <li>83</li> <li>84</li> <li>85</li> <li>86</li> </ol>	sources is responsible for trait variability is crucial for predicting global change responses. If differences in trait values within a population are due to genetic differences they will be heritable, and therefore population-level means will be subject to change through natural selection over generational timescales. Plasticity, on the other hand, enables immediate adjustment of phenotypic traits, which can buffer a population against rapid environmental change. Plasticity tends to slow adaptive evolution in the short term by weakening selective pressures, but by preserving population size and genetic diversity may provide potential for evolutionary responses in the long run (Crispo, 2008). The degree of plasticity exhibited in a trait is typically subject to genetic control and can

if it is costly (Crispo, 2008). See also Gomez-Mestre & Jovani (2013), who
developed a simple heuristic model to illustrate the potential interactions
between plasticity and adaptation.

91 **Epigenetic differences**, which affect gene expression but not the 92 underlying genetic code, can contribute to both categories. They contribute to 93 plastic responses, but some can also be heritable over intermediate timespans 94  $(\sim 1-5 \text{ generations})$ . For instance, differential DNA methylation in plants can 95 affect traits such as flowering-time or drought tolerance, and their heritability 96 (Fieldes & Amyot, 1999; Zhang et al., 2013). Such heritable epigenetic variation 97 may aid population adaptation to global change, but not all epigenetic effects are 98 adaptive: Rats exposed to pesticides show negative fitness effects over at least 3 99 generations, likely due to methylation changes (Manikkam et al., 2012). Broadly 100 speaking, any phenotypic effect not due to genetics might be said to be 101 epigenetic. However, as discussed above, "plasticity" refers to non-heritable 102 changes in phenotype due to environmental variation (Scheiner & Goodnight, 103 1984), while the epigenetic effects that have received the most research 104 attention are heritable (Richards, 2011). As the molecular basis of phenotypic 105 plasticity becomes better understood, we may need to develop new terms 106 distinguishing epigenetic mechanism with short-term vs. long-term effects. 107 Disentangling these different sources of trait variation is often challenging 108 (Gienapp et al., 2008). Plasticity can lead to trait variation between populations 109 even when they are genetically highly similar (Crispo, 2008). Common garden 110 experiments or pedigree studies are needed to distinguish plastic and heritable 111 variation (Wilson et al., 2010; Aitken & Whitlock, 2013; Blanquart et al., 2013),

though distinguishing heritable genetic and epigenetic effects presents furtherchallenges (Richards, 2011).

Moreover, which sources of variation are most important in real ecological systems is still poorly understood. In recent years, more researchers have begun to consider the consequences of genetic variation and evolutionary adaptation in studies of global change responses (Moran & Alexander, 2014), but epigenetic mechanisms and maternal effects are still rarely addressed (Bossdorf *et al.*, 2008). For most species, the cause of observed trait variation remains unknown.

121 When genetic and plastic effects on trait values are disentangled for 122 current global change responses, the contribution of plasticity has been larger in 123 many systems (Hoffmann & Sgro, 2011), but this likely reflects the relatively 124 short timespans involved. Heritability for traits strongly related to fitness (eg. 125 fecundity) is generally lower than for behavioral or morphological traits, but 126 significant heritability (and therefore potential for responses to selection) often 127 exists even in these traits (Mousseau & Roff, 1987). Where heritable changes 128 have been observed in response to global change, it is generally in species with 129 short generation times, high fecundity, and/or large population sizes – the types 130 of species one would expect to exhibit more rapid evolutionary change (Moran & 131 Alexander, 2014).

132

133 <u>Patterns</u>

Within a population or geographic area, trait variation can be structured
(varying in a consistent or predictable way) or unstructured (random) with
respect to space, time, or kinship. For instance, variation due to sex differences

137 in mammals and birds would usually be considered unstructured, because 138 stochastic processes typically determine sex, as would variation due to annual 139 fluctuations in an environmental variable around a stable mean. Persistent 140 differences between individuals in space or time (in behavior, growth rate etc.) 141 are structured variation. Such differences may either be due to heritable 142 differences, or to persistent environmental differences that cause plastic 143 responses. Natural history data can be useful in determining whether variation 144 is likely to be structured (Kendall & Fox, 2002). This can be important - for 145 example, misattributing structured variation to demographic stochasticity can lead to overestimation of extinction probabilities (Fox & Kendall, 2002). 146 147 Structured variation in traits across populations or large geographic areas can often be related to environmental gradients (Figure 1) or to geographic 148 149 structure. If the variation is heritable and dispersal is very low, lack of genetic 150 variation may inhibit adaptation to changing conditions; if dispersal is extremely 151 high, the movement of alleles between heterogeneous environments may 152 counteract local selective pressures (Aitken & Whitlock, 2013). If differences are 153 due adaptive plasticity (trait changes that increase fitness in the new 154 environment), it will facilitate gene flow by reducing the costs of dispersal, 155 whereas maladaptive plasticity (eg. smaller size or lower fecundity when 156 stressed) will do the opposite (Crispo, 2008). 157 158 Influences of intraspecific variation on ecological processes

159 One way that ecological processes can be affected by trait variation is due 160 to non-linearity in the relationship between the variable trait and the response of 161 interest, which is expressed by a mathematical relationship called **Jensen's** 

162 inequality (Ruel & Ayres, 1999; Bolnick *et al.*, 2011). This states that when the 163 relationship between a random variable x and a response variable y is non-164 linear, the expectation for *y* differs depending on whether we use the mean of x, 165 or the whole distribution. Specifically, if the relationship y = f(x) is concave-up, 166 then E[v] > f(E[x]) and if it is concave-down (convex), then E[v] < f(E[x]). To give 167 an example: seeds produced per unit pollen declines with increasing pollen 168 deposition; when pollen deposition varies, the average seeds per flower is lower 169 than predicted based on the average pollen deposited per flower (Richards *et al.*, 170 2009). Similarly, genetic variation in host insects that leads to variation in 171 number of eggs per seed can stabilize host-parasitoid population dynamics via 172 Jensen's inequality (Imura et al., 2003).

