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Fat Composition Changes in Bone Marrow During Chemotherapy and Radiation Therapy

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Summary

Magnetic resonance imaging fat quantification is sensitive to vertebral bone marrow composition changes that result from chemoradiation therapy. These findings indicate that locoregional effects of chemoradiation likely contribute to the clinical manifestation of myelosuppression.

Purpose: To quantify changes in bone marrow fat fraction and determine associations with peripheral blood cell counts.

Methods and Materials: In this prospective study, 19 patients received either highly myelotoxic treatment (radiation therapy plus cisplatin, 5-fluorouracil mitomycin C [FU/MMC], or cisplatin/5-FU/cetuximab) or less myelotoxic treatment (capecitabine-radiation therapy or no concurrent chemotherapy). Patients underwent MR imaging and venipuncture at baseline, midtreatment, and posttreatment visits. We performed mixed effects modeling of the mean proton density fat fraction (PDFF[%]) by linear time, treatment, and vertebral region (lumbar [L] 4-sacral [S]2 vs thoracic [T]10-L3 vs cervical(C)3-T9), while controlling for cumulative mean dose and other confounders. Spearman rank correlations were performed by white blood cell (WBC) counts versus the differences in PDFF(%) before and after treatment.

Results: Cumulative mean dose was associated with a 0.43% per Gy (P<.004) increase in PDFF(%). In the highly myelotoxic group, we observed significant changes in PDFF(%) per visit within L4-S2 (10.1%, P<.001) and within T10-L3 (3.93%, P=.01), relative to the reference C3-T9. In the less myelotoxic group, we did not observe significant changes in PDFF(%) per visit according to region. Within L4-S2, we observed a significant difference between treatment groups in the change in PDFF(%) per visit (5.36%, P=.04). Rank correlations of the inverse log differences in WBC versus the differences in PDFF(%) overall and within T10-S2 ranged from 0.69 to 0.78 (P<.05). Rank correlations of the inverse log differences in absolute...
neutrophil counts versus the differences in PDFF(%) overall and within L4-S2 ranged from 0.79 to 0.81 (P<.05).

**Conclusions**: Magnetic resonance imaging fat quantification is sensitive to marrow composition changes that result from chemoradiation therapy. These changes are associated with peripheral blood cell counts. This study supports a rationale for bone marrow-sparing treatment planning to reduce the risk of hematologic toxicity. Published by Elsevier Inc.

**Introduction**

A limiting factor in cancer treatment with chemoradiation therapy is marrow toxicity (1, 2). Bone marrow is composed of red and yellow marrow. Red marrow consists of hematopoietic stem cells that produce erythrocytes, leukocytes, and thrombocytes. Yellow marrow, like red marrow, contains abundant capillaries but is not directly involved in hematopoiesis. The stroma of the reticular network of yellow marrow is filled primarily with lipids, thus exhibiting a higher fat content. Red marrow is found in flat bones including the pelvis, sternum, and vertebrae, whereas yellow marrow is found in the medullary cavities of long bones. Both chemotherapy and radiation suppress the hematopoietic system, leading to a reduction in red marrow and an increase in yellow marrow (3). This composition change can result in neutropenia and thrombocytopenia that require chemotherapy dose reductions and delays, thus compromising treatment outcomes (4, 5).

T1-weighted magnetic resonance imaging (MRI) provides a qualitative impression of the amount of fat present in bone marrow due to the short T1 of fat compared to that in other tissues. Although this approach is sufficient for distinguishing low-fat from high-fat content, T1-weighting is not reliable when quantitative results and/or finer distinctions are required. A quantitative measure of bone marrow fat fraction is the iterative decomposition of water and fat with echo asymmetrical and least squares estimation (IDEAL) imaging technique, which can be used to create parametric fat fraction maps, providing both quantitative and spatially resolved information on marrow composition (6-10). Liang et al (11) showed that fat fraction maps have sufficient spatial resolution to be used in radiation therapy (RT) planning in patients undergoing pelvic chemoradiation.

