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Authors
Fuller, Trevon L
Saatchi, Sassan S
Curd, Emily E
et al.

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Mapping the risk of avian influenza in wild birds in the US

Trevon L Fuller*1, Sassan S Saatchi1,2, Emily E Curd1,3, Erin Toffelmier1, Henri A Thomassen1, Wolfgang Buermann1,4, David F DeSante5, Mark P Nott5, James F Saracco5, CJ Ralph6, John D Alexander7, John P Pollinger1 and Thomas B Smith1,3

Abstract

Background: Avian influenza virus (AIV) is an important public health issue because pandemic influenza viruses in people have contained genes from viruses that infect birds. The H5 and H7 AIV subtypes have periodically mutated from low pathogenicity to high pathogenicity form. Analysis of the geographic distribution of AIV can identify areas where reassortment events might occur and how high pathogenicity influenza might travel if it enters wild bird populations in the US. Modelling the number of AIV cases is important because the rate of co-infection with multiple AIV subtypes increases with the number of cases and co-infection is the source of reassortment events that give rise to new strains of influenza, which occurred before the 1968 pandemic. Aquatic birds in the orders Anseriformes and Charadriiformes have been recognized as reservoirs of AIV since the 1970s. However, little is known about influenza prevalence in terrestrial birds in the order Passeriformes. Since passerines share the same habitat as poultry, they may be more effective transmitters of the disease to humans than aquatic birds. We analyze 152 passerine species including the American Robin (Turdus migratorius) and Swainson’s Thrush (Catharus ustulatus).

Methods: We formulate a regression model to predict AIV cases throughout the US at the county scale as a function of 12 environmental variables, sampling effort, and proximity to other counties with influenza outbreaks. Our analysis did not distinguish between types of influenza, including low or highly pathogenic forms.

Results: Analysis of 13,046 cloacal samples collected from 225 bird species in 41 US states between 2005 and 2008 indicates that the average prevalence of influenza in passerines is greater than the prevalence in eight other avian orders. Our regression model identifies the Great Plains and the Pacific Northwest as high-risk areas for AIV. Highly significant predictors of AIV include the amount of harvested cropland and the first day of the year when a county is snow free.

Conclusions: Although the prevalence of influenza in waterfowl has long been appreciated, we show that 22 species of song birds and perching birds (order Passeriformes) are influenza reservoirs in the contiguous US.

Background

There is a strong link between influenza in birds and human health because influenza epidemics in human populations occur when viruses that typically inhabit the avian gastrointestinal tract mutate or reassort, enabling them to cross the species barrier to infect people [1]. Mutations arise in avian influenza virus (hereafter “AIV”) due to the high error rate of influenza RNA polymerase and the large population size and short generation time of the virus [2]. Reassortment is the exchange of RNA segments between distinct influenza viruses. When human influenza viruses and AIV reassort, they produce offspring virions that represent a mixture of the parental types’ RNA and are infectious to humans in some cases [2-4]. For example, in 1968, one million people died in an influenza pandemic that resulted from the reassortment of an influenza virus from Ukrainian ducks and a virus that had circulated in people since 1957 [5-8].

Today, outbreaks of H5N1 influenza in Africa, Asia, Europe, and the Middle East further illustrate the human
health implications of influenza in birds. (Influenza viruses are classified into "HA" and "NA" subtypes based on surface proteins.) People contract H5N1 by handling infected poultry or wild birds after which the virus binds to receptors in the pulmonary alveoli, causing pneumonia and death due to respiratory failure [9-11]. Since July 2003, there have been 436 human cases of H5N1 in the Eastern hemisphere with a 60% mortality rate [12-14]. In 75% of these cases, the infected people had contact with birds [10]. However, H5N1 has also evolved limited person-to-person transmission within human families [7,15-17]. Public health authorities are concerned that the evolution of wider human-to-human transmission could result in a H5N1 pandemic that could cause up to 142 million deaths at a cost of $US 4.4 trillion [14,18]. The ongoing human pandemic of H1N1 influenza, which has caused over 296,000 human cases and at least 15,921 deaths since mid-February 2009, contains genes from avian, human, and swine influenza viruses [12].

To date, influenza viruses have been isolated from 105 species of wild birds representing 26 families [16]. In birds, the H5 and H7 AIV subtypes have periodically mutated from a low pathogenicity form (hereafter "LPAI"), which is typically asymptomatic in wild birds, to a highly pathogenic form (hereafter "HPAI") that causes mortality rates of up to 100% in chickens [2,19-21]. (Our analysis did not distinguish between influenza subtypes or differentiate LPAI from HPAI.) HPAI also differs from LPAI in that the former has more amino acids adjacent to the hemagglutinin cleavage site, which allows it to replicate in a broader range of tissues [for details, see [2,22]].

