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High-protein diet selectively reduces fat mass and improves glucose tolerance in Western-type diet-induced obese rats

Andreas Stengel,1,2 Miriam Goebel-Stengel,1,3 Lixin Wang,1 Eugenia Hu,1 Hiroshi Karasawa,1 Joseph R. Pisegna,1 and Yvette Taché1

1CURE/Digestive Diseases Research Center, Center for Neurobiology of Stress, Department of Medicine, Digestive Diseases Division at the University of California Los Angeles, VA Greater Los Angeles Health Care System, Los Angeles, California; 2Charité Center for Internal Medicine and Dermatology, Division Psychosomatic Medicine, Obesity Center Berlin; Charité Universitätmedizin Berlin, Berlin, Germany; and 3Department of Internal Medicine, Institute of Neurogastroenterology and Motility; Martin-Luther Hospital, Berlin, Germany

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Obesity has become epidemic, and by the year 2030, half of the adult population is predicted to be obese in the United States (54). Its prevalence is expanding worldwide in industrialized countries and spreading also to less developed countries; therefore "globesity" is taking over many parts of the world (see World Health Organization homepage). The plethora of health consequences related to obesity involves increased risk of developing adult-onset Type 2 diabetes mellitus, dyslipidemia, arteriosclerosis, and certain forms of cancer, including colorectal and pancreatic cancer (25). Therefore, obesity represents a major medical problem in the United States, and 12% of the entire health care costs are spent taking care of obesity-related comorbidities (52). A modest 5 to 10% body weight loss in obese subjects has been shown to result in improvements of these metabolic disturbances (21, 53).

Although two antiobesity drugs have recently been approved, drug treatment options are still very limited (26), and therefore, dietary interventions remain popular. In particular, the use of a high-protein diet is considered a beneficial strategy to achieve a long-term reduction in body weight (8, 22, 47, 48). However, the means by which an isocaloric high-protein diet reduces body weight are not well understood. There is some evidence that protein decreases appetite by increasing anorexigenic signaling (16, 34, 55) and stimulates thermogenesis (13, 57, 58) in experimental animals and humans without inducing conditioned food aversion (5). The blunting of the reduction in resting energy expenditure associated with a decrease in body weight (20, 59) is another possible mechanism through which protein may facilitate long-term weight loss. It was shown that during energy restriction in human overweight and obese subjects with BMI 27–34 kg/m², the 24-h energy expenditure was reduced following two low-protein diets (15% energy as protein, one high in carbohydrate, and the other one high in protein, one high in carbohydrate, and the other one high in fat), an effect that was reduced by a high-protein diet (36% energy as protein) (59). However, the subsequent alterations of body composition exerted by a high-protein diet under conditions of obesity are not well characterized in rodents.

In the present study, we assessed changes in food intake, body weight, and composition in rats developing obesity in response to a Western-type diet (45% calories from fat, 20% from protein, and 35% from carbohydrates, including 17% from sucrose) that were switched to an isocaloric high-protein (9% calories from fat, 52% from protein, and 39% from carbohydrates) or standard diet (9% calories from fat, 15% from protein, and 76% from carbohydrates) for 14 days. In addition, because gut-derived hormones play an important role in the control of food intake and body weight (23), we also...
investigated whether these different dietary conditions alter postprandial plasma levels of several anorexigenic hormones, namely amylin, gastric inhibitory polypeptide (GIP), insulin, leptin, pancreatic polypeptide (PP), and peptide YY (PYY) in response to a 2-h dark-phase feeding period. Because a modest reduction of body weight was shown to exert beneficial actions on the metabolic comorbidities (21, 53), we also investigated the effects of high-protein diet on glycemic control using oral glucose tolerance and insulin sensitivity tests in diet-induced obese rats.

