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Authors
Pekmezci, M
Villanueva-Meyer, JE
Goode, B
et al

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The genetic landscape of ganglioglioma

Melike Pekmezci1, Javier E. Villanueva-Meyer2, Benjamin Goode1, Jessica Van Ziffle1,3, Courtney Onodera1,3, James P. Grenert1,3, Boris C. Bastian1,3, Gabriel Chamyan4, Ossama M. Maher5, Ziad Khattab5, Bette K. Kleinschmidt-DeMasters6, David Samuel7, Sabine Mueller8,9,10, Anuradha Banerjee8,9, Jennifer L. Clarke10,11, Andrew W. Bollen1, Arie Perry1,9, Tarik Tihan1 and David A. Solomon1,3*

Abstract
Ganglioglioma is the most common epilepsy-associated neoplasm that accounts for approximately 2% of all primary brain tumors. While a subset of gangliogliomas are known to harbor the activating p.V600E mutation in the BRAF oncogene, the genetic alterations responsible for the remainder are largely unknown, as is the spectrum of any additional cooperating gene mutations or copy number alterations. We performed targeted next-generation sequencing that provides comprehensive assessment of mutations, gene fusions, and copy number alterations on a cohort of 40 gangliogliomas. Thirty-six harbored mutations predicted to activate the MAP kinase signaling pathway, including 18 with BRAF p.V600E mutation, 5 with variant BRAF mutation (including 4 cases with novel in-frame insertions at p.R506 in the B3-cC loop of the kinase domain), 4 with BRAF fusion, 2 with KRAS mutation, 1 with RAF1 fusion, 1 with biallelic NF1 mutation, and 5 with FGFR1/2 alterations. Three gangliogliomas with BRAF p.V600E mutation had concurrent CDKN2A homozygous deletion and one additionally harbored a subclonal mutation in PTEN. Otherwise, no additional pathogenic mutations, fusions, amplifications, or deletions were identified in any of the other tumors. Amongst the 4 gangliogliomas without canonical MAP kinase pathway alterations identified, one epilepsy-associated tumor in the temporal lobe of a young child was found to harbor a novel ABL2-GAB2 gene fusion. The underlying genetic alterations did not show significant association with patient age or disease progression/recurrence in this cohort. Together, this study highlights that ganglioglioma is characterized by genetic alterations that activate the MAP kinase pathway, with only a small subset of cases that harbor additional pathogenic alterations such as CDKN2A deletion.

Keywords: Ganglioglioma, Epilepsy, Seizures, Glioneuronal tumor, Targeted next-generation sequencing, Ras-Raf-MEK-ERK, MAP kinase signaling pathway, BRAF, KRAS, RAF1, NF1, FGFR1, FGFR2, ABL2

Introduction
Ganglioglioma is a well-differentiated and typically slow-growing glioneuronal neoplasm composed of dysplastic ganglion cells in combination with neoplastic glial cells [2]. They often arise in the temporal lobe of children and young adults in association with seizures. However, they can occur at any age and throughout the neuraxis including the cerebellum, brainstem, and spinal cord. The neuroimaging appearance is variable, but they often display a mix of solid and cystic components. Most gangliogliomas correspond histologically to WHO grade I and do not recur after complete resection. However, gangliogliomas are both histologically and clinically variable, and tumor recurrence or anaplastic progression occurs in a subset of cases.

The activating p.V600E hotspot mutation in the BRAF oncogene has been identified in a subset of gangliogliomas, ranging from approximately 10–60% depending on the study and anatomic site, with highest frequencies reported in cortical tumors and lower frequency reported in spinal cord tumors [6, 7, 9, 11–13, 16, 21, 27, 30, 31, 36–38]. However, BRAF p.V600E mutation is not specific to ganglioglioma and has been described in a wide spectrum of neuroepithelial tumors including pilocytic astrocytoma, dysembryoplastic neuroepithelial tumor (DNET), pediatric...
IDH-wildtype diffuse astrocytoma, polymorphous low-grade neuroepithelial tumor of the young (PLNTY), pleomorphic xanthoastrocytoma, and epithelioid glioblastoma [6, 12, 17, 22, 30, 31, 36, 38]. Additionally, the genetic alterations responsible for BRAF p.V600 wildtype gangliogliomas are largely unknown, as is the spectrum of any additional cooperating gene mutations or copy number alterations. Herein, we performed comprehensive molecular profiling on a cohort of 40 pathologically-confirmed gangliogliomas in order to evaluate the genetic landscape of this tumor entity and identify any genetic alterations that may correlate with differences in clinical outcomes or imaging and histologic features.