Aside from poultry, no HPAI H5N1 has been detected to date in the US, though six LPAI H5N1 viruses have been detected in North America since 2004 [23]. AIV mutated from LP to HP form in poultry in the US in the 1920s, in 1984, and in 2004 [4,24-29]. Although none of these US outbreaks resulted in the infection of humans with HPAI, it is plausible that HPAI could reassort or mutate to become transmissible to people. As few as five amino acid changes can transform the HP influenza virus into an airborne form that is infectious to mammals [14]. In the event of an HPAI epizootic in migratory birds in the US, these species could spread HPAI across the country along migratory routes because ducks infected with HP H5 remain healthy enough to migrate [16,30]. Indeed, HPAI has already been detected in wild birds in Chad, China, Nigeria, and South Africa [30-32].

We analyze the geographic distribution of AIV in wild birds in the US with the goal of inferring where reassortment events might occur and how HPAI might travel if it enters wild bird populations. Our method for detecting the influenza virus in samples from passerines does not determine whether the virus is LPAI or HPAI (see below). However, 67% of our samples are from non-passerines and are known to be LPAI. Thus, this study assumes that most of our AIV-positive samples are LPAI. We model the geographic distribution of AIV to provide insights about how HPAI might spread if it is introduced to the US in the future. Since we cannot guarantee that the passerine samples are LPAI, when referring to samples that tested positive for influenza virus, we will use the term "AIV" rather than "LPAI". As noted in the Discussion, the characterization of the subtype and pathogenicity of AIVs isolated from passerines in the US remains an important area for future research.

Although the monitoring of HPAI viruses is important, another critical issue in AIV biosafety is the detection of H5 and H7 LPAI viruses. LPAI H7 has been transmitted directly to humans in the US in 1976, 2002, and 2003 [7,33]. These cases resulted in conjunctivitis, fever, and upper-respiratory tract symptoms of influenza-like illness, but no fatalities. LPAI H5 can mutate to HPAI relatively easily given the right environment (for example, poultry sheds). We refer the interested reader to Verdugo et al. [34], who have developed a model for detecting H5 and H7 LPAI in poultry and predicting when they will evolve to HPAI.

The aims of this research are to measure the prevalence of AIV in different species of wild birds in the US and to prioritize geographical regions for future influenza surveillance. Although aquatic birds in the orders Anseriformes and Charadriiformes have been recognized as reservoirs of AIV since the 1970s, much less is known about AIV prevalence in terrestrial birds in the order Passeriformes [16,35-38]. Examples of Anseriformes (ducks) that have high prevalence of influenza in the US include the Mallard (Anas platyrhynchos) and the Northern Pintail (Anas acuta) [39,40]. Shorebirds of the order Charadriiformes in which high influenza prevalence has been detected in the US include the Ruddy Turnstone (Arenaria interpres) and the Red Knot (Calidris canutus) [36,41]. Recent work detected high prevalence of influenza in passerines in China, including the Eurasian Tree Sparrow (Passer montanus) [42,43]. The present study is necessary in order to test the hypothesis that passerines are important reservoirs of AIV in the US. Further motivation for our study comes from the fact that public health agencies have limited funding to test wild birds for AIV. Thus, it is crucial that the establishment of surveillance sites should be as efficient as possible. For example, the number of sites that are monitored should be small but the sites should be located in counties that are most likely to have birds with AIV. We aim to test the hypothesis that environmental variables can be used to predict AIV cases in wild birds. Next, based on the relationship between AIV and environmental predictors, we attempt to identify the US counties most likely to be influenza hotspots for wild birds.
Our study makes the following contributions. First, guidelines formulated by the World Health Organization recognize the importance of epidemiological modelling using tools such as GIS for the control of AIV [44]. Nevertheless, most previous work on the geographic distribution of AIV has analyzed Asia and Africa [e.g. [35,38,45]]. To date, studies of AIV in wild birds in the US have focused on Alaska [39,46,47]. However, there may be overlooked hotspots of AIV in the contiguous US [48]. We contribute the first predictions about AIV cases in the contiguous US at the county scale. Second, we analyze new passerine samples from the Atlantic, Mississippi, and Pacific Flyways supplemented with existing samples from online databases to provide the first comprehensive assessment of AIV prevalence in US passersines. The main finding reported in this article is that the prevalence of influenza in passersines is greater than the prevalence in eight other avian orders. The implication of this finding for human health is that, along with poultry and waterfowl, passersines in the US are a potential vector for the transmission of AIV to people [16,49].