MATERIALS AND METHODS

Animals and diets. Adult male Sprague-Dawley rats (Harlan, San Diego, CA) weighing 230–250 g were housed in pairs under controlled illumination (0600–1800) and temperature (21–23°C) during all experiments unless otherwise specified. Protocols were approved by the Institutional Animal Care and Use Committee of the Veterans Administration (no. 11047–09). Except otherwise stated, all experiments were started between 0900 and 1000 and during nonexperimental days, the maintenance was performed during that time. Animals had free access to water and different types of diet (Table 1): a standard diet (D10012M, calories: protein 15%, fat 9%, and carbohydrates 76%, 3.8 kcal/g), Western-type diet (D12451, calories: protein 20%, fat 45%, and carbohydrates 35%, 17% of calories from carbohydrates were derived from sucrose, 4.7 kcal/g) and a high-protein diet (D10011304, calories: protein 52%, fat 9%, and carbohydrates 39%, 3.8 kcal/g) purchased from Research Diets (New Brunswick, NJ). The main fat source of the Western-type diet was lard, shown to induce the most pronounced obesity compared with other fat sources, such as coconut fat and fish oil in rats (9, 24). Diets were stored at 4°C until use.

Measurements

Body composition. Body composition was assessed in conscious lightly restrained (~1 min) rats using a quantitative nuclear magnetic resonance analysis apparatus (EchoMRI-700 Composition Analyzer; Echo Medical Systems, Houston, TX), as described before (49). Changes in fat mass, lean mass, and total water were determined by assessing body composition before and 2 wk after switching to standard or high-protein diets.

Glucose tolerance. Rats were food deprived overnight with free access to water and received an orogastric gavage of 2 g·kg⁻¹·h⁻¹ glucose (Sigma-Aldrich, St. Louis, MO) in double-distilled H₂O, as described before (27). Blood was obtained by tail prick before and at 15, 30, 60, 120, 180, and 240 min after injection, and blood glucose was assessed using standard glucose test strips (One-Touch Ultra; LifeScan, Milpitas, CA). The total area under the curve (24) was calculated using the formula: blood glucose at [0 min + 15 min] + (15 min + 30 min) + (30 min + 60 min) + (60 min + 120 min) + (120 min – 60 min)]/2.

Insulin sensitivity. Rats were fasted overnight and received an intraperitoneal injection of 1 IU·kg⁻¹·ml⁻¹ insulin (Humulin, Eli Lilly, Indianapolis, IN), as detailed before (11). Blood was obtained by tail prick before and at 15, 30, 60, 120, and 240 min after injection, and blood glucose was assessed using standard glucose test strips (One-Touch Ultra). The area under the curve was calculated as described above.

Plasma hormone panel. Blood (1 ml) was obtained by decapitation and collected in ice-cooled tubes containing EDTA (7.5%, 20 µl/1 ml blood; Sigma-Aldrich) and aprotinin (1, 2 trypsin inhibitory unit/1 ml blood; ICN Pharmaceuticals, Costa Mesa, CA) and centrifuged at 3,000 rpm for 10 min at 4°C. The supernatant was separated and plasma was stored at −80°C until further processing. The measurement of the gut peptide hormones, amylin, GIP, insulin, and PYY, as well as the adipocyte-derived protein hormone leptin was performed using the Luminex xMAP technology for rat gut hormones (RGT-88K, Millipore, Billerica, MA) as used before (50). This panel allows simultaneous determination of multiple hormones without cross-reactivity between the antianalyte antibodies (manufacturer’s information; Millipore). The assay accuracy for amylin, GIP, insulin, leptin, PYY, and PP is 92, 90, 93, 87, 88, and 86%, respectively (manufacturer’s information; Millipore). All samples were processed in one batch and read via the Luminex 100 (Luminex, Austin, TX); the intra-assay variability was <10%. The multiplex assay technology has been validated against individual immunoassays for various biomarkers in several previous studies (36, 40, 45).

Table 1. Composition of standard diet, Western-type diet, and high-protein diet

<table>
<thead>
<tr>
<th>Ingredient</th>
<th>Standard Diet</th>
<th></th>
<th>Western-Type Diet</th>
<th>g kcal</th>
<th></th>
<th>High-Protein Diet</th>
<th>g kcal</th>
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<td>1982.8</td>
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<td>339</td>
<td>310.5</td>
<td>1242</td>
<td></td>
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<tr>
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<td>500</td>
<td>116.5</td>
<td>466</td>
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<td>0</td>
<td></td>
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<tr>
<td>Sucrose</td>
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<td>400</td>
<td>201.3</td>
<td>805</td>
<td>50</td>
<td>200</td>
<td></td>
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<tr>
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<td>58.3</td>
<td>0</td>
<td>50</td>
<td>0</td>
<td></td>
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<tr>
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<td>291.3</td>
<td>262.8</td>
<td>40</td>
<td>360</td>
<td></td>
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<tr>
<td>Lard</td>
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<td>1862</td>
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<td>0</td>
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<td>0.008</td>
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<td>11.7</td>
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<tr>
<td>Vitamin mix</td>
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<td>11.7</td>
<td>47</td>
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<td>0</td>
<td>2.5</td>
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<td>4727</td>
<td>999.9</td>
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<td></td>
</tr>
</tbody>
</table>