173 Biology is rife with non-linear relationships (Benton *et al.*, 2006), and 174 therefore there are many situations where using the trait mean in a process does 175 not lead to the same result as using the whole trait distribution. This can be 176 particularly important in predicting population growth and viability. For 177 instance, unstructured variation in survival probabilities or offspring number 178 between years typically reduces the long-term average growth rate in population 179 models (Boyce, 1977), which increases the extinction probability in small 180 populations (Kendall & Fox, 2002). The form of the non-linear relationship determines how variation affects the responses. If a matrix model is constructed 181 182 with even age/size bins, survival for the individuals within the bin will be 183 estimated well for a linear type II survival curve, but for a type III curve with 184 high juvenile mortality the survival of the younger classes will be 185 underestimated, and for a type I curve with late mortality the survival of older

classes will be overestimated. This, in turn, leads to under- or over-estimation of
population growth, respectively (Grear & Elderd, 2008).

188 The type of variation most frequently included in population models is 189 **demographic stochasticity**, which refers to variation in population growth 190 rates resulting from random variation in survival or reproduction (Bolnick *et al.*, 191 2011). This process is most important in small populations, because as 192 population size increases the mean survival or reproduction in a given year will 193 approach the true population mean. Genetic drift resulting from demographic 194 stochasticity can lead to random evolutionary changes in traits. 195 Intraspecific variation can also have positive effects on population growth 196 or stability. The **portfolio effect**, for example, arises when intraspecific or 197 interspecific variation reduce the variability of population growth or an 198 ecosystem service in a temporally variable environment (Bolnick et al., 2011). 199 For instance, Schindler et al. (2010) estimated that asynchrony between local 200 populations and variation within cohorts of Bristol Bay salmon led to 2.2-fold 201 lower variability in annual returns than if the population were homogeneous. 202 Intraspecific variation (in the form of trait diversity) can also positively affect 203 average productivity. This can be due to **sampling effects**, whereby the larger 204 the number of species or genotypes included the more likely it is that one with 205 high productivity will be present (similar to the portfolio effect, but in a spatial 206 context), or to **complementarity effects**, whereby species or genotypes use 207 resources differently and thus reduce competition. For example, ecological 208 experiments (Crutsinger et al., 2006) and restoration studies (Bischoff et al., 209 2010) suggest that intraspecific genetic variation (more maternal families or 210 clones) in plants will often increase primary productivity (eg. 50% increase in

NPP in 12-genotype vs. 1-genotype plantings of *Solidago altissima*) and reduce
variation in establishment success. Genetic differences in environmental
responses or resource use, cohort or microsite effects, and sub-population effects
are all examples of structured variation, though unstructured variation could
potentially contribute to these stabilizing or productivity-enhancing effects.

216

217 Besides having implications for population- or species-level responses to 218 global change, intraspecific variation may also affect species interactions and 219 community and ecosystem responses. There has been a renewed effort in recent 220 years to link species traits to environmental responses and ecosystem processes 221 (Lavorel & Garnier, 2002), and the need to understand the role of species 222 interactions in global change responses has become increasingly well recognized 223 (Gilman et al., 2010). Variation between individuals in environmental responses, 224 diet, etc. broadens the species niche for the species as a whole (though 225 population- and individual-level tolerances may be narrow), and broadens the 226 range of effects a species may have on its environment, including on other 227 species. Through heritable or plastic effects, global change drivers have the 228 potential to alter species trait means distributions in ways that affect species 229 interactions.

The amount of heritable variation in traits may be particularly important for species interactions. Overall, intraspecific genetic diversity in parameters affecting species interactions (eg. prey choice) tends to increase the number of connections and decrease interaction strength, which in turn tends to increase the stability of the network by dampening population oscillations and reducing the probability of cascading extinctions when a member of the community is lost

236 (Fridley & Grime, 2010; Bolnick et al., 2011; Wolf & Weissing, 2012). Similarly, 237 mathematical models have shown that heritable variation in prey preference 238 within a predator population can alleviate apparent competition between prey 239 species and affect the dynamics of predator and prey populations (Schreiber et 240 *al.*, 2011) and higher levels of heritable variation in both predator and prev can 241 lead to more stable dynamics (Saloniemi, 1993). The emerging sub-field of 242 community genetics has demonstrated that genetic variation in a "foundation 243 species" (e.g. in phytochemistry of a common plant) can have cascading effects 244 on the abundance and community composition of soil and leaf arthropods. 245 nutrient cycling, and even higher trophic levels (Crutsinger et al., 2006; Whitham 246 et al., 2008). For instance, Populus genotypes in a common garden experiment 247 explained more than 70% of microbial community composition and 78% of the 248 variation in microbial biomass (Schweitzer et al., 2008). The amount of variation 249 in plastic responses to global change factors, including shifts in timing and body 250 size, could also affect species interactions. For instance, if a plant and a 251 pollinator shift their phenology different amounts in response to climate change, 252 this could result in a mismatch in timing that may negatively affect one or both 253 populations (Gilman *et al.*, 2010) – but if either population exhibits variation in 254 the extent of the shift, this mismatch would be alleviated.