**Methods**

**Influenza samples from wild birds**

The data set comprised 13,046 samples from 136 counties or parishes in 41 US states (Figure 1, Additional file 1, Table S1, and Additional file 2, Table S1). The new samples included in this analysis comprise cloacal swabs collected from December 2005 to 2008 primarily during the Spring breeding season at banding stations that are part of the Monitoring Avian Productivity and Survival (MAPS) program, during the Fall as part of the Landbird Monitoring Network of the Americas (LaMNA), and during the Winter as part of the Monitoring Avian Winter Survival (MAWS) program in collaboration with UCLA’s Center for Tropical Research [50,51]. Since it is routine for the samples collected by the banding network to be stored at room temperature for prolonged periods, we utilized 100% ethanol as a storage medium. Viral RNA was extracted using a commercial magnetic bead kit (Ambion MagMAX viral RNA isolation kit). The vRNA was then converted to cDNA in a real-time reverse transcription PCR reaction using an Ambion AgPath-ID one-step PCR kit and run on a 7900 HT Fast Real-Time PCR System. PCR primers targeted a conserved region of the Matrix 1 gene: 5-GAR ATC GCG CAG ARA CTT GA-3 and 5-CAC TGG GCA CGG TGA GC-3 are forward and reverse primers, respectively. For analysis of the sensitivity of the MagMAX and AgPath-ID kits, see [47]. We used High Resolution Melt Analysis to identify putative positives based on melt temperature and then confirmed amplicon length (143 bp) on a 3% agarose gel. Viral cDNA was purified using a Zymoclean DNA gel recovery kit. We used the same PCR primers in BigDye Terminator sequencing reactions and then products were run on an Applied Biosystems 3730 system. Chromatograms were visualized in Geneious and then confirmed as AIV through a BLAST search of the NCBI database. The field and laboratory methods utilized in this study were approved by an Institutional Review Board at the University of California, Los Angeles. AIV-positive samples have been deposited in GenBank under Accession Numbers HM355888 to HM355917.

Since our samples were cloacal swabs with no measurable volume, we could not estimate the viral load of influenza in passersines defined as the number of copies of the virus per unit of body fluid. We can nevertheless infer that our copy number is very low based on cycle of threshold (Ct) values, which is the cycle at which the samples began to amplify. Since the Ct values were fairly high, we estimate that our copy number is uniformly low across the passerine samples. (Ct values >35 are typically interpreted as indicating the absence of influenza in passersines [52].)

**Environmental variables**

We investigated the association between AIV cases in wild birds in the contiguous US and 12 predictor variables, which measured agricultural and commercial activity as well as climate (Table 1). The predictor variables are all publicly available. First, we analyzed measures of agricultural activity because rice crop production is correlated with H5N1 outbreaks in Southeast Asia [38]. We included the amount of harvested cropland in the county in units of hectares, the mean size of farms in the county, and the % of the county in cropland [53]. We also examined the % of family-owned farms in the county because the use of employees who were not family members was significant for explaining AIV outbreaks in Virginia in 2002 [26]. In addition, we analyzed the density of roads in each county that connect major population centers and the human population density (as a surrogate measure of trade activity) because proximity to trade routes such as highways has been hypothesized to explain the spread of H5N1 from Asia to Europe in 2005 and proximity to roads was found to be a significant variable for explaining outbreaks of HPAI H5N1 in poultry in Romania in 2005 [54,55]. The density of poultry in the county was included in the analysis to test the hypothesis that poultry spread AIV to wild birds [56].

We incorporated total annual precipitation and minimum temperature data interpolated from weather stations using the Parameter-elevation Relationships on Independent Slopes Model (PRISM) method, which accounts for physiographic features such as terrain barriers that result in rain shadows and is considered the most accurate representation of US climate patterns [57]. Previous work has posited a relationship between tempera-
ture and influenza cases. For example, the 1918 pandemic coincided with an unusually hot winter in eastern North America and north central Asia caused by one of the strongest El Niño/Southern Oscillation (ENSO) events of the twentieth century [58]. We hypothesized that precipitation and minimum temperature might affect influenza prevalence in wild birds because AIV can be transmitted to birds in a moist environment and cold weather affects bird dispersal [59], which may influence the spread of the virus [60,61]. (For these and the following meteorological variables, we used the average value of the variable from 2006 to 2008 because our AIV samples came from December 2005 to 2008). Finally, we examined: (i) a binary variable set to one if the majority of the land in the county was ever covered by snow/ice and set to zero otherwise, (ii) the day of the year when the majority of the land in the county was first covered by snow/ice, and (iii) the day of the year when the majority of the land in the county was first free of snow/ice, all of which were inferred from satellite radar [62]. We included these variables related to freeze and thaw date because the timing of spring migration depends on the first day of thaw and fall migration depends on the first day of freeze. These variables may indirectly explain AIV spread by affecting bird dispersal [63]. Variables were iteratively removed from the spatial regression model via backward elimination until only variables with a $t$-statistic in the 95th percentile remained [64]. Forward selection gave the same result.