**Methods**

**Patients and tumor tissue**

We searched our institutional pathology archives for cases with a diagnosis of ganglioglioma, spanning years 1990 to 2017. Cases with available diagnostic slides and tissue blocks containing sufficient tumor tissue for genetic analysis were included. All tumor specimens had been fixed in 10% neutral-buffered formalin and embedded in paraffin. Pathologic review of all tumor samples was performed to confirm the diagnosis by a group of five expert neuropathologists (M.P., A.W.B., A.P., T.T., and D.A.S.) with a unanimous consensus diagnosis established for all included cases. All tumors contained an unequivocal ganglion cell component admixed with a neoplastic glial component. Tumors that were better classified as other diagnostic entities (e.g. pleomorphic xanthoastrocytoma, pilocytic astrocytoma, DNET, PLNTY, multinodular and vacuolating neuronal tumor of the cerebrum [MVNT], and low-grade glial/glioneuronal neoplasm not further classifiable) were excluded. Histologic features including morphology of the glial component and presence of eosinophilic granular bodies, Rosenthal fibers, calcifications, myxoid background, CD34-immunopositive ramiﬁed cells, perivascular lymphocytes, mitotic activity, necrosis, microvascular proliferation, and leptomeningeal spread were assessed. Pre-operative imaging was reviewed for all available cases (n = 29) by an expert neuroradiologist (J.E.V.). Imaging features assessed were tumor location, size, circumscription, cortical involvement, subcortical white matter involvement, multinodularity, cystic component, T1 intensity, T2 intensity, contrast enhancement, calcifications, hemorrhage, and overlying bony remodeling. Clinical data was extracted from institutional electronic medical records including patient age, sex, presenting symptomatology, duration of symptoms, extent of surgery, adjuvant therapy, and follow-up interval. Event-free survival was defined as time until recurrence after gross total resection or disease progression after subtotal resection based on either imaging impression or pathologic conﬁrmation.

**Genomic DNA extraction and targeted next-generation sequencing**

Genomic DNA was extracted from tumor tissue that had been macrodissected from formalin-fixed, paraffin-embedded blocks or unstained sections using the QIAamp DNA FFPE Tissue Kit (Qiagen) according to the manufacturer’s protocol. Tumor tissue from the initial resection was used in 35 patients, and tumor tissue from a second surgery after recurrence/progression was used in 5 patients (SF-GG-3, SF-GG-5, SF-GG-18, SF-GG-23, and SF-GG-35). Capture-based next-generation DNA sequencing was performed as previously described at the UCSF Clinical Cancer Genomics Laboratory, using an assay that targets all coding exons of 479 cancer-related genes, TERT promoter, select introns and upstream regulatory regions of 47 genes to enable detection of structural variants including gene fusions, and DNA segments at regular intervals along each chromosome to enable genome-wide copy number and zygosity analysis, with a total sequencing footprint of 2.8 Mb (UCSF500 Cancer Panel; Additional file 1: Table S1) [20]. Sequencing libraries were prepared from genomic DNA, and target enrichment was performed by hybrid capture using a custom oligonucleotide library (Roche NimbleGen). Sequencing was performed on an Illumina HiSeq 2500. Duplicate sequencing reads were removed computationally to allow for accurate allele frequency determination and copy number calling. The analysis was based on the human reference sequence (NCBI build 37) using the following software packages: BWA, Samtools, Picard tools, GATK, CNVkit, Pindel, SATK, Annovar, Freebayes, and Delly. Single nucleotide variants, insertions/deletions, and structural variants were visualized and verified using the Integrated Genome Viewer. Genome-wide copy number analysis based on on-target and off-target reads was performed by CNVkit and Nexus Copy Number (Biodiscovery). As the majority of cases were analyzed as tumor-only without a paired normal sample to accurately confirm the somatic status of variants, only those variants classified as pathogenic or likely pathogenic are reported herein. Variants of unknown signiﬁcance are not reported, given that the vast majority of these likely represent rare or private germline variants and not somatic mutations.

**Statistical analysis**

Statistical analysis was performed using GraphPad Prism software version 7. Kaplan-Meier event-free survival analysis for patients with ganglioglioma stratified by molecular alterations was performed using Log-rank (Mantel-Cox) test. Comparison of clinical, imaging, and histologic features stratified by molecular alterations was performed using Fisher’s exact test.
Results

demographic and clinical features of the ganglioglioma cohort

Forty patients with pathologically confirmed ganglioglioma were included in this study (Table 1 and Additional file 1: Table S2). The 23 male and 17 female patients ranged from 0 to 64 years of age (median 21 years). The presenting symptoms were variable and ranged from seizures in patients with temporal lobe tumors to extremity weakness in patients with spinal cord tumors. Thirty-one tumors (78%) were located in the cerebral hemispheres with 19 in the temporal lobe, three in the frontal lobe, four in the parietal lobe, and five in the occipital lobe. Four tumors were located in the cerebellum, two were located in the thalamus, and three were located in the spinal cord (Additional file 1: Table S3). The available clinical follow-up after initial surgical intervention ranged from 0 to 29 years (median 1.8 years). Gross total resection was achieved in 26 patients, two of which had subsequent tumor recurrence at 1.2 and 7.8 years. Subtotal resection was performed in 11 patients, four of which showed subsequent tumor progression (0.6 to 10 years later). Extent of resection was unknown in three patients, two of which had subsequent tumor progression at 1.4 and 1.8 years.