Evolutionary processes not only can alter trait means and variability (Figure 2D), but in some cases can affect population dynamics directly. In **'hard selection'**, there is a threshold that individuals must pass in order to survive or reproduce (e.g., surviving a minimum temperature) independent of population size. The removal of individuals that do not pass the threshold can have a strong impact on population size and persistence (Saccheri & Hanski, 2006). If a

261 population is exposed to altered environmental conditions, hard selection can 262 reduce its maximum population growth rate below replacement levels. If the 263 population evolves a higher intrinsic rate of population increase  $(r_{max})$  in the new 264 environment before it goes extinct, then it can increase again; this is called 265 evolutionary rescue (Kinnison & Hairston Jr., 2007; Aitken & Whitlock, 2013). 266 In **'soft selection'**, the strength of selection is frequency and/or density 267 dependent (e.g. the largest 20% of individuals survive), and thus selection may 268 not directly affect population size – though it can affect population cycles if, for 269 example, the competing genotypes have reproductive rates that respond 270 differently to their own density (Saccheri & Hanski, 2006). For colonizing 271 populations, small evolutionary increases in r<sub>max</sub> in the first few generations can 272 increase the probability of establishment by enabling super-exponential 273 population growth (Kinnison & Hairston Jr., 2007). This seems to have occurred 274 in some invasive species. For instance, invasive populations of salmon in New 275 Zealand accumulated changes in a variety of traits over 26 generations that more 276 than doubled survival and fecundity relative to non-local genotypes (Kinnison & 277 Hairston Jr., 2007).

278 **Shifts in the mean trait value of a population** may change the selective 279 environment experienced by co-occurring species, resulting in **eco-evolutionary** 280 interactions (Post & Palkovacs, 2009). Such eco-evolutionary interactions can 281 alter competitive and trophic dynamics. Several notable examples come from 282 cases of introduced species or biological control. Lankau and Strauss (2007) 283 found that genetic variation in *Brassica nigra's* production of sinigrin, an 284 allelopathic chemical, promoted community diversity, and vice versa. High-285 sinigrin plants did well in heterospecific (newly-invaded) but not conspecific

neighborhoods, causing low-sinigrin plants to tend to increase in areas
dominated by *B. nigra*, finally enabling other species to re-invade low-sinigrin
patches of *B. nigra* where there was a 1.5 to 8-fold increase in seedling density
compared to high-sinigrin patches. In communities invaded by the related *Allaria petiolata*, some resident species also exhibit evolution of increased
tolerance to sinigrin, exhibiting up to a 2.8-fold difference in response to
competition (Lankau, 2012).

293 Starting in the 1960's, many models have examined the effect of evolution 294 on the dynamics of two-species systems and have found that evolution can either 295 stabilize or destabilize interactions (Fussman et al., 2007; Hartig et al., 2014). 296 However, the ecological assumptions in these models tend to be relatively simple 297 and there has been limited work on multi-species systems (Fussman *et al.*, 298 2007). One of the best documented empirical examples is that of myxoma virus 299 and rabbits in Australia: the virus was introduced as a biological control but, as 300 has been observed in other host-parasite systems, evolution led to both reduced 301 virulence of the virus and reduced susceptibility of the host (Fussman et al., 302 2007), dampening oscillations and making population crashes less likely. 303 304 **Dispersal** is a further crucial component for understanding the effects of

304 Dispersal is a further crucial component for understanding the effects of
 305 trait variability on global change responses within species and communities, as it
 306 affects both the probability of range shifts and population expansion as

307 well as the potential for evolution (Aitken *et al.*, 2008; Kubisch *et al.*, 2014).

308 The interaction between dispersal and evolution, coupled with interspecific

309 interactions, will likely determine the extent to which species respond to climate

change via either range shifts or local adaptation (Brooker *et al.*, 2007; Aitken *et al.*, 2008; Atkins & Travis, 2010; Bocedi *et al.*, 2013).

312 Dispersal ability is a trait that can evolve and exhibit plasticity itself 313 (Ronce, 2007). Variation in dispersal affects population spread rates. Dispersers 314 are likely to be a non-random subset of the population in terms of their behavior 315 (Wolf & Weissing, 2012), size, or other traits (Nathan *et al.*, 2011). This may 316 result in important structured variation between new and old populations, and 317 can result in selection on dispersal traits (Shine *et al.*, 2011). Such increases in 318 dispersal ability have been documented in the invasive cane toad and invasive 319 plants, as well as in insects undergoing climate-linked range expansions, and can 320 accelerate range shifts (Shine *et al.*, 2011). Conversely, if most dispersers have 321 low survival due to unsuitability of the matrix between habitat patches, or if poor 322 dispersers are left behind, selection or trait sorting can favor reduced dispersal 323 ability (Shine et al., 2011; Travis et al., 2012; Moran & Alexander, 2014). 324 However, these changes in dispersal traits may be transitory on a population 325 scale, as dispersal ability can trade off with other traits (Travis *et al.*, 2012), such 326 as competitive ability, that are more important in high-density populations. 327 Besides evolving itself, dispersal ability strongly affects the amount of 328 local genetic variation in populations. This local variation tends to increase the 329 speed of evolutionary responses because alleles advantageous in new 330 environment may already be present at moderate frequencies (Barrett & 331 Schluter, 2007; Wolf & Weissing, 2012). Because species that have small or 332 highly fragmented populations and low genetic diversity may have limited 333 potential for adaptation to changing future conditions, managers are increasingly 334 considering "genetic restoration" or assisted movement to boost local genetic