Bolan et al (12) showed that water-fat MRI could be used to assess changes in bone marrow fat content in patients with gynecologic malignancies pre- and postchemotherapy and RT. These investigators showed that chemotherapy-induced changes are uniform in space and that radiation-induced changes are consistent with red-to-yellow marrow transformation. Although they showed an increase in marrow fat fraction at the lumbar (L)4 level from baseline to 6 months posttreatment they did not provide quantitative data for other vertebrae. Furthermore, they did not test differences in the magnitude, rate, and pattern of change between treatment groups or how fat fraction changes related to clinically significant variables such as the development of neutropenia.

The primary aim of this study was to assess the magnitude, rate, and pattern of change in vertebrae bone marrow fat fraction for patients receiving highly myelotoxic versus less myelotoxic pelvic chemoradiation therapy and versus RT alone. The study’s secondary aim was to determine associations between peripheral blood cell counts and fat fraction changes. Measuring composition changes over the course of treatment may provide quantitative spatial evidence to support bone marrow-sparing treatment planning approaches.

**Methods and Materials**

**Study design and patients**

This prospective, longitudinal, single-site clinical study was approved by the institutional review board and is compliant with the Health Insurance Portability and Accountability Act. The population consisted of patients undergoing pelvic chemoradiation therapy or RT alone for gynecologic, anorectal, or genitourinary malignancies. Subjects treated with palliative intent were excluded. Subjects were recruited from the institution’s radiation oncology clinics. All subjects provided informed consent. Our research budget afforded screening of 27 subjects, from November 2008 to July 2012, with 19 patients electing to participate (9 cervical cancer, 2 endometrial cancer, 3 anal cancer, 1 rectal cancer, and 4 prostate cancer patients).

**Treatment**

All patients received intensity modulated RT (IMRT). Four prostate cancer patients received pelvic RT only, 45.0 Gy in 1.8-Gy fractions to the pelvis, followed by a sequential boost to a total dose of 75.6 to 81.0 Gy. Two endometrial cancer patients underwent hysterectomy followed by adjuvant therapy with carboplatin (area under the curve = 6) and paclitaxel (175 mg/m², intravenously [IV]) for 6 cycles, followed by pelvic RT, 45.0 to 50.4 Gy in 1.8-Gy fractions. Anal cancer patients received 45 to 58.2 Gy in 1.5- to 2.0-Gy fractions with either concurrent 5-fluorouracil (5-FU; 1000 mg/m² IV, on days 1-4 and 29-32) and mitomycin-C (MMC; 10 mg/m² IV, on days 1 and 29; 2 patients) or cisplatin (75 mg/m² IV every 28 days × 2), 5-FU.
(1000 mg/m² IV on days 1-4 every 28 days × 2), and cetuximab (400 mg/m² loading dose and then 250 mg/m²/week, IV, every 6-8 weeks; 1 patient). Nine cervical cancer patients received pelvic RT, 45.0 to 50.4 Gy in 1.8-Gy fractions, with concurrent cisplatin, 40 mg/m² weekly, followed by high-dose rate brachytherapy (30 Gy in 5 fractions to point A). One rectal cancer patient received pelvic RT, 50.4 Gy in 1.8-Gy fractions, with concurrent capectabine therapy (825 mg/m², twice daily for 5 days per week). One of 3 anal cancer patients and 1 of 2 uterine cancer patients received pelvic-inguinal radiation. Two of 9 cervical cancer patients received extended field (ie, pelvic-para-aortic) RT. All patients completed chemoradiation therapy without treatment delays, and no patients received granulocyte colony-stimulating factors.

For this study, we broadly classified treatment into 2 main categories: highly myelotoxic treatment and less myelotoxic treatment (Table 1). The highly myelotoxic treatment group consisted of cervical cancer patients treated with cisplatin and anal cancer patients treated with 5-FU/MMC or cisplatin/5-FU/cetuximab. The less myelotoxic treatment group consisted of patients who received no concurrent chemotherapy or concurrent capecitabine.