Histopathologic features of the ganglioglioma cohort

All 40 gangliogliomas contained dysmorphic ganglion cells admixed with a neoplastic glial component (Additional file 1: Table 4 and Additional file 2: Figure S1). The glial component demonstrated astrocytic morphology in 37 cases (93%) and oligodendrogial morphology in three cases (8%). Eosinophilic granular bodies were present in 27 cases (68%), and Rosenthal fibers were present in six cases (15%). Calcifications were present in 19 cases (48%), and were extensive in eight of these. CD34 immunopositive ramified cells were present in 16 of 19 evaluated cases (84%). None of the tumors harbored anaplastic features, such as high mitotic index (more than 2 per 10 high power fields), necrosis, or microvascular proliferation.

Genetic alterations identified in the ganglioglioma cohort

Targeted next-generation sequencing that provides assessment of mutations, gene fusions, amplifications, deletions, and chromosomal copy number alterations was performed on the cohort of 40 gangliogliomas (Fig. 1 and Additional file 1: Tables S5–S7). Twenty-seven of the tumors harbored pathogenic alterations in the BRAF oncogene, including 18 with p.V600E hotspot mutation, five with non-V600E variant mutations (p.L505delinsLEYLS, p.R506delinsRVL[R in two cases], p.R506delinsRSTQ, and p.T599_W604delinsTDG), and four with in-frame gene fusions (two with KIAA1549 as the fusion partner, one with KLHL7, and one with CDC42BPB). In those 13 gangliogliomas lacking identifiable BRAF alteration, nine contained other genetic alterations predicted to activate the MAP kinase signaling pathway. Two harbored KRAS p.Q61K hotspot mutation, one harbored an in-frame ERC2-RAFI gene fusion, one harbored a hotspot missense mutation (p.N546K) in the kinase domain of FGFR1, one harbored an in-frame FGFR1-TACC1 gene fusion, one harbored a mutation affecting the exon 17 splice acceptor sequence of the FGFR2 gene, and two harbored in-frame FGFR2 gene fusions (one with INA as the fusion partner and the other with KIAA1598). One patient with a clinical diagnosis of neurofibromatosis type 1 harbored a germline heterozygous frameshift mutation in the NF1 gene with somatic loss of the remaining wildtype allele in the tumor. These genetic alterations involving BRAF, KRAS, RAF1, NF1, FGFR1, and FGFR2 were mutually exclusive (i.e. no tumor harbored any two of these variants simultaneously). In total, 36 of the 40 tumors (90%) were identified to harbor a genetic alteration predicted to cause activation of the MAP kinase signaling pathway. Among the remaining four tumors, three did not contain identifiable pathogenic alterations, and one epilepsy-associated ganglioglioma in the temporal lobe of a young child (SF-GG-37) was found to harbor a novel ABL2-GAB2 gene fusion predicted to result in an in-frame fusion protein containing the entirety of the kinase domain of the encoded Abelson-related protein tyrosine kinase, similar to the ABL2 fusions that have been described in a subset of pediatric leukemias [32, 34]. Three gangliogliomas with BRAF p.V600E mutation had concurrent CDKN2A homozygous deletion (SF-GG-3, SF-GG-9, and SF-GG-11) and one of these three tumors additionally harbored a subclonal missense mutation in the PTEN tumor suppressor gene (SF-GG-3). Otherwise, no additional pathogenic mutations, fusions, amplifications, or deletions were identified in any of the 40 gangliogliomas. As such, the BRAF, KRAS, RAF1, NF1, FGFR1, or FGFR2 variants were the solitary pathogenic alteration identified in 33 cases (83%). No tumors harbored pathogenic alterations affecting the IDH1, IDH2, H3F3A, HIST1H3B, HIST1H3C, SETD2, TP53, ATRX, TERT (including promoter region), CIC, FLI1P1, MYB, MYBL1, EGRF, PDGFERA, MET, PIK3CA, PIK3RI, MAP2K1, PRKCA, BCOR, BCORLI, NTRK1, NTRK2, NTRK3, ALK, RELA, or NF2 genes.