335	diversity (Hoffmann & Sgro, 2011; Aitken & Whitlock, 2013). The direction of
336	gene flow also matters: gene flow from historically warmer areas and between
337	areas of similar historical climate may increase the probability that alleles
338	advantageous under climate change will be present; gene flow from the range
339	center toward the trailing edge, however, is likely to be unfavorable (Moran &
340	Alexander, 2014).
341	
342	Modeling approaches and current applications incorporating intraspecific
343	variation
344	Population models
345	Three common approaches for modeling trait variability in local
346	populations are matrix models (MM), integral projection models (IPMs), and
347	individual-based models (IBMs). All three can incorporate unstructured
348	variation by adding stochastic effects to account for "random" environmental
349	variation or demographic stochasticity. All can also include population-
350	ecological relationships such as density dependence. They differ more strongly
351	in their ability to include structured variation. All approaches can be
352	incorporated into landscape- or range-wide models, as discussed in the next
353	section.
354	Matrix models are an extension of classical population models – instead
355	of modeling one population with uniform trait values they divide a population
356	into several age or size classes, each of which is uniform in its trait values. A
357	transition matrix represents fecundity, mortality and the transition between
358	classes. The obvious limitation of these models is that they neglect variation in
359	demographic rates within classes (Boyce, 1977). Particularly if there is size-

dependent growth or growth correlations within classes (2B), this can lead to
errors in prediction (Pfister & Stevens, 2003). Careful use of natural history to
define age or stage divisions (Figure 2A) will reduce, but may not eliminate, this
problem (Grear & Elderd, 2008).

Matrix models can include structured variation in phenotypic traits, even heritable variation, but only if the traits are discrete, and the mode of inheritance simple. For instance, Kruger and Lindstrom (2001) divided buzzards into three color morphs which differed in reproductive success. In stage-based models, the distribution of ages within a stage or variation in the length of time an individual occupies a stage can affect dynamics, especially transient dynamics (de Valpine *et al.*, 2014).

Integral projection models (IPMs) can be viewed as an extension of
matrix models, where differences within a population are modeled by
continuous distributions instead of discrete classes (Ellner & Rees, 2007).
Compared to matrix models, this makes it easier to incorporate multiple
differences between individuals, such as differences within a size or age class
(Zuidema *et al.*, 2010; de Valpine *et al.*, 2014).

377 Heritability can be included in more complex IPMs by introducing a 378 distribution of trait values within an age class, and making this distribution in 379 offspring traits dependent on parental traits (Coulson *et al.*, 2010). Changes in 380 the environment (Hoffmann & Merila, 1999) and in the amount of genetic 381 variation in a population due to immigration, selection, or bottlenecks (Bryant et 382 al., 1986; van Straalen & Timmermans, 2002; Lavergne & Molofsky, 2007) can 383 affect heritability. Model projections based on a fixed heritability estimate 384 should therefore only be applied over short time periods. While cumulative

385 effects (such as survival effects of past reproduction) are typically not included 386 in an IPM, age-class models can identify differences in individual growth rates 387 because fast growing individuals will be unusually large for their age. For 388 instance, Jansen et al. (2012) found that in a tropical understory palm growth 389 differences between individuals persisted over a lifetime and that fast growers 390 had a 1.8-fold greater contribution to population growth. IPMs can also be used 391 to explore trait evolution using an evolutionarily stable strategy (ESS) approach. 392 though this does not account for transitory dynamics (Ellner & Rees, 2007).

393 Individual-based models (IBMs), as the name suggests, model a 394 population by describing each individual separately, using state variables such as 395 age, size, location, genotype, and fecundity (Grimm & Railsback, 2005). This 396 allows tracking cumulative changes over the lifecycle of each individual (e.g. 397 increasing size or reproductive history), as well as any conceivable variation 398 between individuals, including genetic variation in environmental responses. 399 Besides evolution, they can also incorporate spatial heterogeneity and individual 400 movement (DeAngelis & Mooij, 2005; Dytham et al., 2014).

401 There are a number of additional advantages associated with an 402 individual-based approach. Like real populations, IBMs have a discrete number 403 of individuals rather than a continuous value, making them naturally suitable for 404 capturing effects of demographic stochasticity in small populations, as well as for 405 simulating spatial dynamics within a population. Also, they allow one to 406 explicitly model allelic frequency and diversity, avoiding the problem of 407 assuming constant heritability. For instance, Kuparinen et al. (2010) simulated 408 the evolution of growing-season timing in *Pinus* and *Betula*, using a simple multi-

409 locus model of bud-burst and bud-set date, over 100 years of climate change, and410 found that a higher adult mortality rate reduced evolutionary lags.