**Spatial regression model**

The statistical model represents the number of AIV cases per county as a Poisson-distributed random variable, which is appropriate for analyzing disease cases in which some geographic units have many cases but most units have few or no cases [65]. In the model, the number of
AIV cases per county depends on the environmental variables as well as spatial proximity to other counties with AIV cases. In particular, we utilized a spatial regression model to account for autocorrelation in AIV cases among counties that are close together geographically. Failure to address autocorrelation results in underestimation of the degrees of freedom of the data, which decreases the standard errors of the parameter estimates in a regression model [66]. Thus, ignoring autocorrelation may lead to the erroneous conclusion that a variable is significant for explaining AIV cases when the variable is in fact non-significant. The model incorporates spatial autocorrelation by constructing a semivariogram and also accounts for differences in sampling effort among counties [67]. After the model was fitted to the 136 counties for which we had AIV samples, we predicted the number of AIV cases in the other 2973 US counties by applying the model to the unsampled counties. We utilized a generalized linear mixed model, which is a form of kriging with a semivariogram, rather than a conditional autoregressive model because the latter cannot readily be extended to non-Gaussian data [67]. The Poisson distribution provided a better fit to our data on flu cases than the Gaussian distribution (see Additional file 3). The accuracy of the spatial regression was assessed using two measures. First, we calculated the generalized chi-squared statistic. If a regression model is accurate, then the generalized chi-squared statistic divided by the degrees of freedom of the data should be close to one [67,68]. The model provided a good fit to the data insofar as the generalized chi-squared statistic divided by the degrees of freedom was 0.9. Collinearity among the regressors was also acceptably low (variance inflation factors <1.75) [68]. Second, we used a leave-one-out procedure that fitted the model to the data from 135 counties and then measured the model’s accuracy on the remaining county. When the procedure was repeated 136 times, the root mean squared error (r.m.s.e.) was 6.33 AIV cases per county. This r.m.s.e. is acceptable based on the rule of thumb that the r.m.s.e. should be no greater than one-quarter of the range [69]. (For our data set, the range was 76.) Further details of the statistical model can be found in Additional file 3.

**Results**

Among the eight avian orders analyzed here, Passeriformes had the largest number of species in which AIV was detected (22 species) followed by ducks (order Anseriformes, 16 species) and shorebirds (order Charadriiformes, 1 species). We tested 4,341 samples from passerine birds, of which 0.89% were AIV-positive (Figure 2). Table 2 lists the top five species in terms of AIV-prevalence for each avian order.

**Influenza prevalence in Passeriformes**

We evaluated the hypothesis that AIV was equally prevalent among 11 avian orders by testing the null hypothesis
that AIV prevalence was the same among the orders. The null hypothesis was rejected, indicating that AIV is more common in some orders than in others (Kruskal-Wallis $KW = 124$, $df = 10$, $p < 0.0001$). Next, we ranked the orders based on AIV prevalence. The number of passerine species that tested positive for AIV ($n = 22$) was greater than the number of positive species detected in ten other orders, including waterfowl. However, our sampling was also biased towards passerines. To address this, we calculated fraction of AIV-positive samples from each order. This corrects for the fact that we had more samples from some orders than from others. The fraction of samples from passerines that tested positive for AIV was greater than the prevalence in eight other orders of birds in the contiguous US (Figure 2). We note, however, that for some of these eight orders the number of species and samples represented in our database is small. Nevertheless, AIV prevalence in Passeriformes was greater than in Falconiformes, an order in which we tested four species and 270 samples, and Piciformes, and order in which we tested 14 species and 105 samples.

**Environmental predictors of influenza prevalence in the sampled counties**

For the remainder of the analysis, we pooled the samples from all 11 orders so as to increase the geographic region represented by data. Significant predictors of the number of AIV cases per county in wild birds were thaw date, the % of harvested cropland in the county, and minimum temperature (Table 3). We interpret each of these variables in turn beginning with thaw date. Freeze/thaw dynamics have previously been hypothesized to explain the prevalence of AIV in ducks in Europe (Andrew Dobson, Princeton University, personal communication, 2009). We posit that thaw date affects influenza in wild birds in the US according to the following mechanism. Waterfowl have large populations with high annual turnover rates, so that a large fraction of the population is immunologically naïve each year [70]. In particular, hatchlings are susceptible to infection from adults via fecal-oral transmission at breeding grounds, which have densities of up to 210 birds/m² [20,37]. At these sites, adults shed the virus into the water in feces and the young are infected by ingesting the water [4]. Prevalence of AIV in gulls in the US is typically highest in spring and in summer [71]. The sign of the coefficient for thaw date is negative, indicating that if the thaw date occurs later in the year, then the number of cases of AIV is predicted to decrease [Table 3, for details see [67]]. We conjecture that thaw date explains AIV cases because if a site thaws earlier, then waterfowl can occupy the site sooner, and there are more opportunities for adults to infect juveniles than if the site were to become free of snow and ice later in the year. In our data set, the prevalence of influenza among hatching year birds is significantly greater than in second-year or adult birds ($x^2_{1} = 64.87$, one-sided $p = 4 \times 10^{-8}$).