Chromosomal copy number analysis revealed no chromosomal gains, losses, or focal amplifications or deletions in 26 of the gangliogliomas (Additional file 1: Table S7). Among the other 14 cases, the quantity of chromosomal aberrations per tumor ranged from one to
<table>
<thead>
<tr>
<th>Tumor ID</th>
<th>Age/sex</th>
<th>Tumor location</th>
<th>Presenting symptoms</th>
<th>Radiographic pattern</th>
<th>Glial component</th>
<th>Pathogenic genetic alterations identified</th>
<th>Chromosomal gains/losses</th>
<th>Extent of resection</th>
<th>Recurrence or progression</th>
<th>Time to recurrence or progression (years)</th>
<th>Length of follow-up (years)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SF-GG-01</td>
<td>5 M</td>
<td>Temporal lobe</td>
<td>seizures</td>
<td>cystic and solid</td>
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<td>BRAF p.V600E</td>
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<td>0.1</td>
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<td>8 M</td>
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<td>astrocytic</td>
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<td>0.4</td>
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<td>Occipital lobe</td>
<td>seizures</td>
<td>N/A</td>
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<td>+distal 3q, -9</td>
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<td>1.8</td>
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<td>0.0</td>
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<td>gross total</td>
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<td>0.0</td>
<td>0.0</td>
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<td>SF-GG-08</td>
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<td>BRAF p.V600E</td>
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<td>0.1</td>
<td>1.8</td>
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<td>SF-GG-09</td>
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<td>BRAF p.V600E, CDKN2A/B HD</td>
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<td>gross total</td>
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<td>0.4</td>
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<td>SF-GG-10</td>
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<td>9.2</td>
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<td>BRAF p.V600E</td>
<td>-1p, +19p</td>
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<tr>
<td>SF-GG-14</td>
<td>15 F</td>
<td>Cerebellum</td>
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<td>BRAF p.V600E</td>
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<td>148</td>
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<td>BRAF p.V600E</td>
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<td>30 F</td>
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<td>BRAF p.V600E</td>
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<td>Thalamus</td>
<td>headache</td>
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<td>BRAF p.V600E</td>
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<td>gross total</td>
<td>no</td>
<td>14.1</td>
<td>14.1</td>
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<td>SF-GG-20</td>
<td>19 F</td>
<td>Temporal lobe</td>
<td>seizures</td>
<td>solid</td>
<td>astrocytic</td>
<td>BRAF p.R306delinsRVLR</td>
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<td>gross total</td>
<td>no</td>
<td>0.2</td>
<td>0.2</td>
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<tr>
<td>SF-GG-21</td>
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<td>Temporal lobe</td>
<td>seizures</td>
<td>cystic and solid</td>
<td>astrocytic</td>
<td>BRAF p.R306delinsRSTQ + 5, + 6, + 7, + 11, + 18</td>
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<td>no</td>
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<td>1.8</td>
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<td>SF-GG-22</td>
<td>23 F</td>
<td>Temporal lobe</td>
<td>seizures</td>
<td>cyst with mural nodule</td>
<td>astrocytic</td>
<td>BRAF p.L505delinsELYLS</td>
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<td>SF-GG-23</td>
<td>41 F</td>
<td>Parietal lobe</td>
<td>visual distortion</td>
<td>solid</td>
<td>astrocytic</td>
<td>BRAF p.R306delinsRVLR</td>
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<td>8.1</td>
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<td>Spinal cord</td>
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<td>KIAA1549-BRAF fusion</td>
<td>none</td>
<td>subtotal</td>
<td>no</td>
<td>0.2</td>
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</tr>
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</table>
Table 1 Summary of the clinicopathologic features and molecular alterations in the ganglioglioma patient cohort (Continued)