411 Due to these advantages, and supported by growing computing power 412 which makes it possible to simulate large numbers of individuals, IBMs have 413 become increasingly popular. A common application is individual-based forest 414 gap models that project the influence of environmental factors on productivity 415 and species composition (Smith *et al.*, 2001). Individual-based models are also 416 used for fisheries models and for population viability analysis (DeAngelis & 417 Mooij, 2005; Frank et al., 2011). Comparing IBMs with different levels of detail 418 to data from real systems can help to identify which types of variation are most 419 important in these systems (DeAngelis & Mooij, 2005; Benton, 2012). IBM 420 simulations can also be used to test when the simplifying assumptions of other 421 model classes are likely to lead to problems (Pfister & Stevens, 2003). 422

#### 423 Modelling landscape- or range-scale dynamics

424 In many global change applications, the focus is not so much on 425 describing the change in local populations, but rather on capturing spatial 426 structure and large-scale dynamics. For this purpose, two approaches exist that 427 allow including trait variability. The first consists of using known spatial 428 extensions of the above-described process-based population models. The second 429 is to extend widely used statistical approaches such as correlative species-430 distribution models to include trait variability. We cover both options in this 431 section, as well as the application of these concepts in dynamic vegetation models. 432

433

434 Metapopulation or metacommunity models are extensions of non-435 spatial population models that describe spatial structure through assuming a 436 number of connected local populations (Ovaskainen & Hanski, 2004). Within 437 each local population (patch), population dynamics may be modeled using any of 438 the approaches previously described. For example, the CD FISH model, a 439 simulator incorporating population genetics, demography, and stream 440 connectivity, has shown how landscape-level connectivity in stream systems can 441 constrain population vulnerability to environmental change in salmonid species 442 (Landguth et al., 2014). Spatial Integral Projection Models (SIPMs) combine 443 IPMs with models of dispersal to predict spread across a landscape (Jongejans et 444 al., 2011), which can be useful in understanding the spread of invasive species or 445 native species under climate change. In principle, this approach allows one to 446 naturally scale-up the effects of intra-specific variation within and between 447 populations to examine its impact on larger scales. In practice, the 448 computational demands of this approach often limit the size of the landscape 449 that can be simulated.

450

451 Species distribution models (SDMs) aim to predict the occurrence 452 probability of a species given environmental variables and therefore the limits 453 and size of its geographic range. The simplest and still most common approach 454 is to correlate presences (and absences if available) with environmental 455 variables. Various criticisms have been against **correlative SDMs**, mostly 456 relating to the fact that the realized niche (where the species does occur) can 457 differ from the fundamental niche (where it could occur) for a number of 458 reasons, including disequilibrium with the environment, biotic interactions, and

459 dispersal and source-sink dynamics (Guisan & Thuiller, 2005). Still, correlative 460 SDMs are widely used because they make use of the museum or herbarium 461 records and climate data that are easily available (Moran & Alexander, 2014). 462 Due to the challenges of using correlative SDMs to predict range 463 dynamics, particularly in the presence of biotic interactions, intraspecific 464 variability and evolutionary potential, there has been a steady movement 465 towards including more processes in these models (Dormann et al., 2012). For 466 example, physiological SDMs make use of knowledge about physiological limits 467 (Porter et al., 2002; Buckley, 2008; Rödder et al., 2009; Buckley et al., 2011) and behavior (Kearney et al., 2009). While most physiological SDMs to date have 468 469 been applied to ectothermic organisms or plants, known relationships of body 470 size to metabolic rate and/or heat exchange (Yarbrough, 1971)could be more 471 widely applied to birds or mammals as well. Hybrid SDMs add a dispersal 472 model on top of a classical SDM (De Cáceres & Brotons, 2012)(Marion et al., 473 2012), while **Dynamic Range Models** include both explicit population dynamics 474 and dispersal (Schurr *et al.*, 2012). SDMs can also be "stacked" to examine effects 475 of environmental changes on biodiversity, either assuming that species 476 distributions are independent (Guisan & Rahbeck 2010) or interacting (Clark et 477 al., 2013). 478 In all such extensions of classical SDMs, intraspecific differences can be 479 included in principle, but such applications are still rare. When intraspecific

480 differences are included, the most common approach is to describe **differences** 

481 **in the environmental response of subpopulations** by fitting separate SDMs to

482 each subpopulation across a species' range. This has mainly been done in trees,

483 for which good provenance (common garden) study data are often available

484 (Savolainen et al., 2007). An example is the Phenofit model, which predicts tree 485 presence based on the link between phenology and reproductive success, can 486 allow different populations different responses (Chuine & Beaubien, 2001). 487 Incorporating intraspecific population variation in that way can alter predictions 488 of global change responses, making them either more (Valladares *et al.*, 2014) or 489 less (Garzon *et al.*, 2011) severe. For instance, incorporating population-level 490 differences in *Pinus contorta* vielded similar results to a species-level model for 491 current distribution, but the models incorporating genetic diversity projected 492 less than half as much range loss even without migration (Oney et al., 2013). 493 Ideally, such studies will be based on or complemented by data confirming 494 genetic differences in environmental responses, as apparent differences between 495 subpopulations could also be caused by biotic interactions or other non-genetic 496 factors.