### Table 2: AIV prevalence among avian orders.

<table>
<thead>
<tr>
<th>AIV-positive species</th>
<th>Common name</th>
<th>Prevalence (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anseriformes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cygnus olor</td>
<td>Mute Swan</td>
<td>100</td>
</tr>
<tr>
<td>Anas rubripes</td>
<td>American Black Duck</td>
<td>50</td>
</tr>
<tr>
<td>Aythya Americana</td>
<td>Redhead</td>
<td>33.3</td>
</tr>
<tr>
<td>Anas carolinensis</td>
<td>Green-winged Teal</td>
<td>7.36</td>
</tr>
<tr>
<td>Anas platyrhynchos</td>
<td>Mallard</td>
<td>5.9</td>
</tr>
<tr>
<td>Charadriiformes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Arenaris interpres</td>
<td>Ruddy Turnstone</td>
<td>16.3</td>
</tr>
<tr>
<td>Falconiformes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Haliaeetus leuecephalus</td>
<td>Bald Eagle</td>
<td>1.23</td>
</tr>
<tr>
<td>Passeriformes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Regulus satrapa</td>
<td>Golden-crowned Kinglet</td>
<td>50</td>
</tr>
<tr>
<td>Passerella iliaca</td>
<td>Fox Sparrow</td>
<td>10</td>
</tr>
<tr>
<td>Piranga ludoviciana</td>
<td>Western Tanager</td>
<td>9.09</td>
</tr>
<tr>
<td>Seiurus noveboracensis</td>
<td>Northern Waterthrush</td>
<td>9.09</td>
</tr>
<tr>
<td>Carpodacus cassini</td>
<td>Cassin’s Finch</td>
<td>8.33</td>
</tr>
</tbody>
</table>

Two orders (Charadriiformes and Falconiformes) had only one AIV-positive species. For orders in which influenza was detected in at least five species, the top five species in terms of prevalence are listed below.
That finding is compatible with the hypothesis that a high fraction of hatchlings are infected by adults at the breeding grounds. The amount of harvested cropland per county was very highly significant for explaining AIV cases. This result is consistent with previous analyses that showed a significant effect of agricultural production on AIV cases in Southeast Asia [38,72]. We hypothesize that agricultural activity results in increased AIV prevalence because it reduces the amount of natural habitat available to avian migrants. For example, the conversion of wetlands to farms may create bottlenecks at stopover sites along migratory corridors, concentrating waterfowl into high densities, such as the congregations of Teal and Snow Geese at Kesterson National Wildlife Refuge in California’s Central Valley during migration. The resulting crowding and intermingling of different species is thought to increase the probability that a bird will be infected with influenza [22].

Minimum temperature also emerged as significant for explaining AIV cases. The sign of the coefficient that represents the effect of minimum temperature was estimated to be negative, meaning that if temperature increases, then the number of AIV cases is predicted to decrease (Table 3). This finding is compatible with the ecology of AIV, which is known to survive longer outside the host in cold conditions [2,73]. During the 1984 Pennsylvania outbreak of H5N2, the virus survived in barns for as long as 105 days during cool weather [28]. In addition, winters are long and cold at Qinghai Lake, China, which was the site of an outbreak of HPAI in wild birds in 2005 [32]. It has also been conjectured that cold snaps explain the spread of HPAI by wild birds in Europe insofar as cold weather events prompt the dispersal of infected birds, resulting in the spread of the virus [16,63,74,75]. For example, in 2006, mute swans that typically winter on the Black Sea became infected when bad weather forced them to leave. This led to the discovery of infected mute swans in Azerbaijan, Georgia, Iran, Kazakhstan, and 20 European countries [31]. Additionally, frost is one of the stress factors associated with dead wild birds infected with HPAI [16].