### Concurrent imaging and laboratory blood tests

Patients underwent MRI with quantitative IDEAL (IDEAL-IQ) at baseline (within a 30-day window), midtreatment (within a 14-day window), and posttreatment (within 30 days of completion). MRI scans were performed with a 3.0-T scanner (Signal HDx; GE Healthcare, Milwaukee, WI). A spinal phased array coil was used. The IDEAL-IQ sequence is a multiple echo 3-dimensional (3D) spoiled gradient echo with a typical TR of 10 ms; a flip angle of 3°; and 6 echo times of between 1.0 and 6.0 ms. The technique estimates fat fraction and R2 with corrections included for B0 inhomogeneity, fat spectrum, and noise bias. Sagittal spinal scans were obtained with each study. White blood cell counts (WBC) and absolute neutrophil counts (ANC) were collected at baseline and weekly thereafter for patients undergoing chemoradiation therapy. Prostate cancer patients did not undergo serial blood count measurements. Peripheral blood counts obtained closest to the time of each MRI acquisition (within a 7-day window) were used for the analysis.

### Image analysis and dose calculation

A 2D region of interest (ROI) was drawn at the center of each subject’s vertebrae (cervical C3 to sacral S2) from serial sagittal spine MRIs, using the Digital Images and Communication in Medicine (DICOM) viewer (version 4.1; Osirix; Osirix Foundation, Geneva, Switzerland) (13). Each ROI contained approximately 100 to 200 voxels. The vertebral ROIs were positioned to include vertebral body bone marrow and to avoid areas of partial volume and

### Table 1  Patient, tumor and treatment characteristics

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Highly myelotoxic</th>
<th>Less myelotoxic</th>
<th>All</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of patients</td>
<td>12</td>
<td>7</td>
<td>19</td>
<td></td>
</tr>
<tr>
<td>Mean age (±SD), yrs</td>
<td>48 (±13)</td>
<td>62 (±0)</td>
<td>53 (±13)</td>
<td>.006</td>
</tr>
<tr>
<td>No. of women (%)</td>
<td>10 (83%)</td>
<td>3 (43%)</td>
<td>13</td>
<td>.17</td>
</tr>
<tr>
<td>Racial distribution, n (%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Caucasian</td>
<td>7 (58%)</td>
<td>6 (86%)</td>
<td>13</td>
<td>.34</td>
</tr>
<tr>
<td>Hispanic</td>
<td>4 (33%)</td>
<td>0 (0%)</td>
<td>4</td>
<td></td>
</tr>
<tr>
<td>Asian</td>
<td>1 (8%)</td>
<td>1 (14%)</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Mean body mass index (±SD), kg/m²</td>
<td>26.2 (±6)</td>
<td>26.3 (±6.1)</td>
<td>26.2 (±5.8)</td>
<td>.99</td>
</tr>
<tr>
<td>Cancer site: stage (n)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cervix: IBI (4), IB2 (2), IIB2 (2), IIIB (1)</td>
<td></td>
<td>Uterus: IIB (1), IV (1)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anus: IIIA (3)</td>
<td>CDDP (9)</td>
<td>Rectum: IIIA (1)</td>
<td>Prostate: III (4)</td>
<td></td>
</tr>
<tr>
<td>5-FU/MMC (2)</td>
<td>CDDP/5-FU/cetuximab (1)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Chemotherapy (n)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No. undergoing pretreatment chemotherapy (%)</td>
<td>0 (0%)</td>
<td>3 (43%)</td>
<td>3</td>
<td>.04</td>
</tr>
<tr>
<td>No. undergoing IMRT (%)</td>
<td>12 (100%)</td>
<td>7 (100%)</td>
<td>19</td>
<td>1.0</td>
</tr>
<tr>
<td>No. undergoing pelvic-aortic RT (%)</td>
<td>2 (17%)</td>
<td>0 (0%)</td>
<td>2</td>
<td>.51</td>
</tr>
<tr>
<td>No. undergoing pelvic-inguinal RT (%)</td>
<td>1 (8%)</td>
<td>1 (14%)</td>
<td>2</td>
<td>1.0</td>
</tr>
<tr>
<td>Mean WBC $\times 10^3/\mu$ (±SD)</td>
<td>7.25 (±2.5)</td>
<td>6.3 (±0.5)</td>
<td>16</td>
<td>.58</td>
</tr>
<tr>
<td>Mean ANC $\times 10^3/\mu$ (±SD)</td>
<td>4.6 (±1.9)</td>
<td>3.7 (±0.4)</td>
<td>14</td>
<td>.22</td>
</tr>
</tbody>
</table>