497

498 There are multiple examples in the recent literature of metapopulation or 499 SDM models being used to investigate the potential impact of evolution on global 500 change responses. For instance, using sequentially updated SDMs, Urban et al. 501 (2007) showed that the maximum annual temperature of areas occupied by the 502 invasive cane toad (Chaunus [Bufo] marinus) had increased from 1975 to 2005, 503 interpreted by the authors as niche expansion into increasingly extreme habitats. 504 Models can also identify when such adaptation could alter dynamics. Kearney et 505 al. (2009) developed a mechanistic niche model incorporating a quantitative 506 genetic model to investigate whether evolution of egg desiccation resistance in 507 the dengue mosquito Aedes aegypti might enable it to expand its range, and 508 found that it would take 17.4% less time to reach Darwin under climate change

with a heritability 0.5 vs. 0.15. Similarly, a dynamic vegetation model developed
by Kramer *et al.* (2010) to explore climate change responses in European beech
incorporated a quantitative genetic submodel for budburst and stomatal
conductance, and found that adaptive changes could occur at the leading and
trailing edge over just 2-3 generations. Interactions between landscape features,
the evolution of dispersal traits, and spread rates can also be investigated with
IBMs (Travis *et al.*, 2012).

516 An important caveat is that most models that aim to project the effects of 517 evolutions, with a few exceptions (Kramer et al., 2010), have not been 518 systematically validated, for example by reproducing the current pattern of local 519 adaptation. We suggest that results should therefore not be interpreted as 520 predictions, but rather as indicators that suggest a potential impact of evolution 521 on environmental responses. It is also unclear how well the true genetic 522 structure of ecologically important traits are approximated by the assumptions 523 made in particular models (e.g. ten two-allele loci), and it is increasingly 524 recognized that models of species presence or persistence need to take into 525 account phenotypic plasticity (Chevin *et al.*, 2010; Valladares *et al.*, 2014). 526

527 Dynamic global vegetation models (DGVMs) are individual- or
528 population-based population models that have a focus on predicting the
529 composition and dynamics of the vegetation by describing physiological
530 processes such as photosynthesis and water uptake, biotic interactions, and
531 disturbances. In principle, the structure of these models is well suited to assess
532 the effects of intraspecific variability in those processes. However, due to data
533 limitations and computational constraints, most DGVMs currently still describe

vegetation dynamics and community interactions in terms of broad functional
types that summarize a potentially large group of similar species. Hence,
potential for improvement exists regarding the representation of interspecific as
well as intraspecific trait differences(Hartig *et al.*, 2012).

538 In recent years, various studies have considered options to include trait 539 variability in DGVMs. One approach is motivated by the observation that 540 environmental drivers explain 52% of global variation in traits (van Bodegom et 541 *al.*, 2014). Allowing traits to vary within PFTs following observed trait-climate 542 relationships can therefore improve the match of predicted to observed 543 vegetation types (Verheijen et al., 2013). Similarly, Stahl (2014) demonstrated 544 that a few easily-measured species-level traits could help predict environmental 545 range limits in trees, which could impose useful constraints on a DGVM. Rödder 546 et al. (2009) found that including physiological limits in an SDM for slider turtles 547 enabled better prediction of their invasive range; including similar limits on 548 vegetation types could provide similar benefits in DGVMs. Trait variation could 549 also be assumed to arise from a large potential for plasticity (Nicotra *et al.*, 2010) 550 or genetic variation (Kramer et al., 2010; Valladares et al., 2014). Adaptive 551 global dynamic vegetation models construct communities of trait combinations 552 by filtering those traits that are most competitive and able to coexist in a given 553 environment (Scheiter et al., 2013).

554

555 To conclude the review of the state-of-the-art in the field of modelling 556 trait variability: all commonly used population model types could accommodate 557 trait variability to a much greater degree than is the current practice. When trait 558 variability is included, conclusions can be substantially altered. The inclusion of

trait variability has been limited by computational and data constraints. The
former constraint is easing as computers become more powerful, the latter will
be discussed in the next section. Finally, we emphasize that no one modeling
approach is ideal for all situations, but that model choice rather depends on how
the model will be used and whether there is sufficient data and/or
understanding of process to parameterize and justify a more complex model.

565

### 566 Data and methods for parameterizing models that include trait variation

## 567 and trait evolution

Recent advances in modeling techniques (see above) provide increasing flexibility to ecologists and evolutionary biologists to model and understand the consequences of trait variation. However, they also highlight the need for gathering new data that allows more direct estimates of structured and unstructured trait variation, and the need of making existing data better available to researchers.

574 Data sources

575 Especially at the local scale, useful data for estimating variation in 576 important traits or demographic rates often already exist. For example, data on 577 phenotype, growth, fecundity, and survival are typically collected at the 578 individual level (Clark et al., 2011), enabling one to construct trait distributions. 579 If multiple measurements are available for each individual, structured variation 580 (e.g. individuals that consistently grow faster or slower) can be distinguished from unstructured variation. Such longitudinal data can also be used to 581 582 disentangle plastic versus evolutionary responses to global change, using 583 approaches such as the 'animal model' (Gienapp et al., 2008) – especially if family

relationships are also known. For instance, phenotypic plasticity in red squirrels
accounted for 62% of an 18-day advance in average birth date, while 13% was
attributable to evolutionary change (Berteaux *et al.*, 2004).

587 However, data are rarely synthesized in this manner, and instead are 588 usually aggregated for analysis – for instance, into species-level means and 589 variances (Clark *et al.*, 2011). Unless the original data are included as an 590 appendix, it is usually impossible to obtain the individual-level measurements 591 from the published literature, which greatly complicates meta-analysis and 592 means that information can easily be lost. Fortunately, journals and funding 593 agencies have been encouraging or, more recently, requiring researchers to 594 archive data in a more accessible format. Trait databases are currently being 595 constructed for many taxa. Plants seem to be better represented than animals 596 (but see the linked trait databases at http://scales.ckff.si/scaletool/). The TRY 597 database, for instance, contains 5.6 million trait entries for 100,000 plant species 598 around the world, and preliminary analyses showed that up to 40% of overall 599 variation in a trait can be intraspecific (Kattge *et al.*, 2011). Such data can be 600 used to better define plant functional types, and to give an idea of the range of 601 values a trait may take within a species, though for many species it is as yet too 602 patchy to, for example, compare trait distributions between multiple populations 603 of a species.