**Influenza predictions for the unsampled counties**

The analysis at the county scale identifies the Mississippi River basin as a hotspot for AIV cases in the contiguous US (Figure 3(a)), which may be because wetlands in the basin with shallow pools of water are conducive to the transmission of the virus [37]. The Pacific Northwest is also classified as a hotspot, though this may be due in part to the fact that many of our samples of passerines came from this region (Figure 1(b), Additional file 4, Figure S1). At the state scale, Minnesota is predicted to have the most cases of AIV in the contiguous US (Figure 4). This is not surprising since AIV is known to have been introduced to turkey farms in Minnesota 135 times since 1968 [76]. At the county scale, there is a pronounced north-to-south gradient in the predicted number of AIV cases in the contiguous US (Figure 3(a)). This finding is compatible with the theory that the virus persists better in the colder environments of the northern US than the warmer environment of southern US states. Finally, the model predicts a large number of AIV cases in the Corn Belt of the Central US (Figure 3(a)). This area is

![Figure 2 Influenza prevalence among avian orders](image)

Each bar represents one order of birds in the contiguous US. The height of the bar indicates that number of species belonging to the order that tested positive for AIV. The number above each bar is the % of cloacal samples that were AIV-positive. An = Anseriformes, Ch = Charadriiformes, Ci = Ciconiiformes, Co = Columbiformes, Cu = Cuculiformes, Fa = Falconiformes, Gr = Gruiformes, Pa = Passeriformes, Pe = Pelecaniformes, Pi = Piciformes, and St = Strigiformes. The first number below the bar is the number of species that we tested from that order. The second number is the number of samples that we tested from all of the species belonging to the order (see Additional file 2, Table S1 for details).

### Table 3: Effect of the environmental variables on AIV cases in wild birds in the contiguous US, 2005-2008.

<table>
<thead>
<tr>
<th>Effect</th>
<th>Est.</th>
<th>Std. Err.</th>
<th>t-value (df = 132)</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intercept</td>
<td>-3.279</td>
<td>0.58</td>
<td>-5.65</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>First day of the year free of ice or snow Range: 0 (snow-free) to day of the year 150</td>
<td>-0.0282</td>
<td>0.0116</td>
<td>-2.43</td>
<td>0.0164</td>
</tr>
<tr>
<td>Minimum temperature Range: -7.15 to 19.67°C</td>
<td>-0.2143</td>
<td>0.0734</td>
<td>-2.92</td>
<td>0.0041</td>
</tr>
<tr>
<td>Percent harvested cropland per county: Range: 0-100</td>
<td>0.0433</td>
<td>0.00951</td>
<td>4.56</td>
<td>&lt;.0001</td>
</tr>
</tbody>
</table>
known to be a major migratory flyway for ducks and also has intensive agricultural production [53], which is a significant risk factor for AIV according to our model. If we reformulate the regression model to estimate the probability of AIV occurrence rather than estimating the number of cases of AIV, the spatial pattern of AIV in the contiguous US remains qualitatively similar (Additional file 5, Figure S1).

**Discussion**

**Implications for influenza surveillance and control in wild birds**

Although the role of ducks and wading birds as influenza reservoirs has long been appreciated, our analysis shows that land birds constitute an important natural host of the influenza virus in the US. Analysis of 225 avian species indicates that influenza prevalence is higher in passerines than in eight other orders of birds in the contiguous US. Thus, the implication of this study for surveillance is that passerines should be monitored as a potential vector for transmitting AIV to humans, in addition to water birds and domesticated birds.

Since vaccinating against or stamping out AIV in all wild birds would be impossible, it is important to prioritize populations for such management activities [16]. Our model predicts that the risk of AIV outbreaks in wild birds is highest in California, the Great Plains, Minnesota, Texas, and Washington (Figures 3 and 4). The Plains region is predicted to have the highest number of AIV cases due to its significant agricultural production and cold winter temperatures, which allow AIV to persist outside the host in environmental reservoirs (Figure 4). Our prediction that there will be an AIV hotspot in the Pacific Northwest is driven by our samples from passerine birds because almost all of the positive samples from Passeriformes came from this region. The Passeriformes with

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**Figure 3 Predictions of the spatial model**

(a) Predictions for AIV cases in sampled and unsampled counties (n = 3109). R.m.s.e.: 6.33 cases per county; (b) Uncertainty associated with the model predictions in (a): standard error of the number of predicted cases of AIV in wild birds; (c) An important environmental variable for explaining AIV cases: % cropland per county; (d) Another important environmental variable: first day of the year with no snow or ice. Most counties with a high % of AIV positive-wild birds have a high % crop of cropland and cold climate measured as snow or ice-cover in January. This is compatible with the observation that the AI virus can survive outside of the host for a longer time period in a cold environment.
the highest AIV-prevalence was the Golden-crowned Kinglet (*Regulus satrapa*), but this result should be interpreted cautiously since we had few samples for this species (Table 2, Additional file 2, Table S1). The passerine bird that showed the highest prevalence among the species for which we had a large number of samples was Swainson’s Thrush (*Catharus ustulatus*). Swainson’s Thrush is a Neotropical migrant whose breeding sites include remote areas of the Pacific Northwest that are isolated from human settlements or farms [57]. The fact that we detected high AIV-prevalence in Swainson’s Thrush supports the hypothesis that passerines birds constitute a reservoir of AIV even without the spillover of influenza from domestic birds to wild birds at farms in the breeding range. However, further work is needed to investigate the possible exposure of Swainson’s Thrush to poultry in its winter range.