**Abbreviations:** 5-FU = 5-fluorouracil; ANC = absolute neutrophil count; CAP = capecitabine; CDDP = cisplatin; IMRT = intensity modulated radiation therapy; MMC = mitomycin C; RT = radiation therapy; SD = standard deviation; WBC = white blood cell count.
image artifact. Each ROI was inspected and manually adjusted if necessary to account for subject motion between scans. The mean signal intensity within the ROI provided corresponding vertebral marrow proton density fat fraction (PDFF) values. PDFF in a given sample is defined as the number of protons in fat (triglycerides) divided by the total number of protons (water plus fat) expressed as percentages (PDFF[%]) from 0 to 100 (6). ROIs were grouped into 3 vertebral column regions: lumbar (L)4 to S2 (the area consistently receiving large doses within the pelvic field), thoracic (T)10 to L3 (the area superior to the pelvic field receiving variable doses), and C3 to T9 (area receiving no dose). Grand mean PDFF(%) values were calculated for each subject according to region. From the radiation treatment plans, cumulative mean doses (Gy) were calculated for each subject according to region at mid- and posttreatment visits.

Statistical analysis

Student’s t, Mann-Whitney U, and Fisher’s exact tests were used to analyze baseline differences in age, race, sex, body mass index (BMI), WBC, and ANC between the highly myelotoxic and less myelotoxic treatment groups. To estimate the effects of myelotoxic chemotherapy on bone marrow fat fraction, we performed Spearman rank correlations for the differences in mean proton density fat fraction (ΔPDFF(%)]) between the end of treatment (t2) and baseline (t0) versus −log(WBCt2/WBCt0), −log(ANCt2/ANCt0), and WBC and ANC nadirs (lowest values occurring during RT). Rank correlation statistics were computed overall and according to vertebral column region.

All tests were two-tailed, with significant correlations at a P value of ≤.05. Data were prepared and analyzed using R version 2.15.1 software (http://www.r-project.org).

Results

Twelve patients received highly myelotoxic treatment and 7 patients received less myelotoxic treatment (Table 1). The median age was 55 years of age (range: 27-81 years). Patients were younger in the highly myelotoxic treatment group. There were no differences in race, BMI, WBC, and ANC between treatment groups.

Fifteen of 19 patients completed all 3 IDEAL-IQ scans (baseline, midtreatment, and posttreatment Two patients completed only the baseline and midtreatment scans, 1 patient completed only the baseline and posttreatment scans, and 1 patient completed only the midtreatment and posttreatment scans. Altogether, 18, 18, and 17 patients completed the baseline, midtreatment, and posttreatment scans, respectively. The mean (±SD) number of days from the baseline IDEAL-IQ scan to the midtreatment and posttreatment scans were 24 (±10) and 55 (±13) days, respectively.

Twelve patients completed all 3 WBC draws (baseline, midtreatment, and posttreatment). Two patients completed only the baseline and midtreatment WBC draws, 1 patient completed only the baseline and posttreatment WBC draws, and 1 patient completed only the baseline WBC draw. Altogether, 16, 14, and 13 patients completed the baseline, midtreatment, and posttreatment WBC measurements, respectively. The mean (±SD) number of days for the WBC measurements relative to the baseline, midtreatment, and posttreatment MRI scans were 2 (±5), −1 (±3), and 1 (±6) days, respectively.