The composition of diets is given according to the manufacturer’s information (Research Diets).
of this period, food intake was determined, and rats (diet) was provided, and rats were left undisturbed for 2 h. At the end phase. Afterward, the remaining rats of the four groups were deprived diet/high-protein diet fed ad libitum) were euthanized by decapitation. Western-type diet/standard diet, and Western-type diet/high-protein diet, resulting in four experimental groups: standard diet/standard diet, standard diet/high-protein diet, Western-type diet/standard diet, and Western-type diet/high-protein diet fed ad libitum). Two-way ANOVA showed a significant influence of diet ($F_{1,342} = 268.0, P < 0.001$), time ($F_{8,342} = 26.2, P < 0.001$), and an interaction of diet and time ($F_{8,342} = 10.1, P < 0.001$). When adjusted for body weight, the food intake (expressed as kcal/300 g body wt/wk) did not differ between the dietary groups after week 2 (Fig. 1B).

Rats (n = 20) fed a standard diet for 9 wk increased their body weight by 196.2 ± 2.9 g. In the Western-type diet group (n = 20), the body weight increase was significantly higher at all weekly measurement time points reaching a 25% higher body weight gain at 9 wk compared with standard diet (244.9 ± 3.1 g body wt change, $P < 0.001$; Fig. 2). Two-way ANOVA indicated a significant influence of diet ($F_{1,342} = 524.0; P < 0.001$), time ($F_{8,342} = 1,045.5; P < 0.001$), and an interaction of diet and time ($F_{8,342} = 13.9; P < 0.001$).

High-protein diet for 14 days reduces body weight and fat mass in Western-type diet-induced obese rats. Rats fed a standard diet and remaining on this diet showed a stable food intake during the first day of the new measurement period (59.1 ± 1.8 kcal/day, Fig. 3A). The same was observed for rats fed a Western-type

**RESULTS**

Western-Type Diet Induces Obesity

Rats fed a Western-type diet consumed significantly more calories during the first week of Western-type diet feeding compared with rats kept on a standard diet (+18%, 602.5 ± 5.0 vs. 509.5 ± 7.1 kcal/wk, $P < 0.001$; Fig. 1A). In the second week, there was a sharp decrease to reach similar food consumption as that of standard diet-fed rats. Thereafter, the food intake of Western-type diet-fed rats was increased during the remaining 7 wk (range: 6 to 10%), which was statistically significant compared with standard diet from weeks 3 to 9 (e.g., week 9: 514.4 ± 6.1 vs. 470.2 ± 3.3 kcal/wk, $P < 0.001$; Fig. 1A). Two-way ANOVA showed a significant influence of diet ($F_{1,342} = 268.0, P < 0.001$), time ($F_{8,342} = 26.2, P < 0.001$), and an interaction of diet and time ($F_{8,342} = 10.1, P < 0.001$). When adjusted for body weight, the food intake (expressed as kcal/300 g body wt/wk) did not differ between the dietary groups after week 2 (Fig. 1B).

**Assessment of glucose tolerance and insulin sensitivity.**

On day 18 after switching diets to standard or high-protein diet, the four groups of rats were fasted overnight and underwent an orogastric gavage with glucose for the glucose tolerance test. Then, rats had a recovery period of 1 wk, during which they were maintained on the same diets (either standard or high-protein diet). Then (on day 25 after switching diets), the four groups of rats were fasted overnight and injected intraperitoneally with insulin to perform the insulin sensitivity test.

**Measurement of hormonal changes between light and dark phase.**

On day 32 after switching diets to standard or high-protein diet, the four groups of rats (n = 5/group; standard diet/standard diet, standard diet/high-protein diet, Western-type diet/standard diet, and Western-type diet/high-protein diet fed ad libitum) were euthanized by decapitation to obtain trunk blood between 3 and 4 PM during the light phase. Afterward, the remaining rats of the four groups were deprived of food for 2 h but had free access to water. At the beginning of the dark phase at 6 PM, preweighed rat chow (standard or high-protein diet) was provided, and rats were left undisturbed for 2 h. At the end of this period, food intake was determined, and rats (n = 5/group, 4 groups) were euthanized by decapitation to obtain trunk blood, and plasma levels of amylin, GIP, insulin, leptin, PP, and PYY were assessed by the rat Luminox panel assay.