Conversely, the Mississippi Flyway in the Plains region had only one AIV-positive sample each from song birds and shorebirds, so our prediction of an AIV hotspot in this part of the country is determined primarily by influenza-positive ducks (Anseriformes). The Anseriformes species with the highest AIV prevalence was the Mute Swan (*Cygnus olor*), which had 100% prevalence in our data set (Table 1). However, this prevalence may not be representative of natural populations insofar as our sample size for this species was small. Among the well-sampled Anseriformes species, AIV-prevalence was high in the Mallard (*Anas platyrhynchos*) and the Green-winged Teal (*Anas carolinensis*) (Additional file 2, Table S1). This result is compatible with previous studies that have detected high prevalence in both of these duck species [reviewed in [72]].

In addition to estimating the mean number of AIV cases in wild birds per county, we predict the standard error of the number of cases (Figure 3(b)). If the standard error for a particular county is large, then there is a great deal of uncertainty about AIV cases in the county. To reduce this uncertainty, counties with large standard errors, such as those in central Washington and central Montana, can be targeted for increased surveillance.

**Human health implications of influenza in wild birds**

Previous spatial models have predicted the occurrence and non-occurrence of AIV [38,45]. The novel contribution of our model is the prediction of the number of cases of influenza in wild birds. Understanding the number of AIV cases in a county, rather than the occurrence or non-
occurrence of the virus, is important because the rate of con-

fection with multiple influenza viruses increases with

number of AIV cases [2]. Co-infection is the source of reassortment events that give rise to new pan-
demic strains of AIV; indeed, such an event preceded the
1918 influenza pandemic that killed 50-100 million people
[2,3,5-8,77,78]. Since the number of AIV cases is pre-
dicted to be highest in the Great Plains and the Pacific
Northwest, we predict that these two geographical
regions will also have a concomitantly higher risk of co-

fection and reassortment. Thus, the threat to human
health due to the evolution of HPAI through reassort-
ment is greatest in those two parts of the country. As a

consequence, implementing biosecurity practices for the
control of HPAI is especially crucial in those two areas
[79].

Efforts undertaken by US health departments to plan
for AIV since 2005 are thought to have facilitated the
response of federal, state, and local agencies to the novel
H1N1 epidemic during the Spring of 2009 [80]. Thus,
modelling the distribution of AIV in the US and allocat-
ing health care resources based on the predictions of such
models may contribute to an improved technological
infrastructure for responding to future epizootics and
epidemics of influenza as well as other public health cri-

ses.

Shortcomings of the analysis and areas for future research
Among the limitations of the analysis is that we did not
construct separate models for AIV in different species of
wild birds. However, this shortcoming may not be severe
because H5 AIV does not show species-specific differ-
ences in North America [16,23]. Moreover, like our anal-
ysis, epidemiological models often analyze the distribution
of the pathogen rather than that of the host species [e.g.
[81]]. Another limitation of the analysis is that our viral
storage and detection methodology does not permit the
characterization of the HA or NA subtype of a sample
that tests positive for influenza. This is a shortcoming
because it may be more important to map the distribu-
tion of subtypes that are highly virulent in mammals such
as H5 or H7. Our models are based on the detection of
influenza A through matrix gene detection rather than the
analysis of the H5 and H7 subtypes. Thus, if these two
subtypes have unique geographic distributions because
they are only transmitted by particular species of wild
birds, then our modelling approach might fail to capture
this. However, wild birds in the US exhibit a high rate of
turnover in serotypes according to a roughly 2-year cycle
[2,16,36,61,82]. Thus, modelling the geographic distribu-
tions of the H5 or H7 subtypes would require detailed
annual surveys. Although such surveys have been con-
ducted for Delaware Bay [49], such data are not available
for sites that represent broad geographic sampling of the
contiguous US.

In the Eastern hemisphere, cases of H5N1 AIV in peo-
ple typically increase in cooler months [9]. We found that
cool temperatures are also a good predictor of AIV cases
in wild birds. Influenza dynamics in wild birds in the US
appear to depend on climatic variables rather than a fixed
annual cycle because the relationship between AIV cases
and day of the year is weak (Pearson’s = 0.17). A hypothe-

sis that emerges from this study is that H5N1 may be more prevalent in humans during cool months because
both the prevalence of the virus in birds and the rate of
avian-to-human transmission are higher in cool weather.
Future field studies could assess the evidence in support
of this hypothesis by testing for influenza in birds and
people simultaneously in the same geographic region
during cool weather. If the hypothesis is not confirmed, a
possible alternative explanation for the increase in human
cases of influenza during cool weather is that people
spend more time indoors during the cool winter months
and thus have greater exposure to infected individuals [3].