Nine patients completed all 3 ANC draws (baseline, midtreatment, and posttreatment). Two patients completed only the baseline and midtreatment ANC draws, 1 patient completed only the baseline and posttreatment ANC draws, 1 patient completed only the midtreatment and posttreatment ANC draws, and 1 patient completed only the baseline ANC test. Altogether, 14, 12, and 11 patients completed the baseline, midtreatment, and posttreatment WBC measurements, respectively. The mean (±SD) number of days for the WBC measurements relative to the baseline, midtreatment, and posttreatment MRI scans were 1 (±5), −1 (±3), and 1 (±6) days, respectively.

Mean PDFF(%) values and cumulative mean doses are shown in Table 2. At baseline, mean PDFF(%) was significantly lower in the highly myelotoxic treatment group than in the less myelotoxic treatment group (37.5% vs 53.6%, respectively, P<.001), likely due to the fact that some patients in the low myelotoxic group received pretreatment chemotherapy. Figure 1 shows a representative image of a subject who received highly myelotoxic chemoradiation therapy (Fig. 1 A-C) compared to a subject who received RT alone (Fig. 1 D-F). Images were produced at baseline, at midtreatment, and upon completion of
treatment. Qualitatively, patients in Figure 1 A–C appear to have higher increases in fat fraction during treatment than those in Figure 1 D–F. To help readers differentiate among signal intensities, we show PDFF(%) values for L5 ROI’s: for the subject receiving highly myelotoxic treatment, L5 progressively increases during treatment (59.4%-78.5%-92.1%, respectively), and for the subject receiving only RT, L5 slightly increases but then decreases at the end treatment (59.5%-65.8%-63.8%, respectively).

Figure 2 shows mean differences in PDFF(%) for regions T10-L3 (Fig. 2A) and L4-S2 (Fig. 2B) relative to the reference (C3-T9) by group and visit. At baseline, the relative differences in PDFF(%) for both regions are higher for the less myelotoxic group. However, over the course of treatment, we observed greater differences in PDFF(%) within both regions in the highly myelotoxic group compared to the less myelotoxic group.

Table 3 shows adjusted mean PDFF(%) estimates by MRI study visit, treatment group, vertebral column region, and confounders, with less myelotoxic treatment than the reference group. In the less myelotoxic group, we observed significant increases at baseline in mean PDFF(%) within T10-L3 and L4-S2 relative to that in C3-T9 (intercept). When we performed the analysis with highly myelotoxic treatment as the reference group (as opposed to less myelotoxic treatment shown in Table 3), we also observed