<table>
<thead>
<tr>
<th>Tumor ID</th>
<th>Age/sex</th>
<th>Tumor location</th>
<th>Presenting symptoms</th>
<th>Radiographic pattern</th>
<th>Glial component</th>
<th>Pathogenic genetic alterations identified</th>
<th>Chromosomal gains/losses</th>
<th>Extent of resection</th>
<th>Recurrence or progression</th>
<th>Time to recurrence or progression (years)</th>
<th>Length of follow-up (years)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SF-GG-25</td>
<td>17 F</td>
<td>Spinal cord</td>
<td>leg weakness</td>
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<td>astrocytic</td>
<td>KIAA1549-BRAF fusion</td>
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<td>gross total</td>
<td>no</td>
<td>4.5</td>
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<td>SF-GG-26</td>
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<td>Temporal lobe</td>
<td>seizures</td>
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<td>astrocytic</td>
<td>CDC42BPB-BRAF fusion</td>
<td>-interstitial 17p</td>
<td>subtotal</td>
<td>yes (R)</td>
<td>2.2</td>
<td>2.2</td>
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<td>SF-GG-27</td>
<td>8 F</td>
<td>Occipital lobe</td>
<td>seizures</td>
<td>solid</td>
<td>astrocytic</td>
<td>KIAA1549-BRAF fusion</td>
<td>+ 7</td>
<td>gross total</td>
<td>no</td>
<td>1.4</td>
<td>1.4</td>
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<tr>
<td>SF-GG-28</td>
<td>48 M</td>
<td>Frontal lobe</td>
<td>seizures</td>
<td>cyst with mural nodule</td>
<td>astrocytic</td>
<td>ERC2-RAF1 fusion</td>
<td>+ 3, -interstitial 3p, + 5, + 7</td>
<td>gross total</td>
<td>no</td>
<td>0.5</td>
<td>0.5</td>
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<tr>
<td>SF-GG-29</td>
<td>32 F</td>
<td>Insula</td>
<td>seizures</td>
<td>cyst with mural nodule</td>
<td>astrocytic</td>
<td>KRAS p.Q61K</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>0.0</td>
<td>0.0</td>
</tr>
<tr>
<td>SF-GG-30</td>
<td>24 M</td>
<td>Temporal lobe</td>
<td>seizures</td>
<td>cyst with mural nodule</td>
<td>astrocytic</td>
<td>KRAS p.Q61K</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>SF-GG-31</td>
<td>28 M</td>
<td>Temporal lobe</td>
<td>headaches</td>
<td>cystic and solid</td>
<td>astrocytic</td>
<td>germline NF1 p.Q514 frameshift w/ somatic LOH</td>
<td>+ 4, + 5, + 7, + 8, + 9, + 12, + 15, -17p, -18p, + 22</td>
<td>gross total</td>
<td>no</td>
<td>5.9</td>
<td>5.9</td>
</tr>
<tr>
<td>SF-GG-32</td>
<td>35 M</td>
<td>Frontal lobe</td>
<td>paresthesias</td>
<td>solid</td>
<td>oligodendroglial</td>
<td>FGFR1-TACC1 fusion</td>
<td>none</td>
<td>subtotal</td>
<td>no</td>
<td>1.4</td>
<td>1.4</td>
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<tr>
<td>SF-GG-33</td>
<td>59 F</td>
<td>Temporal lobe</td>
<td>seizures</td>
<td>multicystic</td>
<td>astrocytic</td>
<td>FGFR1 p.N546 K</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>5.9</td>
<td>5.9</td>
</tr>
<tr>
<td>SF-GG-34</td>
<td>7 M</td>
<td>Temporal lobe</td>
<td>seizures</td>
<td>N/A</td>
<td>oligodendroglial</td>
<td>FGFR2-KIAA1598 fusion</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>5.3</td>
<td>5.3</td>
</tr>
<tr>
<td>SF-GG-35</td>
<td>35 M</td>
<td>Parietal lobe</td>
<td>unknown</td>
<td>complex heterogeneous</td>
<td>oligodendroglial</td>
<td>FGFR2-IN1A fusion</td>
<td>-portions of 10</td>
<td>unknown</td>
<td>yes (P)</td>
<td>9.1</td>
<td>9.1</td>
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<td>SF-GG-36</td>
<td>37 F</td>
<td>Parietal lobe</td>
<td>leg weakness</td>
<td>N/A</td>
<td>astrocytic</td>
<td>FGFR2 exon 17 splice site mutation</td>
<td>none</td>
<td>subtotal</td>
<td>no</td>
<td>7.5</td>
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<tr>
<td>SF-GG-37</td>
<td>2 M</td>
<td>Frontal lobe</td>
<td>seizures</td>
<td>microcystic</td>
<td>astrocytic</td>
<td>ABL2-GAB2 fusion</td>
<td>none</td>
<td>subtotal</td>
<td>no</td>
<td>0.3</td>
<td>0.3</td>
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<tr>
<td>SF-GG-38</td>
<td>53 F</td>
<td>Occipital lobe</td>
<td>asymptomatic</td>
<td>N/A</td>
<td>astrocytic</td>
<td>none identified</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>0.0</td>
<td>0.0</td>
</tr>
<tr>
<td>SF-GG-39</td>
<td>0 F</td>
<td>Cerebellum</td>
<td>nystagmus</td>
<td>cystic and solid</td>
<td>astrocytic</td>
<td>none identified</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>14.9</td>
<td>14.9</td>
</tr>
<tr>
<td>SF-GG-40</td>
<td>10 M</td>
<td>Spinal cord</td>
<td>arm weakness</td>
<td>N/A</td>
<td>astrocytic</td>
<td>none identified</td>
<td>none</td>
<td>gross total</td>
<td>no</td>
<td>14.3</td>
<td>14.3</td>
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</tbody>
</table>

