Title
Artificial control of gene expression in mammalian cells by modulating RNA interference through aptamer-small molecule interaction

Permalink
https://escholarship.org/uc/item/7cz773vx

Journal
RNA-A Publication of the RNA Society, 12(5)

ISSN
1355-8382

Authors
An, Chung-Ii
Trinh, Vu B
Yokobayashi, Yohei

Publication Date
2006-05-01

Peer reviewed
Artificial control of gene expression in mammalian cells by modulating RNA interference through aptamer–small molecule interaction

Chung-II An, Vu B. Trinh and Yohei Yokobayashi

RNA 2006 12: 710-716; originally published online Apr 10, 2006;
Access the most recent version at doi:10.1261/rna.2299306

References
This article cites 33 articles, 11 of which can be accessed free at:
http://www.rnajournal.org/cgi/content/full/12/5/710#References

Email alerting service
Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article or click here

Notes

To subscribe to RNA go to:
http://www.rnajournal.org/subscriptions/

© 2006 RNA Society
Artificial control of gene expression in mammalian cells by modulating RNA interference through aptamer–small molecule interaction

CHUNG-IL AN, VU B. TRINH, and YOHEI YOKOBAYASHI
Department of Biomedical Engineering, University of California, Davis, California 95616, USA

ABSTRACT
Recent studies have uncovered extensive presence and functions of small noncoding RNAs in gene regulation in eukaryotes. In particular, RNA interference (RNAi) has been the subject of significant investigations for its unique role in post-transcriptional gene regulation and utility as a tool for artificial gene knockdown. Here, we describe a novel strategy for post-transcriptional gene regulation in mammalian cells in which RNAi is specifically modulated through RNA aptamer–small molecule interaction. Incorporation of an RNA aptamer for theophylline in the loop region of a short hairpin RNA (shRNA) designed to silence fluorescent reporter genes led to dose-dependent inhibition of RNAi by theophylline. shRNA cleavage experiments using recombinant Dicer demonstrated that theophylline inhibited cleavage of an aptamer-fused shRNA by Dicer in vitro. Inhibition of siRNA production by theophylline was also observed in vivo. The results presented here provide the first evidence of specific RNA–small molecule interaction affecting RNAi, and a novel strategy to regulate mammalian gene expression by small molecules without engineered proteins.

Keywords: RNA interference (RNAi); aptamer; theophylline; Dicer

INTRODUCTION
Gene expression is regulated at multiple steps, ranging from transcription initiation to post-translational modification and/or degradation. Recently, small RNA-mediated gene-silencing phenomena in eukaryotes (Zamore and Haley 2005) have been extensively investigated due to their important regulatory roles in diverse organisms. The gene-silencing mechanism involves trans-acting small RNAs such as small interfering RNAs (siRNAs) and microRNAs (miRNAs) as essential regulatory RNA elements. siRNAs are key molecules in the RNA interference (RNAi) pathway, in which long double-stranded RNAs (dsRNAs) or short hairpin RNAs (shRNAs) are processed by the RNase III enzyme Dicer, and the resulting siRNAs of ~21 nt in length function as guides for specific mRNA degradation in the RNA-induced silencing complex (RISC) (for reviews, see Meister and Tuschl 2004; Mello and Conte 2004). The discovery of RNAi added another layer of functionality of RNA molecules to the already versatile repertoire of biological functions. Furthermore, the universal nature of RNAi mechanism suggested its enormous potential as a tool to control gene expression in cultured cells and organisms. In fact, RNAi is now widely used as a simple and effective method to knock down gene expression in diverse organisms. Furthermore, therapeutic applications of siRNAs are being explored (for review, see Hannon and Rossi 2004).

In this report we describe a novel strategy for artificial gene regulation in mammalian cells using RNAi as a basal system. In this approach, RNAi is specifically modulated by theophylline thorough the binding to its aptamer. Aptamers are RNA or DNA oligonucleotides that can bind specific ligands with high affinity and specificity (Ellington and Szostak 1990; Tuerk and Gold 1990), and they have been used for artificial gene regulation in vivo (Wetstuck and Green 1998; Harvey et al. 2002; Hanson et al. 2003; Suesse et al. 2003; Buskirk et al. 2004; Desai and Gallivan 2004; Bayer and Smolke 2005). Our new approach to controlling gene expression in mammalian cells combines the versatility of siRNAs to target arbitrary genes and the
powerful in vitro selection technology to develop RNA aptamers for a variety of molecules.