<table>
<thead>
<tr>
<th>Treatment group</th>
<th>Region</th>
<th>Baseline PDFF(%)</th>
<th>Cumulative dose (Gy)</th>
<th>Midtreatment PDFF(%)</th>
<th>Cumulative dose (Gy)</th>
<th>Posttreatment PDFF(%)</th>
<th>Cumulative dose (Gy)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Highly myelotoxic</td>
<td>C3-T9</td>
<td>33.0 (11.1)</td>
<td>0</td>
<td>34.9 (11.5)</td>
<td>0</td>
<td>30.5 (15.5)</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>T10-L3</td>
<td>36.0 (13.9)</td>
<td>0</td>
<td>41.2 (14.1)</td>
<td>0.42 (0.78)</td>
<td>42.0 (18.7)</td>
<td>0.60 (1.07)</td>
</tr>
<tr>
<td></td>
<td>L4-S2</td>
<td>43.2 (16.5)</td>
<td>0</td>
<td>64.3 (15.5)</td>
<td>19.3 (5.85)</td>
<td>74.1 (15.5)</td>
<td>28.3 (5.37)</td>
</tr>
<tr>
<td>All vertebrae (C3-S2)</td>
<td></td>
<td>37.5 (14.4)</td>
<td>0</td>
<td>46.5 (18.3)</td>
<td>6.77 (9.79)</td>
<td>49.7 (23.7)</td>
<td>9.93 (13.9)</td>
</tr>
<tr>
<td>Less myelotoxic</td>
<td>C3-T9</td>
<td>47.2 (11.5)</td>
<td>0</td>
<td>48.0 (11.0)</td>
<td>0</td>
<td>41.4 (11.3)</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>T10-L3</td>
<td>52.6 (12.8)</td>
<td>0</td>
<td>51.8 (13.4)</td>
<td>0.17 (0.11)</td>
<td>47.9 (11.7)</td>
<td>0.28 (0.09)</td>
</tr>
<tr>
<td></td>
<td>L4-S2</td>
<td>61.6 (11.2)</td>
<td>0</td>
<td>70.3 (14.2)</td>
<td>18.9 (7.68)</td>
<td>77.8 (12.7)</td>
<td>28.0 (3.86)</td>
</tr>
<tr>
<td>All vertebrae (C3-S2)</td>
<td></td>
<td>53.6 (13.1)</td>
<td>0</td>
<td>57.1 (15.3)</td>
<td>6.67 (10.2)</td>
<td>55.8 (18.9)</td>
<td>9.41 (13.7)</td>
</tr>
</tbody>
</table>

Abbreviations: C = cervical vertebrae; L = lumbar vertebrae; PDFF(%) = proton density fat fraction; S = sacral vertebrae; T = thoracic vertebrae.

Fig. 1. (A–C) MRIs from a subject with cervical cancer who received cisplatin chemotherapy and IMRT. (D–F) MRIs from a subject with prostate cancer who received IMRT alone. (A and D) Baseline results; (B and E) midtreatment; (C and F) immediately after treatment. PDFF(%) values for L5 regions of interest are (A) 59.4%, (B) 78.5%, (C) 92.1%, (D) 59.5%, (E) 65.8%, and (F) 63.8%. IMRT = intensity modulated radiation therapy; MRI = magnetic resonance imaging; PDFF = proton density fat fraction.
significant increases at baseline in mean PDFF(%) within T10-L3 (4.48%, 95% confidence interval [CI]: 0.63%-8.32%, \(P = .02\)) and L4-S2 (11.6%, 95% CI: 7.73%-15.5%, \(P < .001\)) relative to that in C3-T9 (34.5%, 95% CI: 2.78%-1.2%, \(P < .001\)). The adjusted model with less myelotoxic treatment as the reference (Table 2) showed no significant differences between treatment groups for each region at baseline. Similarly, we observed no differences when we excluded patients who received pretreatment chemotherapy. Pretreatment chemotherapy was associated with an increase in baseline PDFF(%) of 12.8% (95% CI: 2.70% to 28.2%), and age was associated with an increase in PDFF(%) of 0.39% per year (95% CI: 5.6e-4% to 0.79%). We observed a significant effect of cumulative mean dose on the mean PDFF(%) (0.43% per Gy, 95% CI: 0.13%-0.72%). In the less myelotoxic group, we observed no significant differences among the changes in mean PDFF(%) per visit according to region. With highly myelotoxic treatment as the reference group, we observed significant differences in the changes in mean PDFF(%) per visit according to T10-L3 (3.93% per visit, 95% CI: 0.90-6.96, \(P = .01\)) and L4-S2 (10.1% per visit, 95% CI: 5.01-15.2, \(P < .001\)) relative to that of C3-T9. These effects were still present when we excluded patients who received extended field RT (T10-L3: 3.44% per visit, 95% CI: 0.12%-6.77%, \(P = .04\); L4-S2: 9.06% per visit, 95% CI: 2.50%-15.6%, \(P = .006\)). For the highly myelotoxic group relative to the less myelotoxic group, we observed significant differences in changes in PDFF(%) per visit within L4-S2 (5.36% per visit, 95% CI: 0.29%-10.4%).