Large-scale observational networks can also provide useful data. Again,
this kind of data tends to be more abundant for plants than for animals. For
instance, Forest Inventory and Analysis (FIA) censuses have tracked tree sizes
and identities in plots across the United States for decades, and such data can be
used to examine variation in global change responses between life stages, species

609 types, or regions, or to parameterize individual-based forest models (Canham et 610 *al.*, 2006; Bell *et al.*, 2014). The new National Ecological Observatory Network 611 (NEON), now coming online, while not explicitly geared to investigate 612 intraspecific variation, will be collecting data such as size and reproductive 613 status for select species, phenology, soil microbial metagenomes, as well as 614 detailed measurements of the physical environment (Keller, 2010). Data on 615 plants' plastic phenological responses to climate can be obtained from both 616 observation networks (eg. <u>https://www.usanpn.org/</u>) and meta-databases. 617 However, to fully investigate the role of intraspecific variation in species' 618 environmental responses, new data specifically collected for this purpose are 619 needed. In addition to the need for accessible trait-variation data for a broader 620 range of species, some research questions, such as those relating to the role of 621 evolution in global change responses, require specialized experiments or 622 observations. For instance, while provenance study data can be useful for 623 understanding tree responses to climate, these studies rarely included very long-624 distance transfers, or populations from or test sites at the environmental limits 625 (Leites *et al.*, 2012). Another important consideration is that covariation in traits 626 can be important for population and eco-evolutionary dynamics, so sampling 627 schemes should be designed to make calculation of covariances possible - long-628 term demographic data are particularly valuable (Saether & Bakke, 2000). 629 Theoretical models can suggest where it is important to consider 630 variation, and therefore where more data is required. For instance, key factors 631 affecting the potential rate of evolutionary change, and therefore the probability 632 of extinction or persistence in a population facing environmental change, include 633 demographic properties of populations (e.g. generation time and maximum

634 population growth rate), the strength of selection, and the available genetic and 635 environmental variance in the key traits (Chevin et al., 2010). They can also 636 suggest when the costs of selection might lead to extinction (Burger & Lynch, 637 1995). Gienapp et al. (2013) provide a good example of applying such models to 638 real populations. Another key area in which more data is needed to understand 639 global change responses is range shifts, as it is unknown in most cases whether 640 species ranges are currently limited by abiotic factors, biotic interactions, 641 dispersal, or lack of genetic variation (Sexton et al., 2009).

642 Distinguishing genetic (or epigenetic) variation from plastic responses to 643 the environment tends to require either detailed, long-term observational data 644 that enables one to build a pedigree (Wilson et al., 2010) or common-garden 645 experiments in which different genotypes are reared in one or more common 646 environments (Blanquart et al., 2013). Unlike the incidental individual-level data 647 discussed above, which may be able to generally distinguish structured vs. 648 unstructured variation, these types of studies are usually specifically designed to 649 investigate the genetic components of intraspecific variation. They are thus 650 extremely valuable for understanding how and when to include genetic variation 651 in models of environmental responses. However, because of high cost in time 652 and money, these studies are rarer than short-term observational trait 653 measurements, and have similar issues of accessibility if the data have not been 654 archived.

If we could better identify the genetic basis of the trait(s) of interest,
extrapolating heritabilities outside of tested environments would become more
feasible. Genome-wide association studies (GWAS), usually making use of singlenucleotide polymorphisms (SNPs) are becoming more widely used as a means of

659 identifying candidate genes for particular traits of interest. SNP variation can be 660 correlated either with variation in phenotype (e.g. water use efficiency), or with 661 variation in environment (e.g. climatic water deficit) (Neale & Kremer, 2011). 662 Results are usually qualitatively consistent with other approaches. However, 663 the amount of variation explained is typically lower than the total heritability 664 because rare variants unique to local families are often missed, multiple 665 functional alleles may exist, and linkage disequilibrium and epistasis may 666 interfere with the analysis (Brachi et al., 2011). Some of the missing heritability 667 may also be attributable to epigenetic variation (Bossdorf et al., 2008). A rangewide sample will have low power to detect significant associations with 668 669 phenotype if adaptive alleles are locally common but globally rare, so sampling 670 must be structured to account for this (Brachi *et al.*, 2011). Methods for 671 incorporating realistic functional genetic submodels into models of species' 672 responses to global change are still in the early stages of development. 673 Neutral genetic variation (e.g. microsatellites), while not always closely 674 correlated with functional variation (Holderegger *et al.*, 2006), is useful for 675 estimating the degree of gene flow between populations, hybridization, or 676 dispersal ability (Aldrich et al., 2003; Ashley, 2010; Moran & Clark, 2012). 677 Dispersal ability and population connectivity can also be estimated based on 678 physical traits in systems for which there are good mechanistic models, such as 679 wind dispersal of seeds (Kuparinen, 2006), via inverse modeling of observations 680 of dispersed individuals (Jones & Muller-Landau, 2008), or through tracking of 681 tagged individuals. Regardless of the method, it is advisable to take into account 682 intraspecific variation – for instance, considering that adult trees may vary in 683 seed production.