**Statistical analysis.** Data are expressed as means ± SE and analyzed by one-way ANOVA followed by Tukey post hoc test or two-way ANOVA followed by Holm-Sidak method. Correlations were determined by univariate linear regression. $P < 0.05$ was considered significant.

Fig. 1. Food intake under conditions of Western-type diet feeding. Rats were fed either a standard or a Western-type diet (those selected as DIO responders) for 9 wk, and food intake was monitored daily and expressed as intake in kilocalories per week (A) or adjusted for body weight in kilocalories per 300 g body wt per week (B). Data are expressed as means ± SE of 20 rats/group. **P < 0.01, and ***P < 0.001 vs. standard diet.

Fig. 2. Body weight gain under conditions of Western-type diet feeding. Rats were fed either a standard or a Western-type diet (those selected as DIO responders) for 9 wk, and body weight was monitored daily and expressed as cumulative weekly body weight change. Data are expressed as means ± SE of 20 rats/group. ***P < 0.001 vs. standard diet.
diet and switched to the standard diet (58.9 ± 1.9 kcal/day, Fig. 3A). In animals that received the standard diet before and were then switched to high-protein diet, the food intake was significantly decreased during the first day (34.8 ± 2.4 kcal/day), which was also observed in the rats switched from a Western-type diet to the high-protein diet (41.2 ± 2.8 kcal/day; P < 0.001; Fig. 3A). During the remaining observation period (days 2–14), rats in the standard diet/standard diet group displayed an overall stable food intake with an average of 67.6 ± 1.0 kcal/day (Fig. 3A). Animals fed Western-type diet before and then switched to high-protein diet displayed a markedly reduced food intake during days 2–5 of the measurement period (44.2 ± 1.2 kcal/day, −24% compared with standard diet/high-protein diet, P < 0.05) and then linearly increased until day 10 when it reached the level of the standard diet/high-protein diet group (Fig. 3A). Two-way ANOVA showed a significant influence of diet (F3,462 = 169.9, P < 0.001), time (F13,462 = 19.6, P < 0.001), and an interaction of diet and time (F39,462 = 4.9, P < 0.001). The same results were observed when food intake was adjusted for body weight (Fig. 3B).

Similar to food intake, body weight remained stable in rats fed a standard diet before and kept on standard diet for 2 wk (Fig. 4). In rats fed Western-type diet before and switched to standard diet, a steady body weight decrease was observed reaching 8.9 ± 2.5 g after 14 days (P < 0.01 compared with standard diet/standard diet; Fig. 4). Rats fed standard diet before and switched to high-protein diet displayed the most pronounced body weight change during the first day and then remained body weight stable over the observation period with a slight increase during the last 2 days, resulting in a nonsignificant difference compared with the standard diet/standard diet group (body weight decrease of 3.5 ± 2.7 g after 14 days; Fig. 4). Lastly, rats fed Western-type diet before and then switched to high-protein diet displayed a steady body weight decrease over the first 9 days reaching a nadir body weight loss of 20.0 ± 1.0 g on day 9 and then showed a slight increase until the end of the 14-day measurement period but remained significantly lower than the standard diet/standard diet and the standard diet/high-protein diet group (P < 0.05; Fig. 4). Two-way ANOVA showed a significant influence of diet (F3,429 = 101.3; P < 0.001), time (F12,429 = 2.7; P < 0.01), and an interaction of diet and time (F36,429 = 1.7; P < 0.05).

Fig. 3. Food intake after switching to standard diet or high-protein diet. After development of diet-induced obesity on a Western-type diet or feeding of standard diet, rats were fed a standard or high-protein diet resulting in four experimental groups: standard diet/standard diet, standard diet/high-protein diet, Western-type diet/standard diet, and Western-type diet/high-protein diet. Food intake was assessed daily for a period of 14 days and expressed as intake in kilocalories per day (A) or adjusted for body weight in kilocalories per 300 g body wt per day (B). Data are expressed as means ± SE of 8–10 rats/group. *P < 0.05, **P < 0.01 and ***P < 0.001 vs. standard diet/standard diet. +P < 0.05, ++P < 0.01 and +++P < 0.001 vs. Western-type diet/standard diet. #P < 0.05, ##P < 0.01, and ###P < 0.001 vs. Western-type diet/high-protein diet.