![Bar plots](image)

**Fig. 2.** Bar plots of the mean difference (95% CI) in proton density fat fraction (PDFF[%]) for (A) T10-L3 and (B) L4-S2 relative to C3-T9 by treatment group and visit. PDFF(%) values are labeled within the bars. PDFF = proton density fat fraction.

---

**Table 3** Adjusted mean PDFF(%) estimates and 95% CI by MRI study visit, treatment group, vertebral column region and confounders

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Mean PDFF(%)</th>
<th>(P)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline estimates for the less myelotoxic group</td>
<td>C3-T9 (intercept)</td>
<td>37.3 (25.7-49.0)</td>
</tr>
<tr>
<td></td>
<td>T10-L3</td>
<td>+5.94 (0.71-11.2)</td>
</tr>
<tr>
<td></td>
<td>L4-S2</td>
<td>+13.9 (8.68-19.2)</td>
</tr>
<tr>
<td>Difference in estimates for the highly myelotoxic group relative to the less myelotoxic group at baseline</td>
<td>C3-T9</td>
<td>-2.83 (18.2 to 12.5)</td>
</tr>
<tr>
<td></td>
<td>T10-L3</td>
<td>-1.45 (-7.95 to 5.03)</td>
</tr>
<tr>
<td></td>
<td>L4-S2</td>
<td>-2.33 (-8.82 to 4.16)</td>
</tr>
<tr>
<td>Change in PDFF(%) per visit for the less myelotoxic group</td>
<td>C3-T9</td>
<td>+1.84 (6.03 to 2.34)</td>
</tr>
<tr>
<td></td>
<td>T10-L3</td>
<td>+0.37 (-3.69 to 4.44)</td>
</tr>
<tr>
<td></td>
<td>L4-S2</td>
<td>+4.76 (-1.00 to 10.5)</td>
</tr>
<tr>
<td>Difference in the change in PDFF(%) per visit between the highly myelotoxic and less myelotoxic groups</td>
<td>C3-T9</td>
<td>+1.50 (-3.74 to 6.73)</td>
</tr>
<tr>
<td></td>
<td>T10-L3</td>
<td>+3.56 (-1.51 to 8.63)</td>
</tr>
<tr>
<td></td>
<td>L4-S2</td>
<td>+5.36 (0.29-10.4)</td>
</tr>
<tr>
<td>Cumulative dose (per Gy)</td>
<td>Pretreatment chemotherapy</td>
<td>+0.43 (0.13-0.72)</td>
</tr>
<tr>
<td>Age (per yr)</td>
<td>+12.8 (2.70 to 28.2)</td>
<td>.08</td>
</tr>
<tr>
<td></td>
<td>+0.39 (5.6e-4 to 0.79)</td>
<td>.05</td>
</tr>
</tbody>
</table>

*Abbreviations:* C = cervical vertebra; L = lumbar vertebra; PDFF(%) = proton density fat fraction (%); S = sacral vertebrae; T = thoracic vertebra.

* \(P\) value is testing differences in mean PDFF(%) for the highly myelotoxic group relative to that of the less myelotoxic group at baseline by vertebral column region.

† \(P\) value is testing differences in the change in mean PDFF(%) per visit for the highly myelotoxic group relative to that of the less myelotoxic group by vertebral column region.
did not observe significant differences between treatment groups for the C3-T9 or T10-L3 region. Of note, when we did not control for region, we also observed a significant difference in the change in PDFF(%) per visit within C3-S2 (4.58% per visit, 95% CI: 0.52%-8.64%, $P = .03$).

Figure 3 shows scatterplots and Spearman rank correlations for the difference in PDFF(%) versus peripheral blood cell counts by (A) C3-S2, (B) C3-T9, (C) T10-L3, and (D) L4-S2. PDFF = proton density fat fraction.