The recent H1N1 pandemic has demonstrated the pub-
lic health significance of reassortment events between
avian and swine influenza viruses. Such reassortment
might be expected to be more frequent in geographic
regions where (i) swine production is intensive and (ii)
there is also high prevalence of influenza in wild birds.
Regions that score high for both (i) and (ii) could have a
greater likelihood of influenza reassortment events in
livestock or wildlife hosts of the influenza virus. For
example, results indicate that the Mississippi Flyway in
the central US has significant swine production along
with a significant number of cases of AIV in ducks (Addi-
tional file 6, Figure S1). However, the risk of an influenza
epidemic in humans may depend on other parameters,
such as contact rates among people, wild birds, poultry,
and swine, and the transmission efficiency of the virus.
The available data on the exposure of humans to AIV is
extremely limited and difficult to interpret due to a lack
of standardized methods for serological testing [reviewed
in [7]]. We consider the improvement of such data an
important avenue for future work and refer the interested
reader to Moffett et al. [83] for the incorporation of con-
tact rates and transmission efficiencies into a model of
epidemiological risk.

Conclusions
The main conclusion of this research is that land birds
(Passeriformes) constitute an important natural reservoir
of influenza in the contiguous US. The importance of this
finding is that since passerine species are common in
urban habitats they could readily transmit highly patho-
genic influenza to people in the event that a highly patho-
genic subtype evolves through mutation or through reassortment in a bird that is co-infected with distinct influenza viruses. Aquatic birds are typically referred to as the most important avian vector of influenza. However, since passerines occupy the same habitat as poultry and there have already been outbreaks of HPAI in US poultry, passerines may be more effective at transmitting HPAI to people than aquatic birds. The geographical regions with the highest risk of influenza in wild birds are the Great Plains and the Pacific Northwest; the threat to human health due to reassortment events that produce pandemic influenza subtypes is also greatest in these two areas.

Additional material

Additional file 1 Influenza samples from wild birds used in the study
This file provides a detailed description of the geographical study region and lists the online databases from which we obtained samples in addition to the samples tested at the UCLA Center for Tropical Research.

Additional file 2 Description of the samples by species. The table in this file reports the prevalence of flu in the 225 avian species analyzed in this study, which represent 11 orders of birds.

Additional file 3 Formulation of the spatial regression model. This file explains how we constructed the semivariogram in the spatial regression model, provides a mathematical formulation of the model, and explains how we fitted the model.

Additional file 4 Geographic locations of AIV-positive samples in the contiguous US (n = 325). This file contains a map showing the bird banding stations where wild birds tested positive for AIV.

Additional file 5 Probability of AIV occurrence in US counties or county equivalents. This file consists of a map constructed by modifying the spatial model to generate probabilistic predictions about the influenza in wild birds, which are restricted to being zero and one, rather than estimates of the number of influenza cases, which range from zero to large numbers of cases per county.

Additional file 6 Hotspots of swine production and AIV cases in wild birds in the contiguous US. This file shows the overlap between areas with intensive swine production in the US and areas in which we predict high prevalence of AIV in wild birds. Reassortment between avian and swine influenza viruses may be more common in such areas.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
TF designed the study’s analytical methods and drafted the manuscript. SS helped design the analytic methods, including the preparation of the satellite data described in the Methods. EC supervised the first year of laboratory activities, helping to design laboratory techniques for influenza testing, and wrote a FileMaker Pro database to summarize the samples. ET supervised the second year of laboratory activities including flu testing and the cataloguing of more than 10,000 cloacal samples. HT helped design the analytic aspects of the study and carried out a review of the literature about flu in wild birds. WB helped design the study’s analytic strategy including the selection of the type of remote-sensed images to be used in the spatial model and selected the spatial scale of these images. DD designed the study’s field activities including the sampling of passerine birds as part of the MAPS bird banding network. PN reviewed and revised a preliminary draft of the manuscript, provided guidance about the study’s analytical approach, and helped design sampling activities as part of the MAPS network. JM supervised field activities as part of the MAWS bird banding network and designed Figure 2 of the manuscript. CJR supervised field activities in the LaMNA bird banding network, which provided flu samples during the migratory period. JA helped supervised the LaMNA bird banding stations and revised a draft of the article. JP directed laboratory activities during both years of flu testing and helped draft the Methods section of the manuscript. TBS helped design the analytic aspects of the study and helped draft the manuscript. All authors read and approved the final manuscript.

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