*HD Homozygous deletion. LOH Loss of heterozygosity. (s), subclonal. (R), radiographic recurrence/progression. (P), pathologically confirmed recurrence/progression*
ten. In the majority of cases, chromosomal copy number changes were limited to gains and losses of whole chromosomes or chromosome arms, without focal gains or losses. No focal amplifications or homozygous deletions were identified other than the three gangliogliomas with focal CDKN2A homozygous deletion. Tumor SF-GG-35 demonstrated multiple regions of segmental loss involving chromosome 10 (containing the FGFR2 and INA loci) consistent with the process of chromosome shattering that has been termed chromothripsis, which was the likely mechanism of generating the FGFR2-INA gene fusion seen in this tumor. Recurrent chromosomal copy number changes in this cohort included trisomy 7 (containing the BRAF locus) seen in six tumors, trisomy 5 seen in five tumors, trisomy 12 seen in three tumors, monosomy 9 seen in three tumors, and monosomy 1p seen in three tumors. Four of the six tumors with trisomy 7 were those harboring BRAF alterations and likely involved gain of the mutant or fused allele. All three of the tumors with monosomy 9 were those harboring a focal deletion event involving the remaining copy of chromosome 9p21 including the CDKN2A gene, resulting in homozygous/biallelic deletion. All three of the tumors with monosomy 1p were located in the cerebral hemispheres of adults and harbored BRAF p.V600E mutation (SF-GG-07, SF-GG-11, and SF-GG-13).

**Association of genetic alterations with clinical and imaging features**

The age at initial diagnosis was not significantly different among patients with gangliogliomas stratified by BRAF p.V600E mutation versus other BRAF alteration, any BRAF alteration versus BRAF wildtype status, or any BRAF alteration versus FGFR alteration (Table 2). Regarding location, all FGFR altered gangliogliomas were located in the cerebral hemispheres, whereas BRAF altered tumors were located throughout the neuraxis. The two thalamic gangliogliomas both harbored BRAF p.V600E mutation, three of the four cerebellar gangliogliomas harbored BRAF p.V600E mutation, and two of three gangliogliomas centered in the spinal cord harbored KIAA1549-BRAF fusion. The remaining cerebellar and spinal cord tumors lacked identifiable pathogenic alterations. The two tumors harboring BRAF fusion with partners other than KIAA1549 were both located in the cerebral hemispheres. All tumors with variant BRAF mutations, KRAS mutation, RAF1 fusion, NF1 mutation, and FGFR alterations were located in the cerebral hemispheres. Imaging features including tumor size, presence

![Oncoprint summary table](image-url)
of a cystic component, circumscription, and contrast enhancement did not show significant correlation with underlying genetic alterations (Table 2 and Additional file 1: Table S3).

**Association of genetic alterations with histologic features**

All three gangliogliomas with a glial component showing oligodendroglial morphology harbored FGFR alterations (Fig. 2 and Additional file 2: Figure S1). However, the other two gangliogliomas with FGFR alterations had a glial component with astrocytic morphology. All BRAF, KRAS, NF1, and RAF1 altered tumors had a glial component with astrocytic morphology. Except for the morphology of the glial component, none of the other histologic features including presence/absence of eosinophilic granular bodies, Rosenthal fibers, calcifications, and perivascular lymphocytes showed a significant correlation with underlying genetic alterations (Table 2 and Additional file 1: Table S4).

**Association of genetic alterations with disease recurrence or progression**

In the two gangliogliomas that recurred after gross total resection, sequencing analysis that was performed on the recurrent tumors demonstrated BRAF p.R506del/insRVLG mutation as the solitary pathogenic alteration without chromosomal copy number alterations in one case. The other demonstrated BRAF p.V600E mutation, CDKN2A homozygous deletion, a subclonal missense mutation in the PTEN tumor suppressor gene, and only two chromosomal copy number aberrations (gain of distal 3q and loss of 9). In the four gangliogliomas that showed tumor progression after subtotal resection, three harbored BRAF p.V600E mutation as the solitary pathogenic alteration without chromosomal copy number aberrations, and the fourth tumor harbored CDC42BPB-BRAF gene fusion. In the two gangliogliomas that progressed after initial resection of unknown extent, sequencing analysis that was performed on the recurrent tumors demonstrated FGFR2-INA fusion in one and BRAF p.V600E mutation in the other. Event-free survival of the patient cohort stratified by BRAF altered versus BRAF wildtype status, BRAF V600E mutant versus other BRAF alteration, BRAF altered versus FGFR altered, and BRAF V600E mutant/CDKN2A intact versus BRAF V600E mutant/CDKN2A deleted was assessed (Fig. 3). No significant differences in event-free survival were found based on underlying genetic alterations in this cohort.