L4-S2 ($r = 0.81, P = .008$) and C3-S2 ($r = 0.79, P = .01$) and was borderline for T10-L3 ($r = 0.62, P = .06$). No significant correlations between ΔPDFF(%) and $-\log(\text{WBC}_{t2}/\text{WBC}_{t0})$ or $-\log(\text{ANC}_{t2}/\text{ANC}_{t0})$ were observed within C3-T9.

We also observed significant negative correlations between ΔPDFF(%) and WBC nadirs within C3-S2 ($r = -0.57, P = .03$) and T10-L3 ($r = -0.69, P = .006$), and a borderline significant correlation for L4-S2 ($r = -0.49, P = .07$), indicating an association between increased fat fraction and lower blood count nadirs. Similarly, we observed borderline significant negative correlations between ΔPDFF(%) and ANC nadirs within C3-S2 ($r = -0.50, P = .10$), T10-L3 ($r = -0.52, P = .08$), and L4-S2 ($r = -0.51, P = .09$). No significant correlations
between ΔPDFF(%) and WBC and ANC nadirs were observed within C3-T9.

Discussion

Multiple studies indicate that increased radiation dose to pelvic bone marrow is associated with greater hematologic toxicity in patients undergoing chemoradiation therapy (11, 17-25). Modern RT techniques can allow the incorporation of functional bone marrow imaging to specifically avoid hematopoietically active marrow subregions, which may reduce hematologic toxicity and increase chemotherapy tolerance (18, 24, 26).

In this study, we used IDEAL-IQ to compare changes in vertebral bone marrow fat fraction in patients undergoing highly myelotoxic chemoradiation therapy versus less myelotoxic chemoradiation therapy or RT alone. Overall, we observed that cumulative mean dose leads to significant increases in PDFF(%) within the vertebral bone marrow. These changes in PDFF(%) also corresponded to decreasing peripheral blood cell counts. In the highly myelotoxic group, we observed significant increases in PDFF(%) over the course of treatment within (eg, L4-S2) and above the pelvic radiation field (eg, T10-L3), even in patients who did not undergo extended field RT. Increases in PDFF(%) within the pelvic radiation field were significantly more pronounced in the highly myelotoxic group relative to that in the less myelotoxic group. Altogether these findings suggest that IDEAL is sensitive to marrow composition changes resulting from chemotherapy and radiation and that locoregional radiation effects contribute to the manifestation of clinical myelosuppression, particularly in patients undergoing myelotoxic chemotherapy.

Although conventional MR imaging may broadly distinguish red from yellow bone marrow (27), quantifying subtle gradations within red marrow in response to radiation therapy is possible only by using recent advances in MR fat quantification methodology. The use of MR with fat quantification allows noninvasive monitoring of the spatial effects of RT and to quantify the degree of red marrow damage as a function of radiation dose. This allows us to model and estimate the impact of conformal radiation techniques on red bone marrow injury.

This study had several limitations. There is significant heterogeneity in the population. To a degree, this heterogeneity was useful to estimate the impact of varying conditions on changes in marrow fat composition; however, further studies are ongoing in more homogeneous populations to control effects of treatment and demographic factors that could influence our findings (32). This longitudinal study included a relatively small sample and was subject to dropout, reducing our statistical power. Although we do not suspect this would have biased our primary effect estimates, this may have limited our ability to detect significant differences in the effects of chemotherapy in regions superior to the pelvic field. Our model form does assume a linear rate of change in fat fraction according to treatment group and spatial location, which may or may not be valid. However, the linear time model has the advantage of simplicity and parsimony and appeared to model the data well.

Conclusions

In summary, we observed increases in bone marrow fat composition over the course of treatment occurring specifically within the radiation treatment field, particularly in the presence of concurrent myelotoxic chemotherapy. These findings indicate that locoregional effects of RT likely significantly contribute to the clinical manifestation of myelosuppression in this setting. Modern RT techniques specifically designed to spare functioning bone marrow may reduce myelosuppression and permit better tolerance to myelotoxic chemotherapy regimens.

References

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