685	A further challenge connected to data on trait variability is statistical
686	analysis and the connection of these data to models. Many of the data types we
687	have discussed above should probably be treated with hierarchical statistical
688	models that allow including underlying dependencies between traits as well as
689	spatial, taxonomic and phylogenetic structure in the data. Such multilevel
690	models describe how individual responses vary according to the context, help
691	avoid over-fitting because they don't use independent "individual" parameters,
692	and enable the proper propagation of uncertainty from parameters to
693	predictions (Clark, 2003). If vital rate calculations are based on capture-
694	recapture methods, it may also be important to account for size- or stage-effects
695	on capture or detection probabilities (Punt et al., 2006).
696	For process-based models such as IPMs, DRMs (Dynamic Range Models),
697	or DGVMs, data can be used in two ways: one can use statistical approaches to
698	infer individual parameters or subsets of the model parameters first, and then
699	use the process-based model to calculate the consequences of those parameter
700	estimates. An alternative is to infer model parameters inversely for the entire
701	model simultaneously, using the raw observations. For DRMs and SDMs, for
702	example, this is often still possible in a hierarchical Bayesian approach (Bykova
703	et al., 2012; Marion et al., 2012). For more complex models such as DGVMs, it
704	may become necessary to resort to simulation-based inference methods to
705	calibrate model parameters (Hartig et al., 2011). Bayesian approaches allow
706	mixing parameter estimates from independent analysis and from calibration,
707	which allows a flexible mix of both approaches (Hartig et al., 2012).

709 <u>Conclusions</u>

### 710 When variation matters

711 Generally, intraspecific variation is likely to be important for 712 understanding responses to global change a) for phenotypic traits that affect 713 demography, b) when the relationship of interest is a non-linear function of the 714 trait (Jensen's inequality), c) in small populations, where demographic 715 stochasticity can cause mean demographic rates or phenotypic traits to fluctuate. 716 d) when variation in the trait of interest is structured (because structured 717 variation cannot be accounted for merely by adding stochasticity to a model). 718 and e) if the timescale of interest spans multiple generations, and the trait is 719 likely to be under directional selection (e.g. drought tolerance in a warming 720 environment). 721 Mechanistic models can be helpful in identifying which traits or 722 demographic rates have the largest effect on a process of interest, and where 723 variation might be most important. For instance, Nathan et al. (2011) found that 724 for wind-dispersed trees, the plant traits most important for migration rate 725 tended to be 1) age at maturity, 2) post-dispersal survival, 3) seed terminal 726 velocity, 4) fecundity, 5) tree height, and 6) time between seed crops. 727 Conversely, natural history data can suggest what traits should be allowed to 728 vary in models: in *Pinus sylvestris*, for instance, there is little variation in seed 729 wing loading because seed mass and wing size are correlated, but isolated trees 730 have more seeds than those in dense populations (Debain *et al.*, 2003). 731 *Incorporating variation in global change models* 732 Although models differ in their ability to include trait variation, it seems 733 from our analysis that the opportunity for including and analyzing trait

variability hasn't been fully explored or exploited in any of the modeling

approaches we considered. The main reason seems the lack of data and

computational restrictions. As both limitations should ease in the coming years,

737 we hope that these capabilities will become more widely used.

738 Data needs for the future

739 Computational limits can be expected to improve without active effort of 740 the ecological community, but the limits on data will not. Existing data collected 741 at the individual level should be more frequently used to investigate intraspecific 742 variation. The growing availability of databases and data archives mean that 743 future meta-analyses and models will be able to rely less heavily on published 744 trait statistics and therefore will be able to address variation more accurately. 745 However, for distinguishing causes of structured variation that may have 746 different effects (e.g. genetic vs. environmental) more specialized experiments 747 are required. Data from these variation-focused studies also needs to be made 748 more widely available.

749 <u>Summary</u>

Intraspecific variation has been gaining attention in ecological theory. Empirical
studies quantify and classify variation in real populations, while developments in
modeling techniques enable the effects of variation to be assessed in more
sophisticated ways. Successfully integrating theoretical insights with these new
data and modeling techniques will be crucial for making robust predictions of
species responses to global change.

756

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761

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- 1116 validation of model output. *Journal of Ecology*, **98**, 345–355.
- 1117
- 1118 <u>Figure Captions</u>

**Figure 1:** At the landscape scale (top), trait variation is shaped by large-scale environmental gradients. Within local sites (bottom), phenotypic variation in each species is shaped by genetic variation and by plastic responses to the local biotic and abiotic environment. This variation affects species interactions with one another and with the environment, ultimately impacting individual fitness.

1124 Differences in fitness feed back to affect genetic variation. Local genetic 1125 variation is also affected by dispersal between populations (white arrows at top). 1126 Figure 2: A) Variation in a trait deriving from developmental processes (e.g. 1127 changes in offspring number with size or age) can be accounted for – for 1128 example, by defining age/size classes to minimize intra-class variation (dashed 1129 lines). However, variation in natural populations is not so simple. B) If growth 1130 rate and offspring number both depend on current size, then small differences in 1131 initial size can cause the relationship between age and fecundity to vary between 1132 individuals. In this example, larger individuals grow faster. C) Even without 1133 such growth correlations, individuals can differ in the relationship between the 1134 predictor and response variables, affecting the shape of the function. D) If such 1135 differences are heritable, then trait distributions and the population processes 1136 that depend on them may change over time.



1137 1138 **Figure 1** 