Fig. 4. Body weight change after switching to standard diet or high-protein diet. After development of diet-induced obesity on a Western-type diet or feeding of standard diet, rats were fed a standard or high-protein diet resulting in four experimental groups: standard diet/standard diet, standard diet/high-protein diet, Western-type diet/standard diet, and Western-type diet/high-protein diet. Body weight was assessed daily for a period of 14 days and expressed as daily body weight change. Data are expressed as means ± SE of 8–10 rats/group. *P < 0.05, **P < 0.01 and ***P < 0.001 vs. standard diet/standard diet; +P < 0.05 and ++P < 0.01 vs. standard diet/standard diet; #P < 0.05, ##P < 0.01 and ###P < 0.001 vs. Western-type diet/standard diet.
Before switching diets, the assessment of the body composition by automated magnetic resonance-based analysis showed that rats fed a Western-type diet displayed higher body weight (Fig. 5A) and fat mass (Fig. 5C) compared with the standard diet-fed groups (P < 0.01). Also, the ratio of lean-to-fat mass was different between the standard and Western-type diet groups with a higher ratio in the standard diet/to be assigned to standard diet (6.0 ± 0.6) and standard diet/to be assigned high-protein diet group (7.3 ± 0.6) compared with the Western-type diet/to be assigned standard diet (4.3 ± 0.4) and Western-type diet/to be assigned high-protein diet group (4.0 ± 0.3, P < 0.05).

After switching diets, on day 15, the most pronounced reduction of body weight was observed in rats that were fed Western-type diet first and then switched to high-protein diet for 14 days (P < 0.05; Fig. 5B). When body composition was reassessed at day 15, fat mass was decreased (−15.4 ± 2.0 g) in the Western-type diet/high-protein diet group compared with −7.1 ± 3.6 g in the standard diet/high-protein diet group, and by −9.0 ± 2.3 g in the Western-type diet/standard diet group compared with an increase (+4.7 ± 1.2 g) in the standard diet/standard diet group (P < 0.05; Fig. 5D). The observed increase in lean mass (average +2.2 ± 0.4 g) did not differ between the four different dietary groups (P > 0.05; Fig. 5E). The ratio of lean to fat mass did not significantly change after the 14-day diet period, with a higher ratio remaining in the standard diet/standard diet (5.7 ± 0.6) and standard diet/high-protein diet group (8.3 ± 0.6) and a lower ratio remained in the Western-type diet/standard diet (4.8 ± 0.4) and Western-type diet/high-protein diet group (4.8 ± 0.5). Lastly, the total water content was decreased in rats fed Western-type diet and switched to high-protein diet (−1.4 ± 1.9 g), while it was unaltered in the Western-type diet/standard diet group (+0.2 ± 0.9 g) and increased in the standard diet/high-protein diet group (+2.5 ± 1.2 g) and in rats constantly fed with standard diet (standard diet/standard diet: +3.8 ± 0.8 g, P < 0.05; Fig. 5F).

High-protein diet improves glucose tolerance, whereas insulin sensitivity is not altered in diet-induced obese rats. In the oral glucose tolerance test performed 18 days after switching diets, no differences were observed in the fasting blood glucose
levels among the four dietary groups \((P > 0.05; \text{Fig. 6A})\). In the Western-type diet/standard diet group, rats displayed the largest increase of blood glucose reaching \(198.9 \pm 7.8\ \text{mg/dl}\) at 15 min postgavage. Blood glucose levels were 12.5 and 17.3% lower in the Western-type diet/high-protein diet group at 15 min and 30 min, respectively (Fig. 4A). Afterward, no significant differences were observed (Fig. 6A). When calculated as area under the curve, two-way ANOVA indicated a significant influence of diet \(F_{3,64} = 7.7, \ P < 0.001\) and time \(F_{3,64} = 1.529.1, \ P < 0.001\).