**Discussion**

This study reveals that ganglioglioma is genetically defined by alterations that activate the MAP kinase signaling pathway in the vast majority of cases, either via BRAF p.V600E mutation or a spectrum of other genetic alterations including alternative BRAF mutations or...
fusions, RAF1 fusion, KRAS mutation, NF1 mutation, or FGFR mutations or fusions. In the majority of cases, the genetic alteration within the MAP kinase pathway was the solitary genetic alteration identified, with few (if any) chromosomal copy number changes, indicating that most gangliogliomas are genetically simple tumors. As such, gangliogliomas are genetically similar to pilocytic astrocytoma, DNET, rosette-forming glioneuronal tumor (RGNT), PLNTY, and MVNT. Gangliogliomas more frequently harbor BRAF p.V600E mutation or other variant BRAF mutations than pilocytic astrocytomas, which most commonly harbor KIAA1549-BRAF fusion [19, 30, 38]. One study of posterior fossa and spinal cord gangliogliomas with a glial component resembling pilocytic astrocytoma found that a subset harbored KIAA1549-BRAF fusion that the authors referred to “pilocytic astrocytoma with focal gangliocytic differentiation” [16]. The two gangliogliomas in our cohort with KIAA1549-BRAF fusion were both located in the spinal cord of children and demonstrated numerous dysmorphic ganglion cells throughout the entirety of the tumor, indicating that classic gangliogliomas can also harbor KIAA1549-BRAF fusion. While the majority of DNETs and RGNTs harbor FGFR1 mutation or rearrangement [14, 33, 37], this is only present in a small subset of pathologically-confirmed gangliogliomas. The recently described entity PLNTY has been reported to harbor either FGFR fusions or BRAF V600E mutation [17], which overlaps with the genetic alterations observed in gangliogliomas. Our recent genetic analysis revealed that MVNTs harbor frequent MAP2K1 exon 2 mutations or small in-frame deletions, as well as BRAF mutations other than V600E [28]; however, MAP kinase pathway activation in gangliogliomas appears to occur independently of MAP2K1 alterations.

Pleomorphic xanthoastrocytoma is a circumscribed glial neoplasm that is genetically characterized by concurrent CDKN2A homozygous deletion and BRAF p.V600E mutation (or less commonly BRAF or RAF1 fusion) [29, 38]. However, our study shows that a small subset of pathologically-confirmed gangliogliomas can

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**Fig. 2** Histologic features of gangliogliomas with different genetic alterations in the MAP kinase signaling pathway. 


d. Ganglioglioma in the spinal cord of a 5 year old girl with KIAA1549-BRAF gene fusion (SF-GG-24). 

e. Ganglioglioma in the frontal lobe of a 48 year old man with ERC2-RAF1 gene fusion (SF-GG-28). 

f. Ganglioglioma in the temporal lobe of a 7 year old boy with FGFR2-KIAA1598 gene fusion (SF-GG-34).
harbor this identical combination of CDKN2A homozygous deletion and BRAF p.V600E mutation, indicating that this genetic pattern is not entirely specific to pleomorphic xanthoastrocytomas. Another recently described tumor entity is diffuse leptomeningeal glioneuronal tumor (also referred to as disseminated oligodendroglioma-like leptomeningeal neoplasm), which is genetically characterized by the combination of monosomy 1p and KIAA1549-BRAF fusion [35]. Three gangliogliomas in our cohort harbored the combination of monosomy 1p and BRAF p.V600E mutation, all of which were intraparenchymal tumors located in the cerebral hemispheres of adults and did not display widespread leptomeningeal dissemination. Of note, a series of intramedullary low-grade glioneuronal tumors of the spinal cord in children harboring BRAF fusion and monosomy 1p without diffuse leptomeningeal spread was recently reported [8]. The relationship of these pediatric spinal tumors and our adult cerebral gangliogliomas harboring the combination of monosomy 1p and BRAF p.V600E mutation is uncertain.

The genetic profile of ganglioglioma appears to be distinct from several glial and glioneuronal neoplasms. No PRKCA fusions or kinase domain mutations were identified in any of the cases, suggesting that gangliogliomas are genetically distinct from the majority of papillary glioneuronal tumors and chordoid gliomas [3, 15]. No IDH1, IDH2, TP53, ATRX, TERT promoter, CIC, or FUBP1 mutations were identified in any of the cases, suggesting that gangliogliomas are genetically distinct from the majority of diffuse lower-grade gliomas in adults (both astrocytomas and oligodendrogliomas) [4]. No MYB or MYBL1 rearrangements were identified in any of the cases, suggesting that gangliogliomas are genetically distinct from the majority of angiocentric gliomas and pediatric IDH-wildtype diffuse astrocytomas [1, 30, 31, 38]. No TSC1 or TSC2 mutations were identified in any of the cases, suggesting that gangliogliomas are also genetically distinct from the majority of subependymal giant cell astrocytomas [5].