RESULTS AND DISCUSSION

RNAi induction with aptamer-fused shRNAs and its inhibition by theophylline

We constructed a human U6 promoter-driven vector that transcribes an shRNA in which the in vitro-selected RNA aptamer for theophylline (Jenison et al. 1994; Zimmermann et al. 1997) is fused to the loop region of the shRNA with a 19-bp double-stranded stem targeting the enhanced green fluorescent protein (EGFP) gene (pE19T) (Fig. 1A). When this construct was cotransfected into HEK293 cells with pEGFP-N1 and pDsRed1-N1 (EGFP and DsRed expression vector, respectively), EGFP silencing was inhibited in a dose-dependent manner to the theophylline concentration (Fig. 1B). DsRed fluorescence was used to normalize for transfection efficiency. The corresponding shRNA expression vector that lacks the aptamer (pE19) (Fig. 1A) retained its RNAi activity up to 1 mM theophylline with small increases in EGFP expression at 5 and 10 mM. The cells grew slightly slower at higher concentrations of theophylline (5 and 10 mM), which may account for the minor increase in EGFP expression, possibly due to nonspecific pleiotropic effects. Addition of up to 1 mM caffeine, which is structurally similar to theophylline but does not bind to the aptamer (Jenison et al. 1994), did not affect RNAi efficiency (data not shown), and higher concentrations of caffeine proved to be toxic to the cultured cells (data not shown).

Similar RNAi inhibition by theophylline was obtained when DsRed gene was targeted instead of EGFP (Fig. 2). The results are consistent with the modular design of the aptamer-fused shRNAs comprised of a gene-targeting stem and a molecular-sensing aptamer that function independently.

Two related constructs (pE20T and pE21T) (Fig. 1A) were examined to probe the operating mechanism of the system. pE20T and pE21T transcripts contain 20 and 21 double-stranded bp in the stem region, respectively. Both constructs retained gene-silencing activity but did not appreciably respond to theophylline (Fig. 1C). The expected Dicer cleavage sites of pE20T and pE21T transcripts lie outside of the theophylline-binding site within the aptamer (Fig. 1A), consistent with their lack of response.

![FIGURE 1](image-url)

FIGURE 1. Design of aptamer-fused shRNAs and effect of theophylline on RNAi induced by the shRNAs. (A) Secondary structures of putative transcripts from pE19, pE19T, pE20T, and pE21T. Boxed nucleotides indicate the anti-sense strand targeting EGFP, gray nucleotides indicate the loop sequence from the pSilencer vector (Ambion) (Brummelkamp et al. 2002), circled nucleotides indicate bases that interact with theophylline (Zimmermann et al. 1997), and arrowheads indicate putative Dicer cleavage sites. RNA polymerase III transcripts are known to have heterogenous 3′ terminus with one to four U residues (Bogenhagen and Brown 1981; Miyagishi and Taira 2002). For simplicity, structures with two U residues are depicted. It should be noted that the pE19 transcript (E19 shRNA) was predicted to have a 21-bp stem region consisting of a 19-bp stem encoding an EGFP sequence and an additional 2-bp stem from the loop sequence recommended by the manufacturer. (B) Dose-dependent inhibition of RNAi with pE19T by theophylline. HEK293 cells were cotransfected with pEGFP-N1 and pDsRed1-N1, and pE19 (without aptamer) or pE19T (with aptamer) in a 96-well microplate. Fluorescence measurements were performed 48 h after transfection, and the ratios of EGFP and DsRed fluorescence intensity were calculated. The ratios were then normalized to those from cells transfected with an shRNA expression vector with a scrambled sequence that has no significant homology with the human genome. The data are averages of triplicate transfections and error bars represent standard deviations. (C) Effect of shRNA stem length on theophylline-dependent RNAi inhibition. HEK293 cells were cotransfected with pEGFP-N1, pDsRed1-N1, and either pE19, pE19T, pE20T, or pE21T in a 96-well microplate.
to theophylline (Fig. 1C). These results suggested that theophylline binding to the aptamer transcript interferes with Dicer cleavage due to the proximity of the binding pocket to the putative Dicer cleavage site (Fig. 1A).