In the insulin sensitivity test performed 25 days after switching diets, all four groups of rats showed a marked reduction of blood glucose with a nadir during the 60–120-min period after intraperitoneal injection of insulin followed by a linear increase thereafter (Fig. 6B). No differences were observed between the four diet groups when calculated either as individual concentrations \(P > 0.05; \text{Fig. 6B}\) or area under the curve (data not shown).

**Circulating anorexigenic gut hormone response to a 2-h dark phase feeding.** Food intake of standard or high-protein diet during the first 2 h of the dark phase did not differ between the four groups (average: \(19.3 \pm 0.6\ \text{kcal/2 h}\)). In the light phase, plasma levels of GIP and insulin were significantly decreased in Western-type diet groups either switched to standard or high-protein diet compared with the standard diet/standard diet group, while the levels of the other gut hormones did not differ among groups during the light phase (Fig. 7). In the standard diet/standard diet group, plasma levels of anorexigenic hormones assessed near the end of the light phase compared with the first 2 h of the dark-phase food intake showed a significant increase of amylin and GIP (Fig. 7, A and B, Table 2). A similar response was maintained for amylin, GIP, and insulin in the other groups of rats switched to the high-protein diet (Fig. 7, A–C, Table 2). The increase of amylin was significantly lower in the Western-type diet/high-protein diet group compared with Western-type diet/standard diet rats \((P < 0.05)\), whereas the increase of GIP was higher \((P < 0.05; \text{Table 2})\). While most gut hormones showed a postprandial increase, leptin levels decreased compared with the light phase (Fig. 7D, Table 2). Although no significant differences were observed for leptin between groups, leptin plasma levels showed a positive correlation with fat mass \((r = 0.75, \ P = 0.001)\). Rats fed Western-type diet initially and then switched to high-protein diet for 32 days displayed a higher dark-phase food intake-related increase of PP (Fig. 7E) and a smaller increase of PYY (Fig. 7F) compared with the standard diet/high-protein diet group \((P < 0.05; \text{Table 2})\).

**DISCUSSION**

In this study evaluating the effects of a high-protein diet (52% protein containing 50% casein) for 2 wk on weight loss in rats fed either Western-type diet or a standard diet, we demonstrated that the high-protein diet induces a greater decrease in food intake and loss in body weight and fat mass in DIO than in lean rats with retention of body lean mass. This is associated with improved glucose tolerance in diet-induced obese rats only, while insulin sensitivity was not different between groups. Lastly, the high-protein diet fed to diet-induced obese rats induced a higher increase in the postprandial satiety hormone response to a 2-h dark phase feeding compared with the standard diet group.

Many studies have characterized the feeding-suppressive effect of a high-protein diet in rats starting the first day with a gradual adaptation during the following days (4, 6, 28, 32). However, less is known on the efficacy of a high-protein diet to curtail spontaneous energy intake in diet-induced obese rats, as studies were set mainly in the context of an additional 40% energy restriction (1, 12, 18), making it difficult to analyze the respective contributions of these interventions. In the current study, the shifting from a 20% to a 52% protein diet markedly reduced the daily energy intake on the first day by 41% and 31% in the standard diet and DIO group, respectively. On day 2, while the standard diet/high-protein diet rats resumed food consumption with a gradual return of caloric intake similar to
the standard diet/standard diet group on day 12, the daily energy intake of the Western-type diet/high-protein diet group remained low for the first 5 days and then increased gradually to reach a plateau on days 10–14, with 5% lower daily energy intake compared with that observed in the standard diet/high-protein diet group. The enhanced magnitude and duration of food intake decrease in the Western-type diet/high-protein diet group may be indicative that diet-induced obese rats display an

Table 2. Fold-changes in intestinal hormones after 2-h dark-phase feeding compared to the preprandial light phase

<table>
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<tr>
<th>Hormone</th>
<th>Standard Diet/Standard Diet</th>
<th>Standard Diet/High-Protein Diet</th>
<th>Western-Type Diet/Standard Diet</th>
<th>Western-Type Diet/High-Protein Diet</th>
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<tr>
<td>Amylin</td>
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<td>3.0 ± 0.4</td>
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<td>16.3 ± 1.0*</td>
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</tbody>
</table>