Malformations of cortical development, including focal cortical dysplasia, constitute one of the major differential diagnoses for ganglioglioma. Genetic evaluation of sporadic focal cortical dysplasias (not associated with another lesion) has revealed frequent post-zygotic somatic mutations in components of the PI3-kinase-Akt-mTOR signaling pathway, most often involving the TSC1, TSC2, AKT3, MTOR, PIK3CA, or PTEN genes [10, 18, 23–26]. None of the gangliogliomas in this cohort showed genetic alterations in components of this pathway, except for one ganglioglioma that recurred after gross total resection and harbored a subclonal PTEN missense mutation (in addition to BRAF p.V600E mutation and CDKN2A homozygous deletion). This indicates that the
PTEN mutation was likely acquired during tumor progression and was not the initiating genetic driver. Thus, gangliogliomas appear to be genetically distinct from the majority of sporadic focal cortical dysplasias, which suggests that genetic evaluation may be potentially informative in cortical resection cases that are challenging to classify based on morphologic features.

Four of the gangliogliomas in this cohort harbored recurrent small in-frame insertions at codon 505 or 506 in the β3-αC loop in the kinase domain of BRAF (p.L505delinsLEYLS, p.R506delinsRVLR [in two cases], and p.R506delinsRSTQ). Among the 52,519 tumors with BRAF mutations currently cataloged in the COSMIC database [version 85 release], only one other tumor (medulloblastoma) with a small in-frame insertion at this site is present. Given this recurrent BRAF alteration in a tumor type with frequent MAP kinase pathway activation and low somatic mutation burden, together with a lack of other identifiable alterations in MAP kinase pathway genes in these four tumors, this very likely represents a novel hotspot BRAF mutation causing activation of the serine/threonine kinase domain in gangliogliomas.

Four of the gangliogliomas in this cohort lacked identifiable alterations in canonical genes associated with the MAP kinase pathway. These cases may potentially harbor cryptic alterations in MAP kinase genes that were not detectable by this sequencing assay. Alternatively, these tumors may harbor novel molecular alterations and represent rare molecular subtypes of ganglioglioma or other glioneuronal tumors. Indeed, one of these four tumors was identified to harbor a novel ABL2-GAB2 gene fusion. Whether this fusion leads to downstream activation of the MAP kinase pathway similar to most other gangliogliomas, or instead drives proliferation via modulation of other intracellular signaling pathway is unknown.

Our study does not reveal any differences in genetic profile of gangliogliomas that correlate with disease progression or recurrence. This may be due to the small size of the cohort in this study, particularly those with less common variants such as RAF1 fusion or KRAS mutation. However, as the predicted biologic consequence of the less common MAP kinase variants identified in this study is activation of the same Ras-Raf-MEK-ERK signaling pathway as BRAF p.V600E mutation, we hypothesize that the specific MAP kinase pathway alteration is unlikely to dictate differences in clinical behavior. Instead, other factors such as tumor location, extent of resection, accompanying genetic alterations, and/or epigenetic differences are more likely to drive the clinical variability in presentation and outcome for patients with ganglioglioma.

**Additional files**

**Additional file 1:** Tables S1-S7. (XLSX 44 kb)

**Additional file 2:** Figures S1-S2. (PDF 7457 kb)

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**Availability of data and materials**

Sequencing data files are available from the authors upon request.

**Competing interest**

The authors declare that they have no competing interests.

**Ethics approval and consent to participate**

This study was approved by the Committee on Human Research of the University of California, San Francisco, with a waiver of patient consent.

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**Author details**

1. Department of Pathology, University of California, San Francisco, CA, USA. 2. Department of Radiology and Biomedical Imaging, University of California, San Francisco, CA, USA. 3. Clinical Cancer Genomics Laboratory, University of California, San Francisco, CA, USA. 4. Department of Pathology, Nicklaus Children’s Hospital, Miami, FL, USA. 5. Department of Pediatric Hematology/Oncology, Nicklaus Children’s Hospital, Miami, FL, USA. 6. Department of Pathology, Neurology, and Neurosurgery, University of Colorado, Aurora, CO, USA. 7. Division of Pediatric Hematology/Oncology, Valley Children’s Hospital, Madera, CA, USA. 8. Division of Pediatric Hematology/Oncology, Department of Pediatrics, University of California, San Francisco, CA, USA. 9. Division of Neurosurgery, University of California, San Francisco, CA, USA. 10. Department of Neurology, University of California, San Francisco, CA, USA. 11. Division of Neuro-Oncology, Department of Neurological Surgery, University of California, San Francisco, CA, USA. 12. Division of Pediatric Hematology/Oncology, UCSF Benioff Children’s Hospital Oakland, Oakland, CA, USA.

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**References**


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