**Inhibition of Dicer-mediated shRNA cleavage by theophylline in vitro**

To test the possibility of Dicer-mediated cleavage of pE19T transcript being inhibited by theophylline, we prepared the predicted RNA transcripts of pE19 and pE19T (Fig. 1A) by in vitro transcription (E19 and E19T, respectively) and reacted them with recombinant Dicer in the absence and the presence of theophylline. As we expected, dose-dependent inhibition of Dicer-mediated siRNA production was observed for E19T but not for E19 (Fig. 3), supporting the proposed mechanism. The same reaction using E20T (Fig. 1A) also resulted in dose-dependent inhibition of Dicer cleavage by theophylline, but the degree of inhibition was smaller than that of E19T. These observations indicate that the proximity of Dicer cleavage site to the theophylline-binding site is important in modulating Dicer activity.

In addition to siRNA, longer cleavage products were detected in the Dicer cleavage reactions of E19T and E20T (indicated by an asterisk in Fig. 3). Judging from their sizes (between 30 and 50 nt; data not shown) and their detectability with the EGFP sense sequence probe, they are expected to be partial cleavage products between the 21st and 22nd nucleotides from the 5’ end of the shRNAs (a 44-nt fragment from E19T and a 46-nt fragment from E20T) (Fig. 1A). This observation suggests that insertion of theophylline aptamer decreased the efficiency of simultaneous cleavage of both strands by an intramolecular heterodimer of the two RNase III domains of Dicer (Zhang et al. 2004; Macrae et al. 2006), presumably due to the relatively large size of the aptamer and its unstructured nature in the absence of theophylline (Zimmermann et al. 1997). Addition of theophylline also resulted in reduction of the longer species, which was more evident with E20T than E19T (Fig. 3), suggesting that cleavage between A21 and U22 in E20T (cf. Fig. 1A) is more susceptible to theophylline binding than that between U21 and A22 in E19T (cf. Fig. 1A). These longer products were also observed in vivo (see below).

Siolas et al. (2005) recently reported that their shRNA with a 19-bp stem and a 4-bp loop was not cleaved by recombinant Dicer in vitro but was still able to induce RNAi in vivo. In contrast, our shRNA with a 19-bp stem...
(E19T) was clearly processed by Dicer in vitro in the absence of theophylline. The difference may be due to the distinct nature of the loop regions (length, structure, etc.) used in the two studies.

**Inhibition of siRNA production by theophylline in vivo**

To investigate the effects of theophylline on siRNA accumulation in vivo, Northern analysis was performed using small RNAs isolated from HEK293 cells, which were cotransfected with pEGFP-N1, pDsRed1-N1, and one of the shRNA expression plasmids (pE19, pE19T, pE20T, or pE21T) and incubated in the presence or the absence of 10 mM theophylline. Strong bands corresponding to the unprocessed shRNAs were detected in all cell extracts regardless of theophylline addition (Fig. 4B). As evident from the bottom picture of Figure 4B, siRNA generation from E19T shRNA was almost completely inhibited by theophylline, whereas those from E20T and E21T were diminished but still detectable. No significant difference in the siRNA levels of pE19-transfected cells in response to theophylline was observed. These observations were consistent with the result of fluorescence measurement of the same samples (Fig. 4A) as well as the in vitro Dicer cleavage results (Fig. 3).

The putative partially cleaved RNA species observed in vitro were also detected in vivo (indicated by an asterisk in Fig. 4B). Additionally, RNA species of ~30 nt were also detected from the cell lysates transfected with the aptamer-fused shRNA expression vectors (indicated by an arrow in Fig. 4B). The intensity of these bands was also diminished in response to theophylline, suggesting that the cleavage site(s) responsible for these species is located within or near the theophylline-binding pocket. Although the exact nature of these by-products awaits further investigation, the similar levels of the RNA species detected in pE19T, pE20T, or pE21T-transfected cells in the presence of theophylline and the difference in observed RNAi among these cells suggest that these RNA species are not involved in the silencing of EGFP.

Taken together, theophylline clearly inhibits siRNA production from aptamer-fused shRNAs in vitro and in vivo, but the highest degree of inhibition is observed with E19T, in which one of the putative Dicer cleavage sites overlaps with the theophylline-binding site (Fig. 1A). Consequently, theophylline-induced RNAi inhibition in vivo was only observed in cells transfected with the aptamer-fused shRNA expression vectors with a 19-bp stem (pE19T and pD19T) (Figs. 1, 2).

**Comparison with other RNAi regulation methods**

Chiu et al. (2005) recently reported two ATP analogs that inhibit RNAi in mammalian cells discovered by screening of a chemical library. These molecules are likely to inhibit a cellular factor involved in the early RNAi pathway to induce general RNAi inhibition, whereas our system was designed to effect specific control of RNAi by a small molecule directly interacting with an shRNA. To our knowledge, our system is the first demonstration of sequence-specific RNAi inhibition by a small molecule at a defined mechanistic step.

Anti-miRNA oligonucleotides (AMOs) have been used to inhibit specific miRNAs and siRNAs in vivo (for review, see Weiler et al. 2005). AMOs are easy to design and can inhibit both endogenous miRNAs and synthetic siRNAs. However, the exact process in which AMOs exert their function in the RNAi pathway is not clear. Delivery and removal of AMOs are also more complicated than many small molecule drugs, which may be more advantageous for experiments requiring temporal control.

The most popular technique for achieving temporal control of RNAi in cultured cells is probably the use of inducible shRNA vectors based on the robust tetracycline or ec dysone-inducible systems using engineered transcription
factors and promoters, or Cre-loxP recombination systems (for review, see Sandy et al. 2005). While these inducible systems offer various features needed for many applications, the aptamer-fused shRNA regulatory strategy operating at the Dicer cleavage step may be used in combination with the existing inducible shRNA expression systems to achieve more complex or improved regulation of RNAi in engineered cells. The ability to control RNAi at the Dicer cleavage step may result in a faster response compared with transcriptional control. Moreover, the modularity of the aptamer and the gene-targeting stem opens the possibility to regulate gene expression in response to endogenous metabolites or proteins for which RNA aptamers have been reported. Werstuck and Green (1998) inserted an aptamer-fused shRNA expression cassette, which will be selected in vitro. Reversibility of the RNA silencing effect will need to be demonstrated using stable transfectants of the aptamer-fused shRNA expression cassette, which will be the subject of future investigation.

pE19T and pD19T transcripts can be classified as RNA genetic switches that allow regulation of exogenous genes without exogenous protein factors. To our knowledge, only two such genetic switches that function in mammalian cells have been reported. Werstuck and Green (1998) inserted a riboswitch in the 5′-untranslated region (UTR) of a reporter transcript whose translation was negatively regulated in the presence of the cognate ligand. Yen et al. (2004) recently reported a system in which a ribosome-integrated 5′ UTR of a reporter mRNA whose self-cleavage was inhibited by small molecule ribozyme inhibitors.

Although not directly demonstrated in this work, the modularity of our system provides a unique potential of regulating endogenous genes not possible with the previously described systems (Werstuck and Green 1998; Yen et al. 2004). RNA genetic switches have some advantages over those based on engineered protein transcription factors such as reduced risk of immunogenic complications and smaller genetic sizes that may be beneficial for in vivo applications. In light of the recent progress in natural and artificial RNA-based gene regulatory mechanisms in bacteria (Isaacs et al. 2004; Davidson and Ellington 2005) and yeast (Suess et al. 2003; Buskirk et al. 2004; Bayer and Smolke 2005), small molecule-regulated RNA genetic switches in mammalian cells should further advance our understanding of the roles of RNA in gene regulation as well as their applications.

MATERIALS AND METHODS

Plasmid construction

pEGFP-N1 and pDsRed1-N1 were obtained from Clontech. pSilencer 2.1-U6 hygro (Ambion) was used for constructing all RNAi vectors. pE19 (E19 stands for 19 bp of shRNA encoding an EGFP target sequence [Meister et al. 2004]), pE19T (T stands for theophylline aptamer inserted in the loop region of shRNA), pD19 (D stands for DsRed), and pD19T were produced by ligating phosphorylated and annealed oligonucleotides with pSilencer 2.1-U6 hygro. Oligonucleotides used were as follows (a pair of annealed oligonucleotides was indicated by “&”):

- pE19: 5′-GATCCGGCGAAGCTTACCTGAGA &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT &5′-CTTTTGCTCTTTGGGCCAAG &5′-AGCTTTCCAAAGAGGAGGAGAAGCTTTGCTAGAGT &5′-TCGAGCTTTGCTAGAGT...

...
Fluorescence measurements

To quantitatively determine in vivo RNAi activity, dual fluorescence reporter systems using EGFP and DsRed (RFP) have been developed and used by many researchers (Chiu and Rana 2002, 2003; Chiu et al. 2004, 2005; Shah et al. 2005; Yang et al. 2005). Our assay procedure was established according to the previous reports with some modifications. EGFP and DsRed fluorescence were detected with Safire® microplate reader (Tecan) according to the manufacturer’s instructions. Before measuring fluorescence intensity, medium was removed from each well, and 150 μL of prewarmed (37°C) phosphate-buffered saline (PBS) was added and incubated at 37°C for 10 min in the instrument. Measurement conditions were as follows: measurement mode, fluorescence bottom well reading; excitation wavelength, 484 nm (for EGFP) or 558 nm (for DsRed); emission wavelength, 510 nm (for EGFP) or 583 nm (for DsRed); excitation bandwidth, 5 nm; emission bandwidth, 5 nm; gain, optimal; number of reads, five; temperature, 37°C. Data were processed as follows: First, fluorescence values of EGFP were divided by those of DsRed to obtain EGFP/DsRed (DsRed fluorescence was used to normalize for transfection efficiency), and the average values and standard deviations of DsRed (DsRed fluorescence was used to normalize for transfection efficiency), and the average values and standard deviations were normalized to control, which expresses an shRNA with the following sequence: 5′-TTCTGCCAAGGGCCTTTCGGCTGGTATAAC (underline indicates T7 promoter sequence). Oligonucleotides used to generate an E19 template were 5′-TCAAGAGAAC &5; double-stranded stem underlined) too b-}

In vitro processing of shRNAs by recombinant Dicer

E19, E19T, and E20T shRNAs were prepared by in vitro transcription using AmpliScribe T7 High Yield Transcription Kit (Epicentre). Template DNAs were prepared by annealing and T4 DNA polymerase-mediated extension of synthetic oligonucleotides. Oligonucleotides used to generate an E19 template were 5′-GCGTTAATACGACTCACTATAGGGCAAGCTGACCCCTGAAGTTCAAGAGAAC &5; double-stranded stem underlined) too b-}

Isolation of small RNAs from transfected cells

A total of 150 ng of pEGFP-N1, 300 ng of pDsRed1-N1, and 1.5 μg of an RNAi vector were cotransfected into HEK293 cells using 20 μL of PolyFect reagent (QIAGEN) in 6-well plates. Cells were incubated in a 5% CO2-humidified incubator at 37°C in the HEK293 medium supplemented with 0 or 10 mM theophylline for 47 h. After measurement of fluorescence intensity as described above, small RNAs were isolated using mirVana miRNA Isolation Kit (Ambion) according to the manufacturer’s instructions.

Northern blotting

A 0.5-μL aliquot of in vitro reaction products (0.6 pmol RNA; see above) or 1 μg of small RNAs from transfected cells (see above) was separated on a 15% polyacrylamide/7 M urea denaturing gel using NorthernMax Formaldehyde Load Dye (Ambion) and electroblotted to BrightStar-Plus Positively Charged Nylon Membrane (Ambion). After prehybridization in ULTRAhyb-Oligo Hybridization Buffer (Ambion) at 25°C for 30 min, blots were incubated with a 5′-biotin-labeled DNA oligonucleotide probe (100 ng/mL) for EGFP (5′-AAAAAGGCAAGCTGACCCCTGAAGTTCAAGAGAAC &5; double-stranded stem underlined) too b-}

REFERENCES