*After development of diet-induced obesity on a Western-type diet or feeding of a standard diet, rats were fed a standard or high-protein diet for 32 days, resulting in four experimental groups: standard diet/standard diet, standard diet/high-protein diet, Western-type diet/standard diet, and Western-type diet/high-protein diet. The four groups fed ad libitum with standard or high-protein diet, respectively, were euthanized by decapitation to obtain trunk blood between 3 and 4 p.m. during the light phase ($n = 5$/group). Afterward, in the remaining rats, food was removed for 2 h. At the beginning of the dark phase at 6 p.m., preweighed rat chow (standard or high-protein diet, respectively) was provided, and rats were left undisturbed for 2 h. After this period, food intake was assessed and rats ($n = 5$/group) were euthanized to obtain trunk blood, and plasma levels of hormones were assessed. *Data are fold changes of plasma levels from light to dark phase and expressed as means ± SE. Significant differences are indicated for the change in plasma levels from light to dark phase. GIP, gastric inhibitory polypeptide; PP, pancreatic polypeptide. *P < 0.05 vs. standard diet/standard diet; †P < 0.05 vs. standard diet/high-protein diet; and #P < 0.05 vs. Western-type diet/standard diet.
increased responsiveness to the food intake-reducing effect of the high-protein diet. Convergent functional and anatomical evidence in rats established that the high-protein diet effect is due to an increased satiety signaling rather than a low palatability of the diet or the induction of the conditioned taste aversion (20). Although an earlier study points to an initial orosensory preabsorptive poor palatability (37), subsequent results obtained using two choices and flavor testing, behavioral satiety sequence, taste reactivity in response to different percentages or sources of proteins support that the determinant of reduced daily energy intake in rats eating a high-protein diet is a protein-specific food intake-suppressive mechanism (4, 5, 19, 30, 31). Of relevance are also choice study experiments showing that rats select a casein-containing diet (40% protein) over other proteins (46), further supporting the enhanced acceptability of casein used in the present study (50% protein from casein, 52% protein in total).

The higher energy intake in rats fed a Western-type diet resulted in a 25% higher body weight increase compared with rats fed a standard diet over a period of 9 wk in agreement with other studies (35). This model is relevant in the context of common obesity, as it involves genetically unaltered, wild-type rats that become obese when exposed to a diet containing 32–45% fat, a proportion similar to that contained in the typical Western diet (56). Switching rats to the high-protein diet induced a marked reduction in body weight in the Western-type diet/high-protein diet group, reaching 14 g at day 14, the highest compared with all other diet groups (standard diet/standard diet, standard diet/high-protein diet, Western-type diet/standard diet). The reduction of body weight in rats that were fed the Western-type diet shifted to high-protein diet was accompanied by the reduction of fat mass (~15.4 g), likely accounting for the observed body weight loss. Likewise, the lower body weight of healthy rats fed a high-protein diet was mainly associated with a reduced body fat mass, in agreement with several previous studies during short (8 wk) (6) or long (6 mo) observation periods (33). In the present study, rats fed with a standard diet presented a higher lean-to-fat mass ratio (~6–7) compared with diet-induced obese rats fed a Western-type diet (~4). Despite the significant loss of fat mass, this ratio did not change significantly after the 14-day dietary period. This could be due to the duration of the period and may require a longer observation time. Of significance, the lean body mass is preserved as shown by the 2-g gain of lean mass, which was not different among the four groups. This is unlikely due to an increase in physical activity as a previous study in C57BL/6J mice indicated that nonexercise physical activity was not different between mice fed Western-type diet and mice fed high-protein diet for 3 mo (29). Similarly, in the same study, the respiratory quotient and energy expenditure did not differ between the Western-type and high-protein diet-fed groups (29). Consistent with our observation, other studies using dairy product diet or its components over 8 wk in diet-induced obese rats showed a similar improvement in body composition (10, 17).

After the moderate weight loss of 15 g and continued feeding with high-protein diet, we performed an oral glucose tolerance test. Rats fed with Western-type diet before and then kept on standard diet showed the largest increase of blood glucose following the orogastric gavage, which was blunted in the Western-type diet/high-protein diet group, indicating a beneficial effect. This improved glycemic control may be due to a stimulation of genes involved in gluconeogenesis, as reported in rats fed a high-protein diet (38). In the present study, the insulin sensitivity was not altered, while in a previous study using Wistar rats fed a high-fat diet (300 g fat/kg diet, 52% of calories from fat) before and switched to high whey protein diet, the insulin sensitivity was increased when calculated by the insulin-to-glucose ratio (3). Whether these differences are due to different measurements applied (insulin-to-glucose ratio vs. injection of insulin and measurement of the glucose response) or strain difference (Wistar vs. Sprague-Dawley rats) warrants further investigation.

The control of food intake and postprandial glycemia is tightly controlled by peptide hormones mainly derived from the gastrointestinal tract (23). Here, we investigated the physiological dark phase meal-stimulated release of different anorexigenic gut hormones among the four dietary conditions. In line with previous studies (44), most food intake-inhibitory hormones, namely amylin, GIP, insulin, PP, and PYY increased during the dark feeding phase to prevent further eating and overeating of the animals. However, leptin levels rather decreased after the dark-phase food intake. Previous studies did not detect alterations of leptin plasma levels between the light and dark photoperiod (2), and in a very recent study, leptin levels were not altered under conditions of fasting followed by 2 h of refeeding (62). These findings are in line with leptin being more involved in the long-term regulation of body weight (60). The increase of amylin was lower in the Western-type diet/high-protein diet group compared with the Western-type diet/standard diet group, which may point toward the restoration of amylin sensitivity reported to be reduced under conditions of chronic high-fat diet feeding (7). On the contrary, the increase of GIP (also known as glucose-dependent insulinotrophic peptide) was higher in the Western-type diet/high-protein diet group compared with the Western-type diet/standard diet group. Because both hormones are involved in glucose homeostasis (41, 51) and affect insulin release and signaling, these changes may contribute to the observed beneficial effect of a high-protein diet on glucose tolerance in diet-induced obese rats. In addition, diet-induced obese rats switched to a high-protein diet displayed a higher dark feeding phase-related fold-change increase of PP and PYY compared with the standard diet/standard diet group, which may contribute to the anorexigenic effect of a high-protein diet. These results cannot be related to differences in food intake as the 2-h dark-phase caloric consumption of standard or high-protein diet did not differ between the four groups. Vagal afferent fibers are the major neuroanatomical pathway linking the alimentary tract and the brain through which nutrients or gut peptides influence food intake (15, 43, 61). However, a previous study indicates that vagal afferents are not the main pathway signaling the feeding-suppressive effects of a high-protein diet in lean rats (32). This was supported by the fact that subdiaphragmatic vagotomy dampened the daily food intake response to a high-protein diet by only 14% compared with sham-operated rats (32). Whether the contribution of the vagus nerve is more prominent under conditions of DIO warrants investigation, especially noting the observed dark phase-related changes in gut hormones and the influence of feeding status on gut hormone receptors in the nodose ganglion (15).
Perspectives and Significance

We showed that a high-protein diet for 2 wk reduces body weight in Western-type diet-induced obese rats by selectively reducing fat, while sparing lean mass. Similar findings were observed in healthy lean rats fed a standard diet and switched to high-protein diet, although less pronounced. Concomitantly, the selective reduction of fat mass is accompanied by improved glucose tolerance in the diet-induced obese rats, which may be related to an altered meal-induced release of gastrointestinal peptides, in particular, the enhanced response of GIP. In view of the paucity of studies on the levels of postprandial circulating gut hormones in response to a high-protein diet in nonobese or diet-induced obese rats, additional investigations on their variations and contributions under these conditions will provide more insight into the role of hormonal mechanisms in the high-protein diet-induced curtailing of food intake. Recent evidence that the melanocortin pathway in the arcuate nucleus, a hypothalamic region with weaker blood-brain barrier and, therefore, sensitive to circulating nutrients and/or hormones (39), is activated by a 50% protein diet (19), supports such as possibility. Future studies will be needed to elucidate the full extent to which alterations in circulating gut hormones and signaling pathways within the arcuate nucleus contribute to the suppression of feeding in diet-induced obese rats fed a high-protein diet. Still, many questions remain unanswered regarding the peripheral and central mechanisms of a high-protein diet underlying its beneficial effect to improve body composition with loss of fat and retention of lean body mass in diet-induced obese rats (20). Taken together, these results support an important role for high-protein diet as an additional column in the multimodal treatment of obesity in addition to dietary coaching (regular eating schedule, slow eating, reduction of fat, and carbohydrate intake), exercise, psychotherapy, drug treatment, and bariatric surgery.